Package 'gespeR'

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Imports Matrix, glmnet, cellHTS2, Biobase, biomaRt, doParallel, parallel, foreach, reshape2, dplyr

Depends methods, graphics, ggplot2, R(>= 2.10)

Suggests knitr

biocViews ImmunoOncology, CellBasedAssays, Preprocessing, GeneTarget, Regression, Visualization

VignetteBuilder knitr

Type Package

Lazyload yes

Title Gene-Specific Phenotype EstimatoR

Version 1.24.0

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Description Estimates gene-specific phenotypes from off-target confounded RNAi screens. The phenotype of each siRNA is modeled based on on-targeted and off-targeted genes, using a regularized linear regression model.

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URL http://www.cbg.ethz.ch/software/gespeR

Collate 'Phenotypes-class.R' 'TargetRelations-class.R' 'gespeR-class.R' 'gespeR-concordance.R' 'gespeR-functions.R' 'gespeR-generics.R' 'gespeR-methods.R' 'gespeR-package.R'

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`gespeR-package` *Package: Gene-Specific Phenotype EstimatoR*

Description

This package provides a model to deconvolute off-target confounded RNAi knockdown phentypes, and methods to investigate concordance between ranked lists of (estimated) phenotypes. The regularized linear regression model can be fitted using two different strategies. (a) Cross-validation over regularization parameters optimising the mean-squared-error of the model and (b) stability selection of covariates (genes) based on a method by Nicolai Meinshausen et al.

Author(s)

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References

Fabian Schmich et. al, Deconvoluting Off-Target Confounded RNA Interference Screens (2014).

See Also

gespeR

Examples

```
# Read phenotypes
phenos <- lapply(LETTERS[1:4], function(x) {
  sprintf("Phenotypes_screen_%s.txt", x)
  })
phenos <- lapply(phenos, function(x) {
  Phenotypes(system.file("extdata", x, package="gespeR"),
             type = "SSP",
             col.id = 1,
             col.score = 2)
})
phenos
plot(phenos[[1]])

# Read target relations
tr <- lapply(LETTERS[1:4], function(x) {
  sprintf("TR_screen_%s.rds", x)
})
tr <- lapply(tr, function(x) {
  TargetRelations(system.file("extdata", x, package="gespeR"))
})
tr[[1]]
tempfile <- paste(tempfile(pattern = "file", tmpdir = tempdir()), ".rds", sep="")
tr[[1]] <- unloadValues(tr[[1]], writeValues = TRUE, path = tempfile)
tr[[1]]
tr[[1]] <- loadValues(tr[[1]])
tr[[1]]

# Fit gespeR models with cross validation
res.cv <- lapply(1:length(phenos), function(i) {
  gespeR(phenotypes = phenos[[i]],
         target.relations = tr[[i]],
         mode = "cv",
         alpha = 0.5,
         ncores = 1)
})
summary(res.cv[[1]])
res.cv[[1]]
plot(res.cv[[1]])

# Extract scores
ssp(res.cv[[1]])
gsp(res.cv[[1]])
head(scores(res.cv[[1]]))
```

```
# Fit gespeR models with stability selection
res.stab <- lapply(1:length(phenos), function(i) {
  gespeR(phenotypes = phenos[[i]],
         target.relations = tr[[i]],
         mode = "stability",
         nbootstrap = 100,
         fraction = 0.67,
         threshold = 0.75,
         EV = 1,
         weakness = 0.8,
         ncores = 1)
})
summary(res.stab[[1]])
res.stab[[1]]
plot(res.stab[[1]])

# Extract scores
ssp(res.stab[[1]])
gsp(res.stab[[1]])
head(scores(res.stab[[1]]))

# Compare concordance between stability selected GSPs and SSPs
conc.gsp <- concordance(lapply(res.stab, gsp))
conc.ssp <- concordance(lapply(res.stab, ssp))

pl.gsp <- plot(conc.gsp) + ggtitle("GSPs\n")
pl.ssp <- plot(conc.ssp) + ggtitle("SSPs\n")

if (require(grid)) {
  grid.newpage()
  pushViewport(viewport(layout = grid.layout(1, 2) ) )
  print(pl.gsp, vp = viewport(layout.pos.row = 1, layout.pos.col = 1))
  print(pl.ssp, vp = viewport(layout.pos.row = 1, layout.pos.col = 2))
} else {
  plot(pl.gsp)
  plot(pl.ssp)
}
```

`annotate.gsp` *annotate.gsp*

Description

Query Biomart HGNC symbols for the entrez identifiers of estimated GSPs. Currently, only implemented for species "hsapiens".

Usage

```
## S4 method for signature 'Phenotypes'
```

```
annotate.gsp(object, organism = "hsapiens")

## S4 method for signature 'gespeR'
annotate.gsp(object, organism = "hsapiens")
```

Arguments

object	A gespeR or Phenotypes object
organism	String indicating the biomaRt organism

Value

data.frame containing gene identifier, gene symbol and phenotypic score

Author(s)

Fabian Schmich

See Also

gsp

ssp

scores

Examples

```
data(stabilityfits)
 gspA <- gsp(stabilityfits$A)
## Not run:
 annotate.gsp(gspA)

## End(Not run)
```

---

as.data.frame,Phenotypes-method

Convert Phenotypes object to a data.frame

Description

Convert Phenotypes object to a data.frame

Usage

```
## S4 method for signature 'Phenotypes'
as.data.frame(x)
```

Arguments

x	A Phenotypes object

Value

A data.frame

Author(s)

Fabian Schmich

Examples

```
phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
type = "SSP",
col.id = 1,
col.score = 2)
as.data.frame(phenos)
```

---

as.data.frame.concordance

Coerce method

Description

Coerce method

Usage

```
## S3 method for class 'concordance'
as.data.frame(x, ...)
```

Arguments

x	concordance object
...	additional arguments

Value

data.frame

Author(s)

Fabian Schmich

c,Phenotypes-method *Concatenate Phenotypes objects*

Description

Concatenate Phenotypes objects

Usage

```
## S4 method for signature 'Phenotypes'
c(x, ..., recursive = FALSE)
```

Arguments

x	A Phenotypes object
...	additional Phenotypes objects
recursive	recursive

Value

A concatenated Phenotypes object

Author(s)

Fabian Schmich

Examples

```
phenos.a <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
type = "SSP",
col.id = 1,
col.score = 2)
phenos.b <- Phenotypes(system.file("extdata", "Phenotypes_screen_B.txt", package = "gespeR"),
type = "SSP",
col.id = 1,
col.score = 2)
c(phenos.a, phenos.b)
```

concordance *Evaluate the concordance between Phenotype objects*

Description

Measures include the correlation (rho) between pairs of phenotypes for the same gene, the rank biased overlap (rbo) of the top and bottom of ranked lists, and the Jaccard index (J) of selected genes.

Usage

```
concordance(..., min.overlap = 10, cor.method = "spearman", rbo.p = 0.98,
  rbo.k = NULL, rbo.mid = 0, uneven.lengths = TRUE)
```

Arguments

...	The phenotypes to be evaluated for concordance
min.overlap	The minimum number of overlapping genes required
cor.method	A character string indicating which correlation coefficient is to be computed
rbo.p	The weighting parameter for rank biased overlap (rbo) in [0, 1]. High p implies strong emphasis on top ranked elements
rbo.k	The evaluation depth for rank biased overlap extrapolation
rbo.mid	The mid point to split a ranked list, e.g. in order to split positive and negative scores choose mid=0
uneven.lengths	Indicator if lists have uneven lengths

Value

A concordance object with the following elements:

pair.test	Indicator of compared phenotypes
cor	The correlation between pairs of phenotypes for the same gene
rbo.top	The rank biased overlap of genes evaluated at the top of the ranked list
rbo.bottom	The rank biased overlap of genes evaluated at the bottom of the ranked list
jaccard	The Jaccard index of selected genes

Author(s)

Fabian Schmich

See Also

Phenotypes

plot.concordance

rbo

Examples

```
data(stabilityfits)
conc <- concordance(gsp(stabilityfits$A), gsp(stabilityfits$B),
gsp(stabilityfits$C), gsp(stabilityfits$D))
plot(conc)
```

---

dim,Phenotypes-method *Dimension of a* Phenotypes *object*

Description

Dimension of a Phenotypes object

Usage

```
## S4 method for signature 'Phenotypes'
dim(x)
```

Arguments

x	Phenotypes object

Value

Dimension of the Phenotypes object

Author(s)

Fabian Schmich

---

gespeR-class *gespeR*

Description

Class that represents a gespeR model. It contains a SSP Phenotypes and TargetRelations representing a siRNA knockdown experiment. When the model is fitted, it additionaly contains estimated GSP Phenotypes.

Usage

```
gespeR(phenotypes, target.relations, ...)

## S4 method for signature 'Phenotypes,TargetRelations'
gespeR(phenotypes, target.relations,
  mode = c("cv", "stability"), alpha = 0.5, nbootstrap = 100,
  fraction = 0.67, threshold = 0.9, EV = 1, weakness = 0.8,
  ncores = 1, ...)

## S4 method for signature 'numeric,Matrix'
gespeR(phenotypes, target.relations, ...)
```

Arguments

phenotypes	The siRNA-spefic phenotypes. Single object for univariate phenotypes and list of Phenotypes objects for multivariate phenotypes.
target.relations	The siRNA-to-gene target relations
...	Additional arguments
mode	The mode of covariate selectino ("cv" or "stability")
alpha	The glmnet mixing parameter
nbootstrap	The number of bootstrap samples
fraction	The fraction for each bootstrap sample
threshold	The selection threshold
EV	The expected value of wrongly selected elements
weakness	The weakness parameter for randomised lasso
ncores	The number of cores for parallel computation

Value

A gespeR object

Slots

SSP The observed siRNA-specific phenotypes

GSP The deconvoluted gene-specific phenotypes

target.relations The siRNA-to-gene target relations, e.g. predicted by TargetScan

is.fitted An indicator wheter the gespeR model was fitted

model The fitted regularized linear regression model

Author(s)

Fabian Schmich

See Also

gespeR-package

plot.gespeR

gsp

ssp

scores

stability

target.relations

Examples

```
phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
type = "SSP",
col.id = 1,
col.score = 2)
trels <- TargetRelations(readRDS(system.file("extdata", "TR_screen_A.rds", package = "gespeR")))
res <- gespeR(phenotypes = phenos,
    target.relations = trels,
    mode = "stability",
    nbootstrap = 100,
    fraction = 0.67,
    threshold = 0.75,
    EV = 1,
    weakness = 0.8,
    ncores = 1)
gsp(res)
```

---

gsp *Retrieve GSPs and SSPs from* gespeR *objects*

Description

Retrieve GSPs and SSPs from gespeR objects

Usage

```
gsp(object)

## S4 method for signature 'gespeR'
gsp(object)

ssp(object)

## S4 method for signature 'gespeR'
ssp(object)
```

Arguments

object	A gespeR object

Value

A Phenotypes object of GSPs and SSPs, respectively

Author(s)

Fabian Schmich

See Also

annotate.gsp

scores

Examples

```
data(stabilityfits)
gsp(stabilityfits$A)
ssp(stabilityfits$B)
```

---

join *join*

Description

Join a TargetRelations object and a Phenotype object

Usage

```
join(targets, phenotypes)

## S4 method for signature 'TargetRelations,Phenotypes'
join(targets, phenotypes)
```

Arguments

targets	A TargetRelations object.
phenotypes	A Phenotypes object.

Value

List containing the matched targets and phenotypes

Author(s)

Fabian Schmich

Examples

```
phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
type = "SSP",
col.id = 1,
col.score = 2)
trels <- TargetRelations(readRDS(system.file("extdata", "TR_screen_A.rds", package = "gespeR")))
phenos <- phenos[1:17]
stripped_down <- join(targets = trels, phenotypes = phenos)
```

`lasso.rand` *Randomized Lasso*

Description

Based on Meinshausen and Buehlmann (2009)

Usage

```
lasso.rand(x, y, weakness = 1, subsample = 1:nrow(x), dfmax = (ncol(x) +
  1), lambda = NULL, standardize = FALSE, intercept = FALSE, ...)
```

Arguments

`x`	The design matrix
`y`	The response vector
`weakness`	The weakness parameter
`subsample`	The data subsample (default: none)
`dfmax`	The maxiumum number of degrees of freedom
`lambda`	The regularisation parameter
`standardize`	Indicator, wheter to standardize the design matrix
`intercept`	Indicator, whether to fit an intercept
`...`	Additional arguments to `glmnet`

Value

A `glmnet` object

Author(s)

Fabian Schmich

Examples

```
y <- rnorm(50)
 x <- matrix(runif(50 * 20), ncol = 20)
 lasso.rand(x = x, y = y)
```

loadValues *Methods for values of* TargetRelations *objects*

Description

Load, unload or write to file the values of a TargetRelations object

Usage

```
loadValues(object)

## S4 method for signature 'TargetRelations'
loadValues(object)

## S4 method for signature 'gespeR'
loadValues(object)

unloadValues(object, ...)

## S4 method for signature 'TargetRelations'
unloadValues(object, writeValues = TRUE,
  overwrite = FALSE, path = NULL)

## S4 method for signature 'gespeR'
unloadValues(object, writeValues = TRUE,
  overwrite = FALSE, path = NULL)

writeValues(object, ...)

## S4 method for signature 'TargetRelations'
writeValues(object, overwrite = FALSE)
```

Arguments

object	A TargetRelations object or gespeR object
...	Additional arguments
writeValues	Indicator, whether to write values
overwrite	Indicator, wheter to overwrite values if file exists at path
path	The path to write out values

Value

A TargetRelations object or gespeR object

Author(s)

Fabian Schmich

Examples

```
data(stabilityfits)
## Not run:
loadValues(stabilityfits$A)

## End(Not run)
```

`na.rem` *Remove NA/Inf values from phenotype vectors*

Description

Remove NA/Inf values from phenotype vectors

Usage

```
na.rem(object)

## S4 method for signature 'Phenotypes'
na.rem(object)
```

Arguments

`object`	A `Phenotypes` object

Value

A `Phenotypes` object without NA scores values

Author(s)

Fabian Schmich

Examples

```
phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
type = "SSP",
col.id = 1,
col.score = 2)
na.rem(phenos)
```

path<- *path*

Description

Set the path of a a TargetRelations object object

Usage

```
path(object) <- value

## S4 replacement method for signature 'TargetRelations,character'
path(object) <- value
```

Arguments

object	A TargetRelations object
value	A string defining the path

Value

A TargetRelations object with set path

Author(s)

Fabian Schmich

Examples

```
trels <- TargetRelations(readRDS(system.file("extdata", "TR_screen_A.rds", package = "gespeR")))
path(trels) <- "/dev/null"
```

Phenotypes-class *Phenotypes*

Description

Class used to represent various types of phenotypes, e.g. from siRNA-specific (SSP) or estimated gene-specific phenotypes (GSP).

Usage

```
Phenotypes(phenotypes, ...)

## S4 method for signature 'character'
Phenotypes(phenotypes, type = c("SSP", "GSP"),
  sep = "\t", col.id = 1, col.score = 2)

## S4 method for signature 'cellHTS'
Phenotypes(phenotypes, channel, sample)

## S4 method for signature 'Matrix'
Phenotypes(phenotypes, ids = NULL, pnames = NULL,
  type = c("SSP", "GSP"))
```

Arguments

phenotypes	The phenotypes as numeric vector, path to a .txt file with two columns (1: identifiers, 2: values), or a cellHTS object
...	Additional arguments
type	The type of phenotype (GSP, SSP)
sep	The separator string
col.id	Column number for the siRNA identifiers
col.score	Column number(s) for the phenotype score
channel	The cellHTS channel identifier
sample	The cellHTS sample index
ids	The siRNA/gene identifiers
pnames	The phenotype identifiers

Value

A Phenotypes object

Slots

type The type of represented phenotypes (i.e., "SSP" or "GSP")

ids The entity identifiers (i.e., siRNA or gene ids)

pnames The phenotype names

values The phenotypic values

Author(s)

Fabian Schmich

See Also

plot.Phenotypes

join

gsp

ssp

scores

concordance

Examples

```
phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
type = "SSP",
col.id = 1,
col.score = 2)
```

---

plot.concordance *Plot concordance*

Description

Plots boxplots of concordance evaluated between multiple Phenotype objects. Measures include the correlation (rho) between pairs of phenotypes for the same gene, the rank biased overlap (rbo) of the top and bottom of ranked lists, and the Jaccard index (J) of selected genes.

Usage

```
## S3 method for class 'concordance'
plot(x, ...)
```

Arguments

x	The data of class concordance
...	Additional parameters for plot

Value

Boxplots of concordance measures

Author(s)

Fabian Schmich

plot.gespeR *Plot method for gespeR objects*

Description

Plot method for gespeR objects

Usage

```
## S3 method for class 'gespeR'
plot(x, ...)
```

Arguments

x	A gespeR object
...	Additional paramters for plot

Value

Histogram of SSPs or GSPs

Author(s)

Fabian Schmich

plot.Phenotypes *Plot method for Phenotype objects*

Description

Plot method for Phenotype objects

Usage

```
## S3 method for class 'Phenotypes'
plot(x, ...)
```

Arguments

x	A Phenotypes object
...	Additional arguments for plot

Value

Histogram of scores phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"), type = "SSP", col.id = 1, col.score = 2) plot(phenos)

Author(s)

Fabian Schmich

---

rbo *Rank biased overlap (Webber et al., 2010)*

Description

Evaluates the rank biased overlap (rbo) of two ranked lists based on formula based on (32) from "A Similarity Measure for Indefinite Rankings" (Webber et al.). Two ranked lists with high rbo are very similar, wheras low rbo indicates dissimilar lists. rbo ranges between 0 and 1. In this method the extrapolated version of rbo is implemented.

Usage

```
rbo(s, t, p, k = floor(max(length(s), length(t))/2), side = c("top",
  "bottom"), mid = NULL, uneven.lengths = TRUE)
```

Arguments

s	List 1
t	List 2
p	Weighting parameter in [0, 1]. High p implies strong emphasis on top ranked elements
k	Evaluation depth for extrapolation
side	Evaluate similarity between the top or the bottom of the ranked lists
mid	Set the mid point to for example only consider positive or negative scores
uneven.lengths	Indicator if lists have uneven lengths

Value

rank biased overlap (rbo)

Author(s)

Fabian Schmich

See Also

concordance

Examples

```
a <- rnorm(26)
b <- rnorm(26)
names(a) <- names(b) <- LETTERS
rbo(a, b, p = 0.95)
```

scores *scores*

Description

Return a named vector of phenotype scores

Usage

```
## S4 method for signature 'Phenotypes'
scores(object)

## S4 method for signature 'gespeR'
scores(object, type = c("GSP", "SSP"))
```

Arguments

object	A gespeR or Phenotypes object
type	The type of phenotype scores (GSP, SSP)

Value

A named vector of scores for each phenotype identifier

Author(s)

Fabian Schmich

See Also

gespeR

Phenotypes

Examples

```
data(stabilityfits)
scores(stabilityfits$A)
```

simData — *Example data: Simulated phenotypes and target relations for 4 screens (A, B, C, D)*

Description

The data set contains simulated data for four screens. Each screen consists of a phenotype vector and target relations between siRNAs and genes, i.e. which siRNA binds which genes (on- and off-targets). The size of each simulated screen is N = 1000 siRNAs x p = 1500 genes. SSPs are generated by first defining GSPs and multiplying the true GSPs with the sampled targe trelation matrices. For sampling the GSPs, we set the number of effect genes to 5 from Normal(0, 3). Target relation matrices are simulated by sampling the number of off-targets per siRNA from Normal(3e-2 * N, 3e-3 * N) and the strength of off-targets is sampled from Beta(2, 5). On-target components are set to 0.75.

Details

The code used to simluate the data can be found in system.file("example", "data_simulation.R", package="gespeR")

Examples

```
pheno.a <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package="gespeR"),
type = "SSP", col.id = 1, col.score = 2)
targets.a <- TargetRelations(system.file("extdata", "TR_screen_A.rds", package="gespeR"))
```

stability — *stability*

Description

Retrieve a Phenotypes object with stability values from a gespeR object.

Usage

```
stability(object)

## S4 method for signature 'gespeR'
stability(object)
```

Arguments

object	A gespeR object

Value

A Phenotypes object of SSPs

Author(s)

Fabian Schmich

Examples

```
phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
type = "SSP",
col.id = 1,
col.score = 2)
trels <- TargetRelations(readRDS(system.file("extdata", "TR_screen_A.rds", package = "gespeR")))
res <- gespeR(phenotypes = phenos,
 target.relations = trels,
 mode = "stability",
 nbootstrap = 100,
 fraction = 0.67,
 threshold = 0.75,
 EV = 1,
 weakness = 0.8,
 ncores = 1)
stab <- stability(res)
ans <- merge(as.data.frame(gsp(res)), as.data.frame(stability(res)), by = "ID")
colnames(ans)[2:3] <- c("Phenotype", "Stability")
ans[order(ans$Stability, decreasing = TRUE),]
```

---

stability.selection *Stability Selection*

Description

Based on Meinshausen and Buehlmann (2009)

Usage

```
stability.selection(x, y, fraction = 0.5, threshold = 0.75, EV = 1,
  nbootstrap = 100, weakness = 1, intercept = FALSE, ncores = 1, ...)
```

Arguments

x	The design matrix
y	The response vector
fraction	The fraction for each bootstrap sample
threshold	The selection threshold
EV	The expected value of wrongly selected elements
nbootstrap	The number of bootstrap samples
weakness	The weakness parameter for randomised lasso
intercept	Indicator, whether to fit an intercept
ncores	The number of cores for parallel computation
...	Additional arguments to lasso.rand

Value

A list containing selected covariates with frequencies, and the fitted model

Author(s)

Fabian Schmich

`stabilityfits` *Example fits for phenotypes from simulated screening data A, B, C and D*

Description

The data set contains four fitted gespeR models using stability selection from the four simulated screens.

Examples

```
data(stabilityfits)
```

`target.relations` *target.relations*

Description

Retrieve siRNA-to-gene target relations from a `gespeR` object.

Usage

```
target.relations(object)

## S4 method for signature 'gespeR'
target.relations(object)
```

Arguments

`object`	A `gespeR` object

Value

A `TargetRelations` object

Author(s)

Fabian Schmich

Examples

```
data(stabilityfits)
target.relations(stabilityfits$A)
```

`TargetRelations-class` *TargetRelations*

Description

Class used to represent siRNA-to-gene on- and off-target relations for a knockdown library and a set of genes.

Usage

```
TargetRelations(targets)

## S4 method for signature 'character'
TargetRelations(targets)

## S4 method for signature 'Matrix'
TargetRelations(targets)
```

Arguments

`targets`	Path to a .rds target relations matrix file or `Matrix` object

Value

A `TargetRelations` object

Slots

`siRNAs` The siRNA identifiers

`genes` The gene identifiers (Entrez)

`path` The path to and `.rds` `TargetRelations` file

`is.loaded` An indicator if target relations values are loaded

`values` The quantitative target relation values between siRNAs and genes

Author(s)

Fabian Schmich

See Also

join

loadValues

unloadValues

writeValues

values

path<-

Examples

```
trels <- TargetRelations(readRDS(system.file("extdata", "TR_screen_A.rds", package = "gespeR")))
```

---

values *values*

Description

Retrieve the numeric values from a TargetRelations or Phenotypes object

Usage

```
values(object)

## S4 method for signature 'TargetRelations'
values(object)

## S4 method for signature 'Phenotypes'
values(object)
```

Arguments

object	A TargetRelations or Phenotypes object

Value

A Matrix object

Author(s)

Fabian Schmich

Examples

```
trels <- TargetRelations(readRDS(system.file("extdata", "TR_screen_A.rds", package = "gespeR")))
values(trels)[1:5, 1:5]
phenos <- Phenotypes(system.file("extdata", "Phenotypes_screen_A.txt", package = "gespeR"),
type = "SSP",
col.id = 1,
col.score = 2)
values(phenos)
```

[,Phenotypes,ANY,ANY,ANY-method

Subsetting for Phenotype objects.

Description

Subsetting for Phenotype objects.

Usage

```
## S4 method for signature 'Phenotypes,ANY,ANY,ANY'
x[i, j, ..., drop = TRUE]
```

Arguments

x	A Phenotypes object
i	The subsetting indices for siRNAs
j	Subsetting indices for multivariate phenotypes
...	Additional parameters
drop	Drop Redundant Extent Information

Value

A Phenotypes object

Author(s)

Fabian Schmich

`[,TargetRelations,ANY,ANY,ANY-method`
Subsetting for TargetRelations objects.

Description

Subsetting for TargetRelations objects.

Usage

```
## S4 method for signature 'TargetRelations,ANY,ANY,ANY'
x[i, j, ..., drop = TRUE]
```

Arguments

`x`	A `TargetRelations` object
`i`	The row subsetting indices (siRNAs)
`j`	The column subsetting indeces (genes)
`...`	Additional parameters
`drop`	Drop Redundant Extent Information

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