Package 'CytoML'

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Type Package

Title GatingML interface for openCyto

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Description This package is designed to use GatingML2.0 as the standard format to exchange the gated data with other software platform.

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LazyData TRUE

Imports flowCore, flowWorkspace (>= 3.21.10), openCyto (>= 1.11.3), XML, data.table, flowUtils (>= 1.35.7), jsonlite, RBGL, ncdfFlow, Rgraphviz, Biobase, methods, graph, graphics, utils, base64enc, plyr

biocViews FlowCytometry, DataImport, DataRepresentation

Suggests testthat, flowWorkspaceData (>= 2.11.1), knitr, ggcyto

LinkingTo Rcpp, flowWorkspace, BH(>= 1.62.0-1)

VignetteBuilder knitr

SystemRequirements xml2, GNU make, C++11

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Collate 'GatingSet2cytobank.R' 'GatingSet2flowJo.R' 'RcppExports.R' 'cytobank2GatingSet.R' 'diva2GatingSet.R' 'flowUtils_functions.R' 'read.gatingML.cytobank.R' 'graphGML_methods.R' 'set.count.xml.R' 'utils.R'

NeedsCompilation yes

R topics documented:

`addCustomInfo` *add customInfo nodes to each gate node and add BooleanAndGates*

Description

add customInfo nodes to each gate node and add BooleanAndGates

Usage

```
addCustomInfo(root, gs, flowEnv, cytobank.default.scale = TRUE, showHidden)
```

Arguments

`root`	the root node of the XML
`gs`	a GatingSet object
`flowEnv`	the environment that stores the information parsed by 'read.GatingML'.
`cytobank.default.scale`	logical flag indicating whether to use the default Cytobank asinhtGml2 settings. Currently it should be set to TRUE in order for gates to be displayed properly in Cytobank because cytobank currently does not parse the global scale settings from GatingML.
`showHidden`	whether to include the hidden population nodes in the output

Value

XML root node

`compare.counts` *compare the counts to cytobank's exported csv so that the parsing result can be verified.*

Description

compare the counts to cytobank's exported csv so that the parsing result can be verified.

Usage

```
compare.counts(gs, file, id.vars = c("FCS Filename", "population"))
```

Arguments

`gs`	parsed GatingSet
`file`	the stats file (contains the populatio counts) exported from cytobank.
`id.vars`	either "population" or "FCS filename" that tells whether the stats file format is one population per row or FCS file per row.

Value

a data.table (in long format) that contains the counts from openCyto and Cytobank side by side.

Examples

```
xmlfile <- system.file("extdata/cytotrol_tcell_cytobank.xml", package = "CytoML")
fcsFiles <- list.files(pattern = "CytoTrol", system.file("extdata", package = "flowWorkspaceData"), full = TR
gs <- cytobank2GatingSet(xmlfile, fcsFiles)
## verify the stats are correct
statsfile <- system.file("extdata/cytotrol_tcell_cytobank_counts.csv", package = "CytoML")
dt_merged <- compare.counts(gs, statsfile, id.vars = "population")
all.equal(dt_merged[, count.x], dt_merged[, count.y], tol = 5e-4)
```

`compensate,GatingSet,graphGML-method` *compensate a GatingSet based on the compensation information stored in graphGML object*

Description

compensate a GatingSet based on the compensation information stored in graphGML object

Usage

```
## S4 method for signature 'GatingSet,graphGML'
compensate(x, spillover, ...)
```

Arguments

x	GatingSet
spillover	graphGML
...	unused.

Value

compensated GatingSet

---

constructTree *Reconstruct the population tree from the GateSets*

Description

Reconstruct the population tree from the GateSets

Usage

```
constructTree(flowEnv, gateInfo)
```

Arguments

flowEnv	the enivornment contains the elements parsed by read.gatingML function
gateInfo	the data.frame contains the gate name, fcs filename parsed by parse.gateInfo function

Value

a graphNEL represent the population tree. The gate and population name are stored as nodeData in each node.

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cytobank2GatingSet *A wrapper that parse the gatingML and FCS files into GatingSet*

Description

A wrapper that parse the gatingML and FCS files into GatingSet

Usage

```
cytobank2GatingSet(xml, FCS)
```

Arguments

xml	the full path of gatingML file
FCS	FCS files to be loaded

Value

a GatingSet

Examples

```
xmlfile <- system.file("extdata/cytotrol_tcell_cytobank.xml", package = "CytoML")
fcsFiles <- list.files(pattern = "CytoTrol", system.file("extdata", package = "flowWorkspaceData"), full = TR
gs <- cytobank2GatingSet(xmlfile, fcsFiles)
#plotGate(gs[[1]])
```

---

divaWorkspace-class *divaWorkspace class Inherited from flowJoWorkspace-class*

Description

divaWorkspace class Inherited from flowJoWorkspace-class

Usage

```
## S4 method for signature 'divaWorkspace'
getSamples(x)

## S4 method for signature 'divaWorkspace'
getSampleGroups(x)

## S4 method for signature 'divaWorkspace'
show(object)

## S4 method for signature 'divaWorkspace'
parseWorkspace(obj, ...)
```

---

extend *extend the gate to the minimum and maximum limit of both dimensions based on the bounding information.*

Description

It is equivalent to the behavior of shifting the off-scale boundary events into the gate boundary that is described in bounding transformation section of gatingML standard.

Usage

```
extend(gate, bound, data.range = NULL, plot = FALSE,
  limits = c("original", "extended"))

## S3 method for class 'polygonGate'
extend(gate, bound, data.range = NULL, plot = FALSE,
  limits = c("original", "extended"))

## S3 method for class 'rectangleGate'
extend(gate, ...)

## S3 method for class 'ellipsoidGate'
extend(gate, ...)
```

Arguments

gate	a flowCore filter/gate
bound	numeric matrix representing the bouding information parsed from gatingML. Each row corresponds to a channel. rownames should be the channel names. colnames should be c("min", "max")
data.range	numeric matrix specifying the data limits of each channel. It is used to set the extended value of vertices and must has the same structure as 'bound'. when it is not supplied, c(-.Machine$integer.max, - .Machine$integer.max) is used.
plot	whether to plot the extended polygon.
limits	character whether to plot in "extended" or "original" gate limits. Default is "original".
...	other arguments

Details

The advantage of extending gates instead of shifting data are two folds: 1. Avoid the extra computation each time applying or plotting the gates 2. Avoid changing the data distribution caused by adding the gates

Normally this function is not used directly by user but invoked when parsing GatingML file exported from Cytobank.

Value

a flowCore filter/gate

Examples

```
library(flowCore)
sqrcut <- matrix(c(300,300,600,600,50,300,300,50),ncol=2,nrow=4)
colnames(sqrcut) <- c("FSC-H","SSC-H")
pg <- polygonGate(filterId="nonDebris", sqrcut)
pg
bound <- matrix(c(100,3e3,100,3e3), byrow = TRUE, nrow = 2, dimnames = list(c("FSC-H", "SSC-H"), c("min", "ma
bound
pg.extened <- extend(pg, bound, plot = TRUE)
```

gating,graphGML,GatingSet-method

Apply the gatingML graph to a GatingSet

Description

It applies the gates to the GatingSet based on the population tree described in graphGML.

Usage

```
## S4 method for signature 'graphGML,GatingSet'
gating(x, y, ...)
```

Arguments

x	graphGML
y	GatingSet
...	other arguments

Value

Nothing. As the side effect, gates generated by gating methods are saved in GatingSet.

GatingSet2cytobank *Convert a GatingSet to a Cytobank-compatible gatingML*

Description

this function retrieves the gates from GatingSet and writes a customed GatingML-2.0 file that can be imported into cytobank.

Usage

```
GatingSet2cytobank(gs, outFile, showHidden = FALSE,
  cytobank.default.scale = TRUE, ...)
```

Arguments

gs	a GatingSet object
outFile	a file name
showHidden	whether to include the hidden population nodes in the output
cytobank.default.scale	logical flag indicating whether to use the default Cytobank asinhtGml2 settings. Currently it should be set to TRUE in order for gates to be displayed properly in Cytobank because cytobank currently does not parse the global scale settings from GatingML.
...	rescale.gate default is TRUE. which means the gate is rescaled to the new scale that is understandable by cytobank. It is recommended not to change this behavior unless user wants to export to a gatingML file used for other purpose other than being imported into cytobank.

Details

The process can be divided into four steps: 1. Read in gate geometry, compensation and transformation from gatingSet 2. Rescale gate boundaries with flowJoTrans() so gates can be displayed properly in Cytobank 3. Save gates and hierarchy structure to R environment 4. Write environment out to gatingML using write.GatingML()

Value

nothing

Examples

```
library(flowWorkspace)

dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))

Rm("CD8", gs)

#output to cytobank
outFile <- tempfile(fileext = ".xml")
GatingSet2cytobank(gs, outFile) #type by default is 'cytobank'
```

`GatingSet2flowJo` *Convert a GatingSet to flowJo workspace*

Description

Convert a GatingSet to flowJo workspace

Usage

```
GatingSet2flowJo(gs, outFile, ...)
```

Arguments

`gs`	a GatingSet object
`outFile`	the workspace file path to write
`...`	other arguments showHidden whether to include the hidden population nodes in the output

Value

nothing

Examples

```
library(flowWorkspace)

dataDir <- system.file("extdata",package="flowWorkspaceData")
gs <- load_gs(list.files(dataDir, pattern = "gs_manual",full = TRUE))

#output to flowJo
outFile <- tempfile(fileext = ".wsp")
GatingSet2flowJo(gs, outFile)
```

---

getChildren,graphGML,character-method
get children nodes

Description

get children nodes

Usage

```
## S4 method for signature 'graphGML,character'
getChildren(obj, y)
```

Arguments

obj	graphGML
y	character parent node path

Value

a graphNEL node

Examples

```
xmlfile <- system.file("extdata/cytotrol_tcell_cytobank.xml", package = "CytoML")
g <- read.gatingML.cytobank(xmlfile)
getChildren(g, "GateSet_722326")
getParent(g, "GateSet_722326")
```

getCompensationMatrices,graphGML-method
Extract compensation from graphGML object.

Description

Extract compensation from graphGML object.

Usage

```
## S4 method for signature 'graphGML'
getCompensationMatrices(x)
```

Arguments

x	graphGML

Value

compensation object or "FCS" when compensation comes from FCS keywords

getGate,graphGML,character-method
get gate from the node

Description

get gate from the node

Usage

```
## S4 method for signature 'graphGML,character'
getGate(obj, y)
```

Arguments

obj	graphGML
y	character node path

Value

the gate information associated with the node

getNodes,graphGML-method

get nodes from graphGML object

Description

get nodes from graphGML object

Usage

```
## S4 method for signature 'graphGML'
getNodes(x, y, order = c("default", "bfs", "dfs",
  "tsort"), only.names = TRUE)
```

Arguments

x	graphGML
y	character node index. When missing, return all the nodes
order	character specifying the order of nodes. options are "default", "bfs", "dfs", "tsort"
only.names	logical specifiying whether user wants to get the entire nodeData or just the name of the population node

Value

It returns the node names and population names by default. Or return the entire nodeData associated with each node.

Examples

```
xmlfile <- system.file("extdata/cytotrol_tcell_cytobank.xml", package = "CytoML")
g <- read.gatingML.cytobank(xmlfile)
getNodes(g)
getNodes(g, only.names = FALSE)
```

getParent,graphGML,character-method

get parent nodes

Description

get parent nodes

Usage

```
## S4 method for signature 'graphGML,character'
getParent(obj, y)
```

Arguments

obj	graphGML
y	character child node path

Value

a graphNEL node

---

getTransformations,graphGML-method *Extract transformations from graphGML object.*

Description

Extract transformations from graphGML object.

Usage

```
## S4 method for signature 'graphGML'
getTransformations(x)
```

Arguments

x	graphGML

Value

transformerList object

---

graphGML-class *A graph object returned by 'read.gatingML.cytobank' function.*

Description

Each node corresponds to a population(or GateSet) defined in gatingML file. The actual gate object (both global and tailored gates) is associated with each node as nodeData. Compensation and transformations are stored in graphData slot.

Details

The class simply extends the graphNEL class and exists for the purpose of method dispatching.

matchPath — *Given the leaf node, try to find out if a collection of nodes can be matched to a path in a graph(tree) by the bottom-up searching*

Description

Given the leaf node, try to find out if a collection of nodes can be matched to a path in a graph(tree) by the bottom-up searching

Usage

```
matchPath(g, leaf, nodeSet)
```

Arguments

g	graphNEL
leaf	the name of leaf(terminal) node
nodeSet	a set of node names

Value

TRUE if path is found, FALSE if not path is matched.

openDiva — *open Diva xml workspace*

Description

open Diva xml workspace

Usage

```
openDiva(file, options = 0, ...)
```

Arguments

file	xml file
options	argument passed to xmlTreeParse
...	arguments passed to xmlTreeParse

Value

a divaWorkspace object

Examples

```
## Not run:
library(flowWorkspace)
library(CytoML)
ws <- openDiva(system.file('extdata/diva/PE_2.xml', package = "CytoML"))
ws
getSampleGroups(ws)
getSamples(ws)
gs <- parseWorkspace(ws, name = 2, subset = 1)
sampleNames(gs)
getNodes(gs)
plotGate(gs[[1]])

## End(Not run)
```

`parse.gateInfo` *Parse the cytobank custom_info for each gate*

Description

Fcs filename and gate name stored in 'custom_info' element are beyong the scope of the gatingML standard and thus not covered by the default 'read.gatingML'.

Usage

```
parse.gateInfo(file, ...)
```

Arguments

`file`	xml file path
`...`	additional arguments passed to the handlers of 'xmlTreeParse'

Value

a data.frame that contains three columns: id (gateId), name (gate name), fcs (fcs_file_filename).

`plot,graphGML,missing-method`
plot the population tree stored in graphGML.

Description

The node with dotted order represents the population that has tailored gates (sample-specific gates) defined.

Usage

```
## S4 method for signature 'graphGML,missing'
plot(x, y = "missing", label = c("popName",
  "gateName"))
```

Arguments

x	a graphNEL generated by constructTree function
y	not used
label	specifies what to be dispaled as node label. Can be either 'popName' (population name parsed from GateSets) or 'gateName'(the name of the actual gate associated with each node)

Value

nothing

Examples

```
xmlfile <- system.file("extdata/cytotrol_tcell_cytobank.xml", package = "CytoML")
g <- read.gatingML.cytobank(xmlfile)
plot(g)
```

---

read.gatingML.cytobank

Parser for gatingML exported by Cytobank

Description

The Default parser (flowUtils::read.gatingML) does not parse the population tree as well as the custom information from cytobank. (e.g. gate name, fcs filename).

Usage

```
read.gatingML.cytobank(file, ...)
```

Arguments

file	Gating-ML XML file
...	additional arguments passed to the handlers of 'xmlTreeParse'

Value

a graphGML that represents the population tree. The gate and population name are stored in nodeData of each node. Compensation and transformations are stored in graphData.

Examples

```
xml <- system.file("extdata/cytotrol_tcell_cytobank.xml", package = "CytoML")
g <- read.gatingML.cytobank(xml) #parse the population tree
#plot(g) #visualize it
```

set.count.xml *save the event counts parsed from xml into c++ tree structure*

Description

It is for internal use by the diva parser

Usage

```
set.count.xml(gh, node, count)
```

Arguments

gh	GatingHierarchy
node	the unique gating path that uniquely identifies a population node
count	integer number that is events count for the respective gating node directly parsed from xml file

Examples

```
## Not run:
set.count.xml(gh, "CD3", 10000)

## End(Not run)
```

show,graphGML-method *show method for graphGML*

Description

show method for graphGML

Usage

```
## S4 method for signature 'graphGML'
show(object)
```

Arguments

object	graphGML

Value

nothing

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