Package 'scATAC.Explorer'

June 27, 2024

Title A Collection of Single-cell ATAC Sequencing Datasets and Corresponding Metadata

Version 1.11.0

Description

This package provides a tool to search and download a collection of publicly available single cell ATAC-seq datasets and their metadata. scATAC-Explorer aims to act as a single point of entry for users looking to study single cell ATAC-seq data. Users can quickly search available datasets using the metadata table and download datasets of interest for immediate analysis within R.

License Artistic-2.0 **Encoding UTF-8** LazyData FALSE **Roxygen** list(markdown = TRUE) RoxygenNote 7.1.1 VignetteBuilder knitr **Suggests** BiocStyle, knitr, rmarkdown, testthat (>= 3.0.0) Imports methods, Matrix **Depends** R (>= 4.1), SingleCellExperiment, BiocFileCache, data.table, utils, S4Vectors biocViews SingleCellData, SequencingData, ExpressionData, GEO, Tissue, Genome, PackageTypeData BugReports https://github.com/shooshtarilab/scATACseq/issues Config/testthat/edition 3 git_url https://git.bioconductor.org/packages/scATAC.Explorer git_branch devel git_last_commit 223d242 git_last_commit_date 2024-04-30 Repository Bioconductor 3.20 Date/Publication 2024-06-27

2 queryATAC

```
Author Arrian Gibson-Khademi [aut, cre],
Erik Christensen [aut],
Parisa Shooshtari [aut]
```

Maintainer Arrian Gibson-Khademi <agibsonk@uwo.ca>

Contents

```
queryATAC 2
saveATAC 4

Index 5

queryATAC A function to query scATAC-seq datasets available in this package
```

Description

This function allows you to search and subset included scATAC-seq datasets. A named list of scATAC-seq_data objects matching the provided options will be returned. Some included datasets are represented using multiple matrices. Each matrix will be a seperate named object within the list. The returned list is named by matrix allow easy identification of data. If queryATAC is called without any options it will retrieve all available datasets in sparse matrix format. This should only be done on machines with a large amount of ram (>64gb) because some datasets are quite large. In most cases it is recommended to instead filter databases with some criteria.

Usage

```
queryATAC(
  accession = NULL,
  author = NULL,
  journal = NULL,
  year = NULL,
  pmid = NULL,
  sequence_tech = NULL,
  score_type = NULL,
  has_cluster_annotation = NULL,
  has_cell_type_annotation = NULL,
  organism = NULL,
  genome_build = NULL,
  broad_cell_category = NULL,
  tissue_cell_type = NULL,
  disease = NULL,
 metadata_only = FALSE,
  sparse = TRUE
)
```

queryATAC 3

Arguments

accession Search by geo accession number. Good for returning individual datasets author Search by the author who published the dataset Search by the journal the dataset was published in. journal Search by exact year or year ranges with '<', '>', or '-'. For example, you can year return datasets newer than 2013 with '>2013' Search by Pubmed ID associated with the study. Good for returning individual pmid datasets sequence_tech Search by sequencing technology used to sample the cells. Search by type of score (TPM, FPKM, raw count) score_type has_cluster_annotation Return only those datasets that have clustering results available, or only those without (TRUE/FALSE) has_cell_type_annotation Return only those datasets that have cell-type annotations available, or only those without annotations (TRUE/FALSE) Search by source organism used in the study, for example human or mouse. organism Return datasets built only using specified genome build (ex. hg19) genome_build broad_cell_category Return datasets based on broad cell categories (ex. Hematopoetic cells). To view all cell categories available, explore the metadata table tissue_cell_type Return datasets based on tissue or cell types sampled (ex. PBMCs, Bone marrow, Oligodendrocytes) disease Return datasets based on sampled disease (ex. carcinoma, leukemia, diabetes) Return rows of metadata instead of actual datasets. Useful for exploring what metadata_only data is available without actually downloading data. Defaults to FALSE Return expression as a sparse matrix. Reccomended to use sparse format, as sparse

Value

A list containing a table of metadata or one or more SingleCellExperiment objects

dense formats tend to be excessively large.

Examples

4 saveATAC

saveATAC	A function to save a scATAC-seq dataset stored in a SingleCellExper- iment

Description

This function allows you to save the counts, peaks, cell ID's/barcodes, and any cell clustering data to disk in csv format. It takes two options: an object to save and a directory to save in. Multiple files will be created in the provided output directory, one for each type of data available in the scATAC_data object (counts, cell ID/Barcode, peak regions, cell type/cluster annotations).

Usage

```
saveATAC(object, outdir)
```

Arguments

object The SingleCellExperiment object to be written to disk, this should be an indi-

vidual dataset returned by queryATAC.

outdir The directory to save the data in, the directory should not exist yet.

Value

Nothing

Examples

```
# Retrieve a previously identified dataset (see queryATAC) and save it to disk
res <- queryATAC(accession = 'GSE89362')[[1]]
saveATAC(res, output_directory_name)</pre>
```

Index

```
* scATAC-seq
saveATAC, 4
* tumour
queryATAC, 2
queryATAC, 2
saveATAC, 4
```