

Package ‘DeProViR’

May 24, 2024

Type Package

Title A Deep-Learning Framework Based on Pre-trained Sequence Embeddings for Predicting Host-Viral Protein-Protein Interactions

Version 1.1.0

Description Emerging infectious diseases, exemplified by the zoonotic COVID-19 pandemic caused by SARS-CoV-2, are grave global threats. Understanding protein-protein interactions (PPIs) between host and viral proteins is essential for therapeutic targets and insights into pathogen replication and immune evasion. While experimental methods like yeast two-hybrid screening and mass spectrometry provide valuable insights, they are hindered by experimental noise and costs, yielding incomplete interaction maps. Computational models, notably DeProViR, predict PPIs from amino acid sequences, incorporating semantic information with GloVe embeddings. DeProViR employs a Siamese neural network, integrating convolutional and Bi-LSTM networks to enhance accuracy. It overcomes the limitations of feature engineering, offering an efficient means to predict host-virus interactions, which holds promise for antiviral therapies and advancing our understanding of infectious diseases.

License MIT+ file LICENSE

Encoding UTF-8

URL <https://github.com/mrbakhsh/DeProViR>

BugReports <https://github.com/mrbakhsh/DeProViR/issues>

Depends keras

Imports caret, data.table, dplyr, fmsb, ggplot2, grDevices, pROC, PRROC, readr, stats, BiocFileCache, utils

VignetteBuilder knitr

Suggests rmarkdown, tensorflow, BiocStyle, RUnit, knitr, BiocGenerics

biocViews Proteomics, SystemsBiology, NetworkInference, NeuralNetwork, Network

RoxygenNote 7.3.1

git_url <https://git.bioconductor.org/packages/DeProViR>

git_branch devel**git_last_commit** 4c1b40b**git_last_commit_date** 2024-04-30**Repository** Bioconductor 3.20**Date/Publication** 2024-05-24**Author** Matineh Rahmatbakhsh [aut, trl, cre]**Maintainer** Matineh Rahmatbakhsh <matinerb.94@gmail.com>

Contents

encodeHostSeq	2
encodeViralSeq	3
gloveImport	4
loadPreTrainedModel	4
loadTrainingSet	5
modelTraining	6
performancePlots	8
predInteractions	8
Index	10

 encodeHostSeq

Host Protein Sequence Encoding with GloVe Embedding Vectors

Description

This function first encodes amino acids as a sequence of unique 20 integers through tokenizer. The padding token was added to the front of shorter sequences to ensure a fixed-length vector of defined size L (i.e., here is 1000). Embedding matrix is then constructed to transform amino acid tokens to pre-training embedding weights, in which rows represent the amino acid tokens created earlier, and columns correspond to 100-dimension weight vectors derived from GloVe word-vector-generation vector map.

Usage

```
encodeHostSeq(trainingSet, embeddings_index)
```

Arguments

`trainingSet` a data.frame containing training information
`embeddings_index` embedding outputted from `gloveImport`

Value

A list containing Embedding matrix and tokenization

Examples

```
# Download and load the index
embeddings_index <- gloveImport()
#load training set
dt <- loadTrainingSet()
#encoding
encodeHostSeq <- encodeHostSeq(dt, embeddings_index)
```

encodeViralSeq

Viral Protein Sequence Encoding with GloVe Embedding Vectors

Description

This function first encodes amino acids as a sequence of unique 20 integers through tokenizer. The padding token was added to the front of shorter sequences to ensure a fixed-length vector of defined size L (i.e., here is 1000). Embedding matrix is then constructed to transform amino acid tokens to pre-training embedding weights, in which rows represent the amino acid tokens created earlier, and columns correspond to 100-dimension weight vectors derived from GloVe word-vector-generation vector map.

Usage

```
encodeViralSeq(trainingSet, embeddings_index)
```

Arguments

`trainingSet` a data.frame containing training information
`embeddings_index` embedding outputted from [gloveImport](#)

Value

A list containing Embedding matrix and tokenization

Examples

```
# Download and load the index
embeddings_index <- gloveImport()
#load training set
dt <- loadTrainingSet()
#encoding
encoded_seq <- encodeViralSeq(dt, embeddings_index)
```

gloveImport *Cache and Load Pre-Trained Word Vectors*

Description

This function cache and loads pre-trained GloVe vectors (100d).

Usage

```
gloveImport(url_path = "https://nlp.stanford.edu/data")
```

Arguments

url_path URL path to GloVe embedding. Defaults to "https://nlp.stanford.edu/data"

Value

glove embedding

Examples

```
options(timeout=240)
embeddings_index <- gloveImport(url_path = "https://nlp.stanford.edu/data")
```

loadPreTrainedModel *Load Pre-Trained Model Weights*

Description

This function loads the pre-trained model weights constructed previously using [modelTraining](#)

Usage

```
loadPreTrainedModel(
  input_dim = 20,
  output_dim = 100,
  filters_layer1CNN = 32,
  kernel_size_layer1CNN = 16,
  filters_layer2CNN = 64,
  kernel_size_layer2CNN = 7,
  pool_size = 30,
  layer_lstm = 64,
  units = 8,
  metrics = "AUC",
  filepath = system.file("extdata", "Pre_trainedModel", package = "DeProViR")
)
```

Arguments

input_dim	Integer. Size of the vocabulary, i.e. amino acid tokens. Defaults to 20. See keras.
output_dim	Integer. Dimension of the dense embedding, i.e., GloVe. Defaults to 100. See keras.
filters_layer1CNN	Integer, the dimensionality of the output space (i.e. the number of output filters in the first convolution). Defaults to 32. See keras
kernel_size_layer1CNN	An integer or tuple/list of 2 integers, specifying the height and width of the convolution window in the first layer. Can be a single integer to specify the same value for all spatial dimensions. Defaults to 16. See keras
filters_layer2CNN	Integer, the dimensionality of the output space (i.e. the number of output filters in the second convolution). Defaults to 64. See keras
kernel_size_layer2CNN	An integer or tuple/list of 2 integers, specifying the height and width of the convolution window in the second layer. Can be a single integer to specify the same value for all spatial dimensions. Defaults to 7. See keras
pool_size	Down samples the input representation by taking the maximum value over a spatial window of size pool_size. Defaults to 30. See keras
layer_lstm	Number of units in the Bi-LSTM layer. Defaults to 64. See keras
units	Number of units in the MLP layer. Defaults to 8. See keras
metrics	Vector of metric names to be evaluated by the model during training and testing. Defaults to "AUC". See keras
filepath	A character string indicating the path contained pre-trained model weights, i.e., inst/extdata/Pre-trainedModel

Value

Pre-trained model.

Examples

```
Loading_trainedModel <- loadPreTrainedModel()
```

loadTrainingSet	<i>Load Demo Training Set</i>
-----------------	-------------------------------

Description

This function loads demo training set.

Usage

```
loadTrainingSet(  
  training_dir = system.file("extdata", "training_Set", package = "DeProViR")  
)
```

Arguments

training_dir dir containing a training data.frame .csv Default set to "extdata/training_testSets".

Value

data.frame

Examples

```
dt <- loadTrainingSet()
```

modelTraining

Predictive Model Training using k-fold Validation Strategy

Description

This function first transforms protein sequences to amino acid tokens wherein tokens are indexed by positive integers, then represents each amino acid token by pre-trained co-occurrence embedding vectors learned by GloVe, followed by applying an embedding layer. Then it employs Siamese-like neural network architecture on densely-connected neural net to predict interactions between host and viral proteins.

Usage

```
modelTraining(  
  url_path = "https://nlp.stanford.edu/data",  
  training_dir = system.file("extdata", "training_Set", package = "DeProViR"),  
  input_dim = 20,  
  output_dim = 100,  
  filters_layer1CNN = 32,  
  kernel_size_layer1CNN = 16,  
  filters_layer2CNN = 64,  
  kernel_size_layer2CNN = 7,  
  pool_size = 30,  
  layer_lstm = 64,  
  units = 8,  
  metrics = "AUC",  
  cv_fold = 10,  
  epochs = 100,  
  batch_size = 128,  
  plots = TRUE,
```

```

    tpath = tempdir(),
    save_model_weights = TRUE,
    filepath = tempdir()
)

```

Arguments

<code>url_path</code>	URL path to GloVe embedding. Defaults to "https://nlp.stanford.edu/data/glove.6B.zip".
<code>training_dir</code>	dir containing viral-host training set. See loadTrainingSet
<code>input_dim</code>	Integer. Size of the vocabulary, i.e. amino acid tokens. Defaults to 20. See keras.
<code>output_dim</code>	Integer. Dimension of the dense embedding, i.e., GloVe. Defaults to 100. See keras.
<code>filters_layer1CNN</code>	Integer, the dimensionality of the output space (i.e. the number of output filters in the first convolution). Defaults to 32. See keras
<code>kernel_size_layer1CNN</code>	An integer or tuple/list of 2 integers, specifying the height and width of the convolution window in the first layer. Can be a single integer to specify the same value for all spatial dimensions. Defaults to 16. See keras
<code>filters_layer2CNN</code>	Integer, the dimensionality of the output space (i.e. the number of output filters in the second convolution). Defaults to 64. See keras
<code>kernel_size_layer2CNN</code>	An integer or tuple/list of 2 integers, specifying the height and width of the convolution window in the second layer. Can be a single integer to specify the same value for all spatial dimensions. Defaults to 7. See keras
<code>pool_size</code>	Down samples the input representation by taking the maximum value over a spatial window of size <code>pool_size</code> . Defaults to 30. See keras
<code>layer_lstm</code>	Number of units in the Bi-LSTM layer. Defaults to 64. See keras
<code>units</code>	Number of units in the MLP layer. Defaults to 8. See keras
<code>metrics</code>	Vector of metric names to be evaluated by the model during training and testing. Defaults to "AUC". See keras
<code>cv_fold</code>	Number of partitions for cross-validation. Defaults to 10.
<code>epochs</code>	Number of epochs to train the model. Defaults to 100. See keras
<code>batch_size</code>	Number of samples per gradient update. Defaults to 128. See keras
<code>plots</code>	PDF file containing performance measures. Defaults to TRUE. See performancePlots
<code>tpath</code>	A character string indicating the path to the project directory. If the directory is missing, PDF file containing performance measures will be stored in the Temp directory. See performancePlots
<code>save_model_weights</code>	If TRUE, save the trained weights. Defaults to TRUE.
<code>filepath</code>	A character string indicating the path to save the model weights. Default to <code>tempdir()</code> .

Value

Trained model and performance measures.

performancePlots *Model Performance Evaluation*

Description

This function plots model performance

Usage

```
performancePlots(pred_label, y_label, tpath = tempdir())
```

Arguments

pred_label	predicted labels
y_label	Ground truth labels
tpath	A character string indicating the path to the project directory. If the directory is missing, PDF file will be stored in the Temp directory.

Value

Pdf file containing performance plots

Examples

```
pred_label <- seq(0,1, length.out = 100)
truth_label <- rep(c(0,1), each = 50)
perf <- performancePlots(pred_label,
                          truth_label,
                          tpath = tempdir())
```

predInteractions *Predict Unknown Interactions*

Description

This function initially constructs an embedding matrix from the viral or host protein sequences and then predicts scores for unknown interactions. Interactions with scores greater than 0.5 are more likely to indicate interaction.

Usage

```
predInteractions(  
  url_path = "https://nlp.stanford.edu/data",  
  Testingset,  
  trainedModel  
)
```

Arguments

<code>url_path</code>	URL path to GloVe embedding. Defaults to "https://nlp.stanford.edu/data/glove.6B.zip".
<code>Testingset</code>	A data.frame containing unknown interactions. For demo, we can use the file in <code>extdata/test_Set</code> .
<code>trainedModel</code>	Pre-trained model stored in <code>extdata/Pre_trainedModel</code> or the training model "\$merge_model" achieved by modelTraining .

Value

Probability scores for unknown interactions

Examples

```
trainedModel <- loadPreTrainedModel()  
# load test set (i.e., unknown interactions)  
testing_set <- data.table::fread(system.file("extdata", "test_Set",  
                                           "test_set_unknownInteraction.csv",  
                                           package = "DeProViR"))  
  
# now predict interactions  
options(timeout=240)  
predInteractions <- predInteractions(url_path = "https://nlp.stanford.edu/data",  
                                     testing_set,  
                                     trainedModel)
```

Index

`encodeHostSeq`, [2](#)
`encodeViralSeq`, [3](#)

`gloveImport`, [2](#), [3](#), [4](#)

`loadPreTrainedModel`, [4](#)
`loadTrainingSet`, [5](#), [7](#)

`modelTraining`, [4](#), [6](#), [9](#)

`performancePlots`, [7](#), [8](#)
`predInteractions`, [8](#)