

Package ‘DAPAR’

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Type Package

Title Tools for the Differential Analysis of Proteins Abundance with R

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Description This package contains a collection of functions for the
visualisation and the statistical analysis of proteomic data.

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boxPlotD	<i>Builds a boxplot from a dataframe</i>
----------	--

Description

Boxplot for quantitative proteomics data

Usage

```
boxPlotD(qData, dataForXAxis = NULL, labels = NULL,
         group2Color = "Condition")
```

Arguments

qData	A dataframe that contains quantitative data.
dataForXAxis	A vector containing the types of replicates to use as X-axis. Available values are: Label, Analyt.Rep, Bio.Rep and Tech.Rep. Default is "Label".
labels	A vector of the conditions (labels) (one label per sample).
group2Color	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

Value

A boxplot

Author(s)

Florence Combes, Samuel Wieczorek

See Also

[densityPlotD](#)

Examples

```
data(UPSep25)
qData <- Biobase::exprs(UPSep25)
types <- c("Label", "Analyt.Rep")
dataForXAxis <- Biobase::pData(UPSep25)[, types]
labels <- Biobase::pData(UPSep25)[, "Label"]
boxPlotD(qData, dataForXAxis, labels)
```

BuildAdjacencyMatrix *Function matrix of appartenance group*

Description

Method to create a binary matrix with proteins in columns and peptides in lines on a MSnSet object (peptides)

Usage

```
BuildAdjacencyMatrix(obj.pep, protID, unique = TRUE)
```

Arguments

obj.pep	An object (peptides) of class MSnbase .
protID	The name of proteins ID column
unique	A boolean to indicate whether only the unique peptides must be considered (TRUE) or if the shared peptides have to be integrated (FALSE).

Value

A binary matrix

Author(s)

Florence Combes, Samuel Wieczorek, Alexia Dorffer

Examples

```
data(UPSep25)
BuildAdjacencyMatrix(UPSep25, "Protein.group.IDs")
```

BuildColumnToProteinDataset

creates a column for the protein dataset after agregation by using the previous peptide dataset.

Description

This function creates a column for the protein dataset after agregation by using the previous peptide dataset.

Usage

```
BuildColumnToProteinDataset(peptideData, matAdj, columnName)
```

Arguments

peptideData	A data.frame of meta data of peptides. It is the fData of the MSnset object.
matAdj	The adjacency matrix used to agregate the peptides data.
columnName	The name of the column in fData(peptides_MSnset) that the user wants to keep in the new protein data.frame.

Value

A vector

Author(s)

Samuel Wieczorek

Examples

```
data(UPSsep25)
protID <- "Protein.group.IDs"
M <- BuildAdjacencyMatrix(UPSsep25, protID, FALSE)
data <- Biobase::fData(UPSsep25)
name <- "organism"
BuildColumnToProteinDataset(data, M, name )
```

BuildSparseAdjacencyMatrix

Function matrix of appartenance group

Description

Method to create a binary matrix with proteins in columns and peptides in lines on a MSnSet object (peptides)

Usage

```
BuildSparseAdjacencyMatrix(obj.pep, protID, unique = TRUE)
```

Arguments

obj.pep	An object (peptides) of class MSnbase .
protID	The name of proteins ID column
unique	A boolean to indicate whether only the unique peptides must be considered (TRUE) or if the shared peptides have to be integrated (FALSE).

Value

A binary matrix

Author(s)

Florence Combes, Samuel Wiczorek, Alexia Dorffer

Examples

```
data(UPSpep25)
BuildSparseAdjacencyMatrix(UPSpep25, "Protein.group.IDs", TRUE)
```

compareNormalizationD *Builds a plot from a dataframe*

Description

Plot to compare the quantitative proteomics data before and after normalization

Usage

```
compareNormalizationD(qDataBefore, qDataAfter, labelsForLegend = NULL,
  indData2Show = NULL, group2Color = "Condition")
```

Arguments

qDataBefore	A dataframe that contains quantitative data before normalization.
qDataAfter	A dataframe that contains quantitative data after normalization.
labelsForLegend	A vector of the conditions (labels) (one label per sample).
indData2Show	A vector of the indices of the columns to show in the plot. The indices are those of indices of the columns in the data.frame qDataBefore.
group2Color	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

Value

A plot

Author(s)

Samuel Wieczorek

Examples

```
data(UPSep25)
qDataBefore <- Biobase::exprs(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
qDataAfter <- normalizeD(qDataBefore,labels,"Median Centering",
"within conditions")
compareNormalizationD(qDataBefore, qDataAfter, labels)
```

corrMatrixD	<i>Displays a correlation matrix of the quantitative data of the exprs() table.</i>
-------------	---

Description

Correlation matrix based on a [MSnSet](#) object

Usage

```
corrMatrixD(qData, samplesData, gradientRate = 5)
```

Arguments

qData	A dataframe of quantitative data.
samplesData	A dataframe where lines correspond to samples and columns to the meta-data for those samples.
gradientRate	The rate parameter to control the exponential law for the gradient of colors

Value

A colored correlation matrix

Author(s)

Florence Combes, Samuel Wiczorek

Examples

```
data(UPSep25)
qData <- Biobase::exprs(UPSep25)
samplesData <- Biobase::pData(UPSep25)
corrMatrixD(qData, samplesData)
```

CountPep

Compute the number of peptides used to aggregate proteins

Description

This function computes the number of peptides used to aggregate proteins.

Usage

```
CountPep(M)
```

Arguments

M A "valued" adjacency matrix in which lines and columns correspond respectively to peptides and proteins.

Value

A vector of boolean which is the adjacency matrix but with NA values if they exist in the intensity matrix.

Author(s)

Alexia Dorffer

Examples

```
data(UPSep25)
protID <- "Protein.group.IDs"
M <- BuildAdjacencyMatrix(UPSep25, protID, FALSE)
CountPep(M)
```

createMSnset	<i>Creates an object of class MSnSet from text file</i>
--------------	---

Description

Builds an object of class [MSnSet](#) from a single tabulated-like file for quantitative and meta-data and a dataframe for the samples description. It differs from the original [MSnSet](#) builder which requires three separated files tabulated-like quantitative proteomic data into a [MSnSet](#) object, including meta-data.

Usage

```
createMSnset(file, metadata = NULL, indExpData, indFData, indiceID = NULL,  
             logData = FALSE, replaceZeros = FALSE, pep_prot_data = NULL)
```

Arguments

file	The name of a tab-separated file that contains the data.
metadata	A dataframe describing the samples (in lines).
indExpData	A vector of string where each element is the name of a column in designTable that have to be integrated in the <code>fData()</code> table of the MSnSet object.
indFData	The name of column in file that will be the name of rows for the <code>exprs()</code> and <code>fData()</code> tables
indiceID	The indice of the column containing the ID of entities (peptides or proteins)
logData	A boolean value to indicate if the data have to be log-transformed (Default is FALSE)
replaceZeros	A boolean value to indicate if the 0 and NaN values of intensity have to be replaced by NA (Default is FALSE)
pep_prot_data	A string that indicates whether the dataset is about peptides or proteins.

Value

An instance of class [MSnSet](#).

Author(s)

Florence Combes, Samuel Wieczorek

Examples

```
exprsFile <- system.file("extdata", "UPSsep25.txt", package="DAPAR")  
metadataFile <- system.file("extdata", "samples.txt", package="DAPAR")  
metadata = read.table(metadataFile, header=TRUE, sep="\t", as.is=TRUE)  
indExpData <- c(56:61)  
indFData <- c(1:55,62:71)  
indiceID <- 64
```

```
createMSnset(exprsFile, metadata, indExpData, indFData, indiceID,  
pep_prot_data = "peptide")
```

deleteLinesFromIndices

Delete the lines in the matrix of intensities and the metadata table given their indice.

Description

Delete the lines of `exprs()` table identified by their indice.

Usage

```
deletelinesFromIndices(obj, deleteThat = NULL, processText = NULL)
```

Arguments

<code>obj</code>	An object of class <code>MSnSet</code> containing quantitative data.
<code>deleteThat</code>	A vector of integers which are the indices of lines to delete.
<code>processText</code>	A string to be included in the <code>MSnSet</code> object for log.

Value

An instance of class `MSnSet` that have been filtered.

Author(s)

Florence Combes, Samuel Wiczorek

Examples

```
data(UPSep25)  
mvFilter(UPSep25, c(1:10))
```

densityPlotD	<i>Builds a densityplot from a dataframe</i>
--------------	--

Description

Densityplot of quantitative proteomics data over samples.

Usage

```
densityPlotD(qData, labelsForLegend = NULL, indData2Show = NULL,  
             group2Color = "Condition")
```

Arguments

qData	A dataframe that contains quantitative data.
labelsForLegend	A vector of the conditions (labels) (one label per sample).
indData2Show	A vector of indices to show in densityplot. If NULL, then all labels are displayed.
group2Color	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

Value

A density plot

Author(s)

Florence Combes, Samuel Wieczorek

See Also

[boxPlotD](#), [varianceDistD](#)

Examples

```
data(UPSep25)  
qData <- Biobase::exprs(UPSep25)  
labels <- lab2Show <- Biobase::pData(UPSep25)[,"Label"]  
densityPlotD(qData, labels)
```

diffAna	<i>This function performs a differential analysis on an MSnSet object (adapted from limma)</i>
---------	--

Description

Performs a differential analysis on an [MSnSet](#) object, based on [limma](#) functions.

Usage

```
diffAna(qData, design)
```

Arguments

qData	A dataframe that contains quantitative data.
design	The design matrix as described in the limma package documentation

Value

A dataframe with the p-value and log(Fold Change) associated to each element (peptide/protein)

Author(s)

Florence Combes, Samuel Wieczorek

Examples

```
data(UPSep25)
qData <- Biobase::exprs(UPSep25)
design <- cbind(cond1=1, cond2 = rep(0,nrow(Biobase::pData(UPSep25))))
rownames(design) <- rownames(Biobase::pData(UPSep25))
labels <- Biobase::pData(UPSep25)[,"Label"]
indices <- getIndicesConditions(labels, "25fmol", "10fmol")
design[indices$iCond,2] <- 1
diffAna(qData, design)
```

diffAnaComputeFDR	<i>Computes the FDR corresponding to the p-values of the differential analysis using</i>
-------------------	--

Description

This function is a wrapper to the function `adjust.p` from the `cp4p` package. It returns the FDR corresponding to the p-values of the differential analysis. The FDR is computed with the function `p.adjust{stats}`..

Usage

```
diffAnaComputeFDR(data, threshold_PVal = 0, threshold_LogFC = 0,  
  pi0Method = 1)
```

Arguments

data	The result of the differential analysis processed by diffAna
threshold_PVal	The threshold on p-value to distinguish between differential and non-differential data
threshold_LogFC	The threshold on log(Fold Change) to distinguish between differential and non-differential data
pi0Method	The parameter pi0.method of the method adjust.p in the package cp4p

Value

The computed FDR value (floating number)

Author(s)

Samuel Wieczorek

Examples

```
data(UPSep25)  
obj <- wrapper.mvImputation(UPSep25, "QRILC")  
condition1 <- '25fmol'  
condition2 <- '10fmol'  
qData <- Biobase::exprs(obj)  
samplesData <- Biobase::pData(obj)  
labels <- Biobase::pData(obj)[,"Label"]  
limma <- diffAnalimma(qData,samplesData, labels, condition1, condition2)  
diffAnaComputeFDR(limma)
```

`diffAnaGetSignificant` *Returns a MSnSet object with only proteins significant after differential analysis.*

Description

Returns a MSnSet object with only proteins significant after differential analysis.

Usage

```
diffAnaGetSignificant(obj)
```

Arguments

obj An object of class `MsnSet`.

Value

A `MsnSet`

Author(s)

Alexia Dorffer

Examples

```
data(UPSep25)
condition1 <- "25fmol"
condition2 <- "10fmol"
resLimma <- wrapper.diffAnaLimma(UPSep25, condition1, condition2)
obj <- diffAnaSave(UPSep25, resLimma, "limma", condition1, condition2)
signif <- diffAnaGetSignificant(obj)
```

diffAnaLimma	<i>Performs differential analysis on an <code>MsnSet</code> object, calling the <code>limma</code> package functions</i>
--------------	--

Description

Method to perform differential analysis on an `MsnSet` object (calls the `limma` package function).

Usage

```
diffAnaLimma(qData, samplesData, labels, condition1, condition2)
```

Arguments

qData A dataframe that contains quantitative data.

samplesData A dataframe where lines correspond to samples and columns to the meta-data for those samples.

labels A vector of the conditions (labels) (one label per sample).

condition1 A vector that contains the names of the conditions considered as condition 1

condition2 A vector that contains the names of the conditions considered as condition 2

Value

A dataframe as returned by the `limma` package

Author(s)

Florence Combes, Samuel Wiczorek

Examples

```
data(UPSep25)
condition1 <- '25fmol'
condition2 <- '10fmol'
qData <- Biobase::exprs(UPSep25)
samplesData <- Biobase::pData(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
diffAnaLimma(qData, samplesData, labels, condition1, condition2)
```

diffAnaSave	<i>Returns a MSnSet object with the results of the differential analysis performed with limma package.</i>
-------------	--

Description

This method returns a [MSnSet](#) object with the results of differential analysis.

Usage

```
diffAnaSave(obj, data, method = "limma", condition1, condition2,
  threshold_pVal = 1e-60, threshold_logFC = 0, fdr = 0,
  calibrationMethod = "pounds")
```

Arguments

obj	An object of class MSnSet .
data	The result of the differential analysis processed by diffAna
method	The method used for differential analysis. Available choices are : "limma", "Welch"
condition1	A vector containing the names (some values of the slot "Label" of <code>pData()</code>) of the first condition.
condition2	A vector containing the names (some values of the slot "Label" of <code>pData()</code>) of the second condition.
threshold_pVal	A float that indicates the threshold on p-value chosen to discriminate differential proteins.
threshold_logFC	A float that indicates the threshold on log(Fold Change) to discriminatedifferential proteins.
fdr	The FDR based on the values of <code>threshold_pVal</code> and <code>threshold_logFC</code>
calibrationMethod	The calibration method used to compute the calibration plot

Value

A MSnSet

Author(s)

Alexia Dorffer, Samuel Wieczorek

Examples

```
data(UPSep25)
condition1 <- '25fmol'
condition2 <- '10fmol'
limma <- wrapper.diffAnalimma(UPSep25, condition1, condition2)
obj <- diffAnaSave(UPSep25, limma, "limma", condition1, condition2)
```

diffAnaVolcanoplot *Volcanoplot of the differential analysis*

Description

Plots a volcano plot after the differential analysis. Typically, the log of Fold Change is represented on the X-axis and the log₁₀ of the p-value is drawn on the Y-axis. When the `threshold_pVal` and the `threshold_logFC` are set, two lines are drawn respectively on the y-axis and the X-axis to visually distinguish between differential and non differential data.

Usage

```
diffAnaVolcanoplot(logFC = NULL, pVal = NULL, threshold_pVal = 1e-60,
  threshold_logFC = 0, conditions = NULL)
```

Arguments

<code>logFC</code>	A vector of the log(fold change) values of the differential analysis.
<code>pVal</code>	A vector of the p-value values returned by the differential analysis.
<code>threshold_pVal</code>	A floating number which represents the p-value that separates differential and non-differential data.
<code>threshold_logFC</code>	A floating number which represents the log of the Fold Change that separates differential and non-differential data.
<code>conditions</code>	A list of the names of condition 1 and 2 used for the differential analysis.

Value

A volcano plot

Author(s)

Florence Combes, Samuel Wiczorek

Examples

```
data(UPSep25)
condition1 <- '25fmol'
condition2 <- '10fmol'
data <- wrapper.diffAnaLimma(UPSep25, condition1, condition2)
diffAnaVolcanoplot(data$logFC, data$P.Value)
```

diffAnaWelch	<i>Performs a differential analysis on a MSnSet object using the Welch t-test</i>
--------------	---

Description

Computes differential analysis on an [MSnSet](#) object, using the Welch t-test (`t.test{stats}`).

Usage

```
diffAnaWelch(qData, labels, condition1, condition2)
```

Arguments

qData	A dataframe that contains quantitative data.
labels	A vector of the conditions (labels) (one label per sample).
condition1	A vector containing the names of the conditions qData as condition 1
condition2	A vector containing the names of the conditions considered as condition 2

Value

A dataframe with two slots : P.Value (for the p-value) and logFC (the log of the Fold Change).

Author(s)

Florence Combes, Samuel Wiczorek

Examples

```
data(UPSep25)
condition1 <- '25fmol'
condition2 <- '10fmol'
qData <- Biobase::exprs(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
diffAnaWelch(qData, labels, condition1, condition2)
```

getIndicesConditions *Gets the conditions indices.*

Description

Returns a list for the two conditions where each slot is a vector of indices for the samples.

Usage

```
getIndicesConditions(labels, cond1, cond2)
```

Arguments

labels	A vector of strings containing the column "Label" of the pData().
cond1	A vector of Labels (a slot in the pData() table) for the condition 1.
cond2	A vector of Labels (a slot in the pData() table) for the condition 2.

Value

A list with two slots iCond1 and iCond2 containing respectively the indices of samples in the pData() table of the dataset.

Author(s)

Florence Combes, Samuel Wiczorek

Examples

```
data(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
getIndicesConditions(labels, "25fmol", "10fmol")
```

getIndicesOfLinesToRemove
Get the indices of the lines to delete, based on a prefix string

Description

This function returns the indice of the lines to delete, based on a prefix string

Usage

```
getIndicesOfLinesToRemove(obj, idLine2Delete = NULL, prefix = NULL)
```

Arguments

obj	An object of class MSnSet .
idLine2Delete	The name of the column that correspond to the data to filter
prefix	A character string that is the prefix to find in the data

Value

A vector of integers.

Author(s)

Samuel Wieczorek

Examples

```
data(UPSpep25)
getIndicesOfLinesToRemove(UPSpep25, "Potential.contaminant", prefix="+")
```

getNumberOf	<i>Number of lines with prefix</i>
-------------	------------------------------------

Description

Returns the number of lines, in a given column, where content matches the prefix.

Usage

```
getNumberOf(obj, name = NULL, prefix = NULL)
```

Arguments

obj	An object of class MSnSet .
name	The name of a column.
prefix	A string

Value

An integer

Author(s)

Samuel Wieczorek

Examples

```
data(UPSpep25)
getNumberOf(UPSpep25, "Potential.contaminant", "+")
```

getNumberOfEmptyLines *Returns the number of empty lines in the data*

Description

Returns the number of empty lines in a matrix.

Usage

```
getNumberOfEmptyLines(qData)
```

Arguments

qData A matrix corresponding to the quantitative data.

Value

An integer

Author(s)

Samuel Wieczorek

Examples

```
data(UPSsp25)
qData <- Biobase::exprs(UPSsp25)
getNumberOfEmptyLines(qData)
```

getPaletteForLabels *Palette for plots in DAPAR*

Description

Selects colors for the plots in DAPAR based on the different conditions in the dataset. The palette is derived from the brewer palette "Dark2" (see [RColorBrewer](#)).

Usage

```
getPaletteForLabels(labels)
```

Arguments

labels A vector of labels (strings).

Value

A palette designed for the data manipulated in DAPAR

Author(s)

Florence Combes, Samuel Wieczorek

Examples

```
data(UPSpep25)
labels <- Biobase::pData(UPSpep25)[,"Label"]
getPaletteForLabels(labels)
```

`getPaletteForReplicates`

Palette for plot the replicates in DAPAR

Description

Selects colors for the plots in DAPAR based on the replicates in the dataset. The palette is derived from the brewer palette "Dark2" (see [RColorBrewer](#)).

Usage

```
getPaletteForReplicates(nColors)
```

Arguments

`nColors` The desired number of colors

Value

A palette designed for the data manipulated in DAPAR

Author(s)

Samuel Wieczorek

Examples

```
data(UPSpep25)
n <- nrow(Biobase::pData(UPSpep25))
getPaletteForLabels(5)
```

getPourcentageOfMV *Percentage of missing values*

Description

Returns the percentage of missing values in the quantitative data (exprs()) table of the dataset).

Usage

```
getPourcentageOfMV(obj)
```

Arguments

obj An object of class [MSnSet](#).

Value

A floating number

Author(s)

Florence Combes, Samuel Wieczorek

Examples

```
data(UPSep25)
getPourcentageOfMV(UPSep25)
```

getProcessingInfo *Returns the contains of the slot processing of an object of class MSnSet*

Description

Returns the contains of the slot processing of an object of class [MSnSet](#).

Usage

```
getProcessingInfo(obj)
```

Arguments

obj An object (peptides) of class [MSnbase](#).

Value

The slot processing of obj@processingData

Author(s)

Samuel Wieczorek

Examples

```
data(UPSpep25)
getProcessingInfo(UPSpep25)
```

getProteinsStats	<i>computes the number of proteins that are only defined by specific peptides, shared peptides or a mixture of two.</i>
------------------	---

Description

This function computes the number of proteins that are only defined by specific peptides, shared peptides or a mixture of two.

Usage

```
getProteinsStats(matUnique, matShared)
```

Arguments

matUnique	The adjacency matrix with only specific peptides.
matShared	The adjacency matrix with both specific and shared peptides.

Value

A list

Author(s)

Samuel Wieczorek

Examples

```
data(UPSpep25)
protID <- "Protein.group.IDs"
MShared <- BuildAdjacencyMatrix(UPSpep25, protID, FALSE)
MUnique <- BuildAdjacencyMatrix(UPSpep25, protID, TRUE)
getProteinsStats(MUnique, MShared)
```

GraphPepProt	<i>Function to create a histogram that shows the repartition of peptides w.r.t. the proteins</i>
--------------	--

Description

Method to create a plot with proteins and peptides on a MSnSet object (peptides)

Usage

```
GraphPepProt(mat)
```

Arguments

mat An adjacency matrix.

Value

A histogram

Author(s)

Alexia Dorffer, Samuel Wieczorek

Examples

```
data(UPSspep25)
mat <- BuildAdjacencyMatrix(UPSspep25, "Protein.group.IDs")
GraphPepProt(mat)
```

heatmap.DAPAR	<i>This function is inspired from the function heatmap.2 that displays quantitative data in the <code>exprs()</code> table of an object of class <code>MSnSet</code>. For more information, please refer to the help of the <code>heatmap.2</code> function.</i>
---------------	--

Description

Heatmap inspired by the `heatmap.2` function.

Usage

```
heatmap.DAPAR(x, col = heat.colors(100), srtCol = NULL, labCol = NULL,
  labRow = NULL, key = TRUE, key.title = NULL, main = NULL,
  ylab = NULL)
```


Arguments

x	A dataframe that contains quantitative data.
col	colors used for the image. Defaults to heat colors (heat.colors).
srtCol	angle of column labels, in degrees from horizontal
labCol	character vectors with column labels to use.
labRow	character vectors with row labels to use.
key	logical indicating whether a color-key should be shown.
key.title	main title of the color key. If set to NA no title will be plotted.
main	main title; default to none.
ylab	y-axis title; default to none.

Value

A heatmap

Author(s)

Samuel Wieczorek

Examples

```
data(testWithoutNA)
qData <- Biobase::exprs(testWithoutNA)
heatmapD(qData)
```

heatmapD	<i>This function is a wrapper to heatmap.2 that displays quantitative data in the <code>exprs()</code> table of an object of class MSnSet</i>
----------	---

Description

Heatmap of the quantitative proteomic data of a [MSnSet](#) object

Usage

```
heatmapD(qData, distance = "euclidean", cluster = "average",
         dendro = FALSE)
```

Arguments

qData	A dataframe that contains quantitative data.
distance	The distance used by the clustering algorithm to compute the dendrogram. See <code>help(heatmap.2)</code>
cluster	the clustering algorithm used to build the dendrogram. See <code>help(heatmap.2)</code>
dendro	A boolean to indicate if the dendrogram has to be displayed

Value

A heatmap

Author(s)

Florence Combes, Samuel Wiczorek

Examples

```
data(testWithoutNA)
qData <- Biobase::exprs(testWithoutNA)
heatmapD(qData)
```

limmaCompleteTest *Computes a hierarchical differential analysis*

Description

This function is a limmaCompleteTest

Usage

```
limmaCompleteTest(qData, Conditions, RepBio, RepTech, Contrast = 1)
```

Arguments

qData	A matrix of quantitative data, without any missing values.
Conditions	A vector of factor which indicates the name of the biological condition for each replicate.
RepBio	A vector of factor which indicates the number of the bio rep for each replicate.
RepTech	A vector of factor which indicates the number of the tech rep for each replicate.
Contrast	Indicates if the test consists of the comparison of each biological condition versus each of the other ones (Contrast=1; for example H0:"C1=C2" vs H1:"C1!=C2", etc.) or each condition versus all others (Contrast=2; e.g. H0:"C1=(C2+C3)/2" vs H1:"C1!=(C2+C3)/2", etc. if there are three conditions).

Value

fdsfdgfdg

Author(s)

Quentin Giai-Gianetto

Examples

```
data(UPSep25)
obj <- wrapper.mvImputation(UPSep25, "QRILC")
condition1 <- '25fmol'
condition2 <- '10fmol'
qData <- Biobase::exprs(obj)
RepBio <- RepTech <- factor(1:6)
conds <- factor(c(rep(condition1, 3), (rep(condition2, 3))))
limma <- limmaCompleteTest(qData,conds,RepBio, RepTech)
```

MeanPeptides	<i>Compute the intensity of proteins as the mean of the intensities of their peptides.</i>
--------------	--

Description

This function computes the intensity of proteins as the mean of the intensities of their peptides.

Usage

```
MeanPeptides(matAdj, expr)
```

Arguments

matAdj	An adjacency matrix in which lines and columns correspond respectively to peptides and proteins.
expr	A matrix of intensities of peptides

Value

A matrix of intensities of proteins

Author(s)

Alexia Dorffer

Examples

```
data(UPSep25)
protID <- "Protein.group.IDs"
matAdj <- BuildAdjacencyMatrix(UPSep25, protID, FALSE)
MeanPeptides(matAdj, Biobase::exprs(UPSep25))
```

MeanPeptidesSpeedUp *Compute the intensity of proteins as the mean of the intensities of their peptides.*

Description

This function computes the intensity of proteins as the mean of the intensities of their peptides.

Usage

```
MeanPeptidesSpeedUp(matAdj, expr)
```

Arguments

matAdj An adjacency matrix in which lines and columns correspond respectively to peptides and proteins.

expr A matrix of intensities of peptides

Value

A matrix of intensities of proteins

Author(s)

Alexia Dorffer

Examples

```
data(UPSsep25)
protID <- "Protein.group.IDs"
matAdj <- BuildSparseAdjacencyMatrix(UPSsep25, protID, FALSE)
MeanPeptidesSpeedUp(matAdj, Biobase::exprs(UPSsep25))
```

mvFilter *Filter lines in the matrix of intensities w.r.t. some criteria*

Description

Filters the lines of `exprs()` table with conditions on the number of missing values. The user chooses the minimum amount of intensities that is acceptable and the filter delete lines that do not respect this condition. The condition may be on the whole line or condition by condition.

Usage

```
mvFilter(obj, type, th, processText = NULL)
```

Arguments

obj	An object of class MSnSet containing quantitative data.
type	Method used to choose the lines to delete. Values are : "none", "wholeMatrix", "allCond", "atLeastOneCond"
th	An integer value of the threshold
processText	A string to be included in the MSnSet object for log.

Details

The different methods are : "wholeMatrix": given a threshold th, only the lines that contain at least th values are kept. "allCond": given a threshold th, only the lines which contain at least th values for each of the conditions are kept. "atLeastOneCond": given a threshold th, only the lines that contain at least th values, and for at least one condition, are kept.

Value

An instance of class [MSnSet](#) that have been filtered.

Author(s)

Florence Combes, Samuel Wieczorek

Examples

```
data(UPSep25)
mvFilter(UPSep25, "wholeMatrix", 2)
```

mvFilterFromIndices *Filter lines in the matrix of intensities w.r.t. some criteria*

Description

Filters the lines of `exprs()` table with conditions on the number of missing values. The user chooses the minimum amount of intensities that is acceptable and the filter delete lines that do not respect this condition. The condition may be on the whole line or condition by condition.

Usage

```
mvFilterFromIndices(obj, keepThat = NULL, processText = NULL)
```

Arguments

obj	An object of class MSnSet containing quantitative data.
keepThat	A vector of integers which are the indices of lines to keep.
processText	A string to be included in the MSnSet object for log.

Details

The different methods are : "wholeMatrix": given a threshold th, only the lines that contain at least th values are kept. "allCond": given a threshold th, only the lines which contain at least th values for each of the conditions are kept. "atLeastOneCond": given a threshold th, only the lines that contain at least th values, and for at least one condition, are kept.

Value

An instance of class `MSnSet` that have been filtered.

Author(s)

Florence Combes, Samuel Wiczorek

Examples

```
data(UPSep25)
mvFilter(UPSep25, c(1:10))
```

mvFilterGetIndices *Filter lines in the matrix of intensities w.r.t. some criteria*

Description

Returns the indices of the lines of `exprs()` table to delete w.r.t. the conditions on the number of missing values. The user chooses the minimum amount of intensities that is acceptable and the filter delete lines that do not respect this condition. The condition may be on the whole line or condition by condition.

Usage

```
mvFilterGetIndices(obj, type, th)
```

Arguments

obj	An object of class <code>MSnSet</code> containing quantitative data.
type	Method used to choose the lines to delete. Values are : "none", "wholeMatrix", "allCond", "atLeastOneCond"
th	An integer value of the threshold

Details

The different methods are : "wholeMatrix": given a threshold th, only the lines that contain at least th values are kept. "allCond": given a threshold th, only the lines which contain at least th values for each of the conditions are kept. "atLeastOneCond": given a threshold th, only the lines that contain at least th values, and for at least one condition, are kept.

Value

An vector of indices that correspond to the lines to keep.

Author(s)

Florence Combes, Samuel Wieczorek

Examples

```
data(UPSsep25)
mvFilterGetIndices(UPSsep25, "wholeMatrix", 2)
```

mvHisto

Histogram of missing values

Description

This method plots a histogram of missing values.

Usage

```
mvHisto(qData, samplesData, labels, indLegend = "auto", showValues = FALSE)
```

Arguments

qData	A dataframe that contains quantitative data.
samplesData	A dataframe where lines correspond to samples and columns to the meta-data for those samples.
labels	A vector of the conditions (labels) (one label per sample).
indLegend	The indices of the column name's in pData() tab
showValues	A logical that indicates wether numeric values should be drawn above the bars.

Value

A histogram

Author(s)

Florence Combes, Samuel Wieczorek

Examples

```
data(UPSsep25)
qData <- Biobase::exprs(UPSsep25)
samplesData <- Biobase::pData(UPSsep25)
labels <- Biobase::pData(UPSsep25)[,"Label"]
mvHisto(qData, samplesData, labels, indLegend="auto", showValues=TRUE)
```

`mvImage`*Heatmap of missing values*

Description

Plots a heatmap of the quantitative data. Each column represent one of the conditions in the object of class `MsnSet` and the color is proportional to the mean of intensity for each line of the dataset. The lines have been sorted in order to visualize easily the different number of missing values. A white square is plotted for missing values.

Usage

```
mvImage(qData, labels)
```

Arguments

`qData` A dataframe that contains quantitative data.
`labels` A vector of the conditions (labels) (one label per sample).

Value

A heatmap

Author(s)

Samuel Wiczorek, Thomas Burger

Examples

```
data(UPSep25)
qData <- Biobase::exprs(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
mvImage(qData, labels)
```

`mvImputation`*Missing values imputation from a matrix*

Description

This method is a wrapper to the `imputeLCMD` package adapted to a matrix.

Usage

```
mvImputation(qData, method)
```


Arguments

qData A dataframe that contains quantitative data.
 method The imputation method to be used. Choices are QRILC, KNN, BPCA and MLE.

Value

The matrix imputed

Author(s)

Samuel Wieczorek

Examples

```
data(UPSpep25)
qData <- Biobase::exprs(UPSpep25)
mvImputation(qData, "QRILC")
```

mvPerLinesHisto *Bar plot of missing values per lines*

Description

This method plots a bar plot which represents the distribution of the number of missing values (NA) per lines (ie proteins).

Usage

```
mvPerLinesHisto(qData, samplesData, indLegend = "auto", showValues = FALSE)
```

Arguments

qData A dataframe that contains the data to plot.
 samplesData A dataframe which contains informations about the replicates.
 indLegend The indice of the column name's in pData() tab
 showValues A logical that indicates wether numeric values should be drawn above the bars.

Value

A bar plot

Author(s)

Florence Combes, Samuel Wieczorek

Examples

```
data(UPSsep25)
qData <- Biobase::exprs(UPSsep25)
samplesData <- Biobase::pData(UPSsep25)
mvPerLinesHisto(qData, samplesData)
```

mvPerLinesHistoPerCondition

Bar plot of missing values per lines and per condition

Description

This method plots a bar plot which represents the distribution of the number of missing values (NA) per lines (ie proteins) and per conditions.

Usage

```
mvPerLinesHistoPerCondition(qData, samplesData, indLegend = "auto",
  showValues = FALSE)
```

Arguments

qData	A dataframe that contains quantitative data.
samplesData	A dataframe where lines correspond to samples and columns to the meta-data for those samples.
indLegend	The indice of the column name's in pData() tab
showValues	A logical that indicates wether numeric values should be drawn above the bars.

Value

A bar plot

Author(s)

Samuel Wiczorek

Examples

```
data(UPSsep25)
qData <- Biobase::exprs(UPSsep25)
samplesData <- Biobase::pData(UPSsep25)
mvPerLinesHistoPerCondition(qData, samplesData)
```

mvTypePlot

*Distribution of missing values with respect to intensity values***Description**

This method plots a scatter plot which represents the distribution of missing values. The colors correspond to the different conditions (slot Label in in the dataset of class `MsnSet`). The x-axis represent the mean of intensity for one condition and one entity in the dataset (i. e. a protein) whereas the y-axis count the number of missing values for this entity and the considered condition. The data have been jittered for an easier vizualisation.

Usage

```
mvTypePlot(qData, labels, threshold = 0)
```

Arguments

qData	A dataframe that contains quantitative data.
labels	A vector of the conditions (labels) (one label per sample).
threshold	An integer for the intensity that delimits MNAR and MCAR missing values.

Value

A scatter plot

Author(s)

Florence Combes, Samuel Wiczorek

Examples

```
data(UPSsep25)
qData <- Biobase::exprs(UPSsep25)
labels <- Biobase::pData(UPSsep25)[,"Label"]
mvTypePlot(qData, labels, threshold=0)
```

normalized

*Normalisation***Description**

Provides several methods to normalize data from a matrix. They are organized in four main families : Strong Rescaling, Median Centering, Mean Centering, Mean CenteringScaling. For the first family, two sub-categories are available : the sum by columns and the quantiles method. For the three other families, two categories are available : "Overall" which means that the value for each protein (ie line in the expression data tab) is computed over all the samples ; "within conditions" which means that the value for each protein (ie line in the matrix) is computed condition by condition.

Usage

```
normalized(qData, labels, family, method)
```

Arguments

qData	A dataframe that contains quantitative data.
labels	A vector of strings containing the column "Label" of the pData().
family	One of the following : Global Rescaling, Median Centering, Mean Centering, Mean Centering Scaling.
method	"Overall" or "within conditions".

Value

A matrix normalized

Author(s)

Florence Combes, Samuel Wiczorek

Examples

```
data(UPSsep25)
qData <- Biobase::exprs(UPSsep25)
labels <- Biobase::pData(UPSsep25)[,"Label"]
normalized(qData, labels, "Median Centering", "within conditions")
```

pepAgregate

Function agregate peptides to proteins

Description

Method to agregate with a method peptides to proteins on a MSnSet object (peptides)

Usage

```
pepAgregate(obj.pep, protID, method = "sum", matAdj = NULL, n = NULL)
```

Arguments

obj.pep	An object (peptides) of class MSnbase .
protID	The name of proteins ID column
method	The method used to aggregate the peptides into proteins. Values are "sum", "mean" or "sum on top n" : do the sum / mean of intensity on all peptides belonging to proteins. Default is "sum"
matAdj	An adjacency matrix
n	The number of peptides considered for the aggregation.

Value

An object of class [MSnbase](#) with proteins

Author(s)

Alexia Dorffer, Samuel Wieczorek

Examples

```
data(UPSspep25)
protID <- "Protein.group.IDs"
mat <- BuildAdjacencyMatrix(UPSspep25, protID, TRUE)
pepAggregate(UPSspep25, protID, "sum", mat)
```

pepAggregateSpeedUp *Function aggregate peptides to proteins*

Description

Method to aggregate with a method peptides to proteins on a MSnSet object (peptides)

Usage

```
pepAggregateSpeedUp(obj.pep, protID, method = "sum overall", matAdj = NULL,
  n = NULL)
```

Arguments

- obj.pep An object (peptides) of class [MSnbase](#).
- protID The name of proteins ID column
- method The method used to aggregate the peptides into proteins. Values are "sum", "mean" or "sum on top n" : do the sum / mean of intensity on all peptides belonging to proteins. Default is "sum"
- matAdj An adjacency matrix
- n The number of peptides considered for the aggregation.

Value

An object of class [MSnbase](#) with proteins

Author(s)

Alexia Dorffer, Samuel Wieczorek

Examples

```
data(UPSep25)
protID <- "Protein.group.IDs"
mat <- BuildSparseAdjacencyMatrix(UPSep25, protID, TRUE)
pepAggregateSpeedUp(UPSep25, protID, "sum overall", mat)
```

proportionConRev *Barplot of proportion of contaminants and reverse*

Description

Plots a barplot of proportion of contaminants and reverse

Usage

```
proportionConRev(obj, idContaminants = NULL, prefixContaminants = NULL,
  idReverse = NULL, prefixReverse = NULL)
```

Arguments

obj An object of class `MsnSet`.
idContaminants The name of a column of Contaminants
prefixContaminants The prefix to identify contaminants
idReverse The name of a column of Reverse
prefixReverse The prefix to identify Reverse

Value

A barplot

Author(s)

Samuel Wieczorek

Examples

```
data(UPSep25)
pref <- "+"
proportionConRev(UPSep25, "Potential.contaminant", pref, "Reverse", pref)
```

removeLines	<i>Removes lines in the dataset based on a prefix string.</i>
-------------	---

Description

This function removes lines in the dataset based on a prefix string.

Usage

```
removeLines(obj, idLine2Delete = NULL, prefix = NULL)
```

Arguments

obj	An object of class MSnSet .
idLine2Delete	The name of the column that correspond to the data to filter
prefix	A character string that is the prefix to find in the data

Value

An object of class [MSnSet](#).

Author(s)

Samuel Wieczorek

Examples

```
data(UPSep25)
removeLines(UPSep25, "Potential.contaminant")
removeLines(UPSep25, "Reverse")
```

SumPeptides	<i>Compute the intensity of proteins with the sum of the intensities of their peptides.</i>
-------------	---

Description

This function computes the intensity of proteins based on the sum of the intensities of their peptides.

Usage

```
SumPeptides(matAdj, expr)
```

Arguments

matAdj	An adjacency matrix in which lines and columns correspond respectively to peptides and proteins.
expr	A matrix of intensities of peptides

Value

A matrix of intensities of proteins

Author(s)

Alexia Dorffer

Examples

```
data(UPSsep25)
protID <- "Protein.group.IDs"
M <- BuildAdjacencyMatrix(UPSsep25, protID, FALSE)
SumPeptides(M, Biobase::exprs(UPSsep25))
```

SumPeptidesSpeedUp *Compute the intensity of proteins with the sum of the intensities of their peptides.*

Description

This function computes the intensity of proteins based on the sum of the intensities of their peptides.

Usage

```
SumPeptidesSpeedUp(matAdj, expr)
```

Arguments

matAdj	An adjacency matrix in which lines and columns correspond respectively to peptides and proteins.
expr	A matrix of intensities of peptides

Value

A matrix of intensities of proteins

Author(s)

Alexia Dorffer

Examples

```

data(UPSep25)
protID <- "Protein.group.IDs"
M <- BuildSparseAdjacencyMatrix(UPSep25, protID, FALSE)
SumPeptidesSpeedUp(M, Biobase::exprs(UPSep25))

```

test	<i>Test dataset</i>
------	---------------------

Description

Partial (small) dataset for unit tests containing missing values.

Format

An object of class [MSnSet](#)

testWithoutNA	<i>Test dataset</i>
---------------	---------------------

Description

Partial (small) dataset for unit tests without any missing values.

Format

An object of class [MSnSet](#)

topMaxUsingPartialSortIndices	<i>Function to return the indices of the n higher values in the vector</i>
-------------------------------	--

Description

Method to return the indices of the n higher values in the vector

Usage

```
topMaxUsingPartialSortIndices(x, n)
```

Arguments

x	A vector of numeric values
n	The number of values to be returned

Value

A vector of the indices of the n highest values

Author(s)

Alexia Dorffer

Examples

```
topMaxUsingPartialSortIndices(c(1:10), 3)
```

TopnPeptides	<i>Compute the intensity of proteins as the sum of the intensities of their n best peptides.</i>
--------------	--

Description

This function computes the intensity of proteins as the sum of the intensities of their n best peptides.

Usage

```
TopnPeptides(matAdj, expr, n)
```

Arguments

matAdj	An adjacency matrix in which lines and columns correspond respectively to peptides and proteins.
expr	A matrix of intensities of peptides
n	The maximum number of peptides used to aggregate a protein.

Value

A matrix of intensities of proteins

Author(s)

Alexia Dorffer

Examples

```
data(UPSsep25)
protID <- "Protein.group.IDs"
matAdj <- BuildAdjacencyMatrix(UPSsep25, protID, FALSE)
TopnPeptides(matAdj, Biobase::exprs(UPSsep25), 3)
```

UPSp25

UPSp25 dataset

Description

This dataset is the final outcome of a quantitative mass spectrometry-based proteomic analysis of two samples containing different concentrations of 48 human proteins (UPS1 standard from Sigma-Aldrich) within a constant yeast background (see Giai Gianetto et al. (2016) for details). It contains the abundance values of the different human and yeast peptides identified and quantified in these two conditions. The two conditions represent the measured abundances of peptides when respectively 25fmol and 10fmol of UPS1 human proteins were mixed with the yeast extract before mass spectrometry analyses. Three technical replicates were acquired for each condition.

To identify and quantify peptides, spectra were searched using MaxQuant (version 1.5.1.2) against the Uniprot database, the UPS database and the frequently observed contaminants database. Maximum false discovery rates were set to 0.01 by employing a reverse database strategy.

The dataset is either available as a CSV file (see `inst/extdata/UPSp25.txt`), or as a [MSnSet](#) structure (UPSp25). In the latter case, the quantitative data are those of the raw intensities.

Usage

```
data(UPSp25)
```

Format

An object of class [MSnSet](#) related to peptide quantification. It contains 6 samples divided into two conditions (25fmol and 10fmol) and 13918 peptides.

The data frame `exprs(UPSp25)` contains six columns that are the quantitation of peptides for the six replicates.

The data frame `fData(UPSp25)` contains the meta data about the peptides.

The data frame `pData(UPSp25)` contains the experimental design and gives few informations about the samples.

Value

An object of class [MSnSet](#).

References

Cox J., Hein M.Y., Lubner C.A., Paron I., Nagaraj N., Mann M. Accurate proteome-wide label-free quantification by delayed normalization and maximal peptide ratio extraction, termed MaxLFQ. *Mol Cell Proteomics*. 2014 Sep, 13(9):2513-26.

Giai Gianetto, Q., Combes, F., Ramus, C., Bruley, C., Coute, Y., Burger, T. (2016). Calibration plot for proteomics: A graphical tool to visually check the assumptions underlying FDR control in quantitative experiments. *Proteomics*, 16(1), 29-32.

varianceDistD	<i>Distribution of variance of proteins</i>
---------------	---

Description

Builds a densityplot of the variance of entities in the `exprs()` table of a object. The variance is calculated for each condition (Label) present in the dataset (see the slot 'Label' in the `pData()` table)

Usage

```
varianceDistD(qData, labels = NULL)
```

Arguments

<code>qData</code>	A dataframe that contains quantitative data.
<code>labels</code>	A vector of the conditions (labels) (one label per sample).

Value

A density plot

Author(s)

Florence Combes, Samuel Wieczorek

See Also

[densityPlotD](#).

Examples

```
data(UPSsep25)
labels <- Biobase::pData(UPSsep25)[,"Label"]
varianceDistD(UPSsep25)
```

wrapper.boxPlotD *Wrapper to the boxplotD function on an object [MSnSet](#)*

Description

This function is a wrapper for using the boxPlotD function with objects of class [MSnSet](#)

Usage

```
wrapper.boxPlotD(obj, dataForXAxis = "Label", group2Color = "Condition")
```

Arguments

obj	An object of class MSnSet .
dataForXAxis	A vector of strings containing the names of columns in pData() to print labels on X-axis (Default is "Label").
group2Color	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

Value

A boxplot

Author(s)

Florence Combes, Samuel Wiczorek

See Also

[wrapper.densityPlotD](#)

Examples

```
data(UPSep25)
types <- c("Label", "Analyt.Rep")
wrapper.boxPlotD(UPSep25, types)
```

`wrapper.compareNormalizationD`*Builds a plot from a dataframe*

Description

Wrapper to the function that plot to compare the quantitative proteomics data before and after normalization

Usage

```
wrapper.compareNormalizationD(objBefore, objAfter, labelsForLegend = NULL,  
  indData2Show = NULL, group2Color = "Condition")
```

Arguments

<code>objBefore</code>	A dataframe that contains quantitative data before normalization.
<code>objAfter</code>	A dataframe that contains quantitative data after normalization.
<code>labelsForLegend</code>	A vector of the conditions (labels) (one label per sample).
<code>indData2Show</code>	A vector of the indices of the columns to show in the plot. The indices are those of indices of the columns in the data.frame <code>qDataBefore</code> .
<code>group2Color</code>	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

Value

A plot

Author(s)

Samuel Wieczorek

Examples

```
data(UPSsep25)  
labels <- Biobase::pData(UPSsep25)[,"Label"]  
objAfter <- wrapper.normalizeD(UPSsep25, "Median Centering",  
  "within conditions")  
wrapper.compareNormalizationD(UPSsep25, objAfter, labels)
```

wrapper.corrMatrixD *Displays a correlation matrix of the quantitative data of the exprs() table*

Description

Builds a correlation matrix based on a [MSnSet](#) object.

Usage

```
wrapper.corrMatrixD(obj, rate = 5)
```

Arguments

obj An object of class [MSnSet](#).
rate A float that defines the gradient of colors.

Value

A colored correlation matrix

Author(s)

Alexia Dorffer

Examples

```
data(UPSep25)  
wrapper.corrMatrixD(UPSep25)
```

wrapper.densityPlotD *Builds a densityplot from an object of class [MSnSet](#)*

Description

This function is a wrapper for using the densityPlotD function with objects of class [MSnSet](#)

Usage

```
wrapper.densityPlotD(obj, labelsForLegend = NULL, indData2Show = NULL,  
                  group2Color = "Condition")
```

Arguments

obj	An object of class MSnSet .
labelsForLegend	A vector of labels to show in densityplot.
indData2Show	A vector of the indices of the columns to show in the plot. The indices are those of indices of the columns in the data frame qDataBefore in the density plot.
group2Color	A string that indicates how to color the replicates: one color per condition (value "Condition") or one color per replicate (value "Replicate"). Default value is by Condition.

Value

A density plot

Author(s)

Alexia Dorffer

See Also

[wrapper.boxPlotD](#), [wrapper.varianceDistD](#)

Examples

```
data(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
wrapper.densityPlotD(UPSep25, labels)
```

wrapper.diffAnaLimma *Performs differential analysis on an MSnSet object, calling the limma package functions*

Description

Method to perform differential analysis on a [MSnSet](#) object (calls the limma package function).

Usage

```
wrapper.diffAnaLimma(obj, condition1, condition2)
```

Arguments

obj	An object of class MSnSet .
condition1	A vector that contains the names of the conditions considered as condition 1.
condition2	A vector that contains the names of the conditions considered as condition 2.

Value

A dataframe as returned by the limma package

Author(s)

Alexia Dorffer

Examples

```
data(UPSep25)
condition1 <- '25fmol'
condition2 <- '10fmol'
wrapper.diffAnaLimma(UPSep25, condition1, condition2)
```

wrapper.diffAnaWelch *Performs a differential analysis on a [MSnSet](#) object using the Welch t-test*

Description

Computes differential analysis on a [MSnSet](#) object, using the Welch t-test (`t.test{stats}`).

Usage

```
wrapper.diffAnaWelch(obj, condition1, condition2)
```

Arguments

obj	An object of class MSnSet .
condition1	A vector containing the names of the conditions considered as condition 1.
condition2	A vector containing the names of the conditions considered as condition 2.

Value

A dataframe with two slots : P.Value (for the p-value) and logFC (the log of the Fold Change).

Author(s)

Alexia Dorffer

Examples

```
data(UPSep25)
condition1 <- '25fmol'
condition2 <- '10fmol'
wrapper.diffAnaWelch(UPSep25, condition1, condition2)
```

wrapper.heatmapD	<i>This function is a wrapper to heatmap.2 that displays quantitative data in the <code>exprs()</code> table of an object of class MSnSet</i>
------------------	---

Description

Builds a heatmap of the quantitative proteomic data of a [MSnSet](#) object.

Usage

```
wrapper.heatmapD(obj, distance = "euclidean", cluster = "average",
  dendro = FALSE)
```

Arguments

obj	An object of class MSnSet .
distance	The distance used by the clustering algorithm to compute the dendrogram. See <code>help(heatmap.2)</code> .
cluster	the clustering algorithm used to build the dendrogram. See <code>help(heatmap.2)</code>
dendro	A boolean to indicate if the dendrogram has to be displayed

Value

A heatmap

Author(s)

Alexia Dorffer

Examples

```
data(testWithoutNA)
wrapper.heatmapD(testWithoutNA)
```

wrapper.mvHisto	<i>Histogram of missing values from a MSnSet object</i>
-----------------	---

Description

This method plots from a [MSnSet](#) object a histogram of missing values.

Usage

```
wrapper.mvHisto(obj, indLegend = "auto", showValues = FALSE)
```

Arguments

obj	An object of class MSnSet .
indLegend	The indices of the column name's in pData() tab.
showValues	A logical that indicates wether numeric values should be drawn above the bars.

Value

A histogram

Author(s)

Alexia Dorffer

Examples

```
data(UPSep25)
wrapper.mvHisto(UPSep25, showValues=TRUE)
```

wrapper.mvImage *Heatmap of missing values from a [MSnSet](#) object*

Description

Plots a heatmap of the quantitative data. Each column represent one of the conditions in the object of class [MSnSet](#) and the color is proportional to the mean of intensity for each line of the dataset. The lines have been sorted in order to vizualize easily the different number of missing values. A white square is plotted for missing values.

Usage

```
wrapper.mvImage(obj)
```

Arguments

obj	An object of class MSnSet .
-----	---

Value

A heatmap

Author(s)

Alexia Dorffer

Examples

```
data(UPSep25)
wrapper.mvImage(UPSep25)
```

wrapper.mvImputation *Missing values imputation from a MSnSet object*

Description

This method is a wrapper to the imputeLCMD package adapted to objects of class [MSnSet](#).

Usage

```
wrapper.mvImputation(obj, method)
```

Arguments

obj An object of class [MSnSet](#).
method The imputation method to be used. Choices are QRILC, KNN, BPCA and MLE.

Value

The object obj which has been imputed

Author(s)

Alexia Dorffer

Examples

```
data(UPSpep25)
wrapper.mvImputation(UPSpep25, "QRILC")
```

wrapper.mvPerLinesHisto
Histogram of missing values per lines from an object MSnSet

Description

This method is a wrapper to plots from a [MSnSet](#) object a histogram which represents the distribution of the number of missing values (NA) per lines (ie proteins).

Usage

```
wrapper.mvPerLinesHisto(obj, indLegend = "auto", showValues = FALSE)
```

Arguments

obj An object of class [MSnSet](#).
indLegend The indice of the column name's in pData() tab .
showValues A logical that indicates wether numeric values should be drawn above the bars.

Value

A histogram

Author(s)

Alexia Dorffer

Examples

```
data(UPSpep25)
wrapper.mvPerLinesHisto(UPSpep25)
```

```
wrapper.mvPerLinesHistoPerCondition
```

Bar plot of missing values per lines and per conditions from an object
[MSnSet](#)

Description

This method is a wrapper to plots from a [MSnSet](#) object a bar plot which represents the distribution of the number of missing values (NA) per lines (ie proteins) and per conditions.

Usage

```
wrapper.mvPerLinesHistoPerCondition(obj, indLegend = "auto",
  showValues = FALSE)
```

Arguments

obj	An object of class MSnSet .
indLegend	The indice of the column name's in pData() tab .
showValues	A logical that indicates wether numeric values should be drawn above the bars.

Value

A bar plot

Author(s)

Samuel Wiczorek

Examples

```
data(UPSpep25)
wrapper.mvPerLinesHistoPerCondition(UPSpep25)
```

wrapper.mvTypePlot *Distribution of missing values with respect to intensity values from a MSnSet object*

Description

This method plots a scatter plot which represents the distribution of missing values. The colors correspond to the different conditions (slot Label in in the dataset of class `MSnSet`). The x-axis represent the mean of intensity for one condition and one entity in the dataset (i. e. a protein) whereas the y-axis count the number of missing values for this entity and the considered condition. The data have been jittered for an easier vizualisation.

Usage

```
wrapper.mvTypePlot(obj, threshold = 0)
```

Arguments

obj An object of class `MSnSet`.
threshold An integer for the intensity that delimits MNAR and MCAR missing values.

Value

A scatter plot

Author(s)

Florence Combes, Samuel Wieczorek

Examples

```
data(UPSep25)  
wrapper.mvTypePlot(UPSep25)
```

wrapper.normalized *Normalisation*

Description

Provides several methods to normalize quantitative data from a `MSnSet` object. They are organized in four main families : Strong Rescaling, Median Centering, Mean Centering, Mean CenteringScaling. For the first family, two sub-categories are available : the sum by columns and the quantiles method. For the three other families, two categories are available : "Overall" which means that the value for each protein (ie line in the expression data tab) is computed over all the samples ; "within conditions" which means that the value for each protein (ie line in the `exprs()` data tab) is computed condition by condition.

Usage

```
wrapper.normalized(obj, family, method)
```

Arguments

obj	An object of class MSnSet .
family	One of the following : Global Rescaling, Median Centering, Mean Centering, Mean Centering Scaling.
method	"Overall" or "within conditions".

Value

An instance of class [MSnSet](#) where the quantitative data in the `exprs()` tab has been normalized.

Author(s)

Alexia Dorffer

Examples

```
data(UPSep25)
wrapper.normalized(UPSep25, "Median Centering", "within conditions")
```

`wrapper.varianceDistD` *Distribution of variance of proteins*

Description

Builds a densityplot of the variance of entities in the `exprs()` table of an object [MSnSet](#). The variance is calculated for each condition (Label) present in the dataset (see the slot 'Label' in the `pData()` table).

Usage

```
wrapper.varianceDistD(obj)
```

Arguments

obj	An object of class MSnSet .
-----	---

Value

A density plot

Author(s)

Alexia Dorffer

See Also

[wrapper.densityPlotD](#)

Examples

```
data(UPSep25)
wrapper.varianceDistD(UPSep25)
```

wrapperCalibrationPlot

Performs a calibration plot on an [MSnSet](#) object, calling the cp4p package functions.

Description

This function is a wrapper to the calibration.plot method of the cp4p package for use with [MSnSet](#) objects.

Usage

```
wrapperCalibrationPlot(vPVal, pi0Method = "pounds")
```

Arguments

vPVal	A dataframe that contains quantitative data.
pi0Method	A vector of the conditions (labels) (one label per sample).

Value

A plot

Author(s)

Samuel Wieczorek

Examples

```
data(UPSep25)
condition1 <- '25fmol'
condition2 <- '10fmol'
qData <- Biobase::exprs(UPSep25)
labels <- Biobase::pData(UPSep25)[,"Label"]
diffAnaWelch(qData, labels, condition1, condition2)
```

writeMSnsetToExcel *This function exports a `MSnSet` object to a Excel file.*

Description

This function exports a `MSnSet` data object to a Excel file. Each of the three data.frames in the `MSnSet` object (ie experimental data, phenoData and metaData are respectively integrated into separate sheets in the Excel file).

Usage

```
writeMSnsetToExcel(obj, filename, id)
```

Arguments

obj	An object of class <code>MSnSet</code> .
filename	A character string for the name of the Excel file.
id	An integer to select in the fdata frame which column has to be used as an index.

Value

A Excel file

Author(s)

Samuel Wieczorek

Examples

```
data(UPSep25)
writeMSnsetToExcel(UPSep25, "foo", 1)
```

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