

Package ‘maftools’

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Type Package

Title Summarize, Analyze and Visualize MAF Files

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Description Analyze and visualize Mutation Annotation Format (MAF) files from large scale sequencing studies. This package provides various functions to perform most commonly used analyses in cancer genomics and to create feature rich customizable visualizations with minimal effort.

URL <https://github.com/PoisonAlien/maftools>

BugReports <https://github.com/PoisonAlien/maftools/issues>

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LazyData TRUE

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survival

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annovarToMaf	<i>Converts annovar annotations into MAF.</i>
--------------	---

Description

Converts variant annotations from Annovar into a basic MAF.

Usage

```
annovarToMaf(
  annovar,
  Center = NULL,
  refBuild = "hg19",
  tsbCol = NULL,
  table = "refGene",
  ens2hugo = TRUE,
  basename = NULL,
  sep = "\t",
  MAFobj = FALSE,
  sampleAnno = NULL
)
```

Arguments

annovar	input annovar annotation file. Can be vector of multiple files.
Center	Center field in MAF file will be filled with this value. Default NA.
refBuild	NCBI_Build field in MAF file will be filled with this value. Default hg19.
tsbCol	column name containing Tumor_Sample_Barcode or sample names in input file.
table	reference table used for gene-based annotations. Can be 'ensGene' or 'refGene'. Default 'refGene'
ens2hugo	If 'table' is 'ensGene', setting this argument to 'TRUE' converts all ensemble IDs to hugo symbols.
basename	If provided writes resulting MAF file to an output file.
sep	field separator for input file. Default tab separated.
MAFobj	If TRUE, returns results as an MAF object.

sampleAnno annotations associated with each sample/Tumor_Sample_Barcode in input annovar file. If provided it will be included in MAF object. Could be a text file or a data.frame. Ideally annotation would contain clinical data, survival information and other necessary features associated with samples. Default NULL.

Details

Annovar is one of the most widely used Variant Annotation tools in Genomics. Annovar output is generally in a tabular format with various annotation columns. This function converts such annovar output files into MAF. This function requires that annovar was run with gene based annotation as a first operation, before including any filter or region based annotations. Please be aware that this function performs no transcript prioritization.

e.g. `table_annovar.pl example/ex1.avinput humandb/ -buildver hg19 -out myanno -remove -protocol (refGene),cytoBand,dbnsfp30a -operation (g),r,f -nastring NA`

This function mainly uses gene based annotations for processing, rest of the annotation columns from input file will be attached to the end of the resulting MAF.

Value

MAF table.

References

Wang, K., Li, M. & Hakonarson, H. ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Res* 38, e164 (2010).

Examples

```
var.annovar <- system.file("extdata", "variants.hg19_multianno.txt", package = "maftools")
var.annovar.maf <- annovarToMaf(annovar = var.annovar, Center = 'CSI-NUS', refBuild = 'hg19',
tsbCol = 'Tumor_Sample_Barcode', table = 'ensGene')
```

clinicalEnrichment *Performs mutational enrichment analysis for a given clinical feature.*

Description

Performs pairwise and groupwise fisher exact tests to find differentially enriched genes for every factor within a clinical feature.

Usage

```
clinicalEnrichment(
  maf,
  clinicalFeature = NULL,
  annotationDat = NULL,
  minMut = 5,
  useCNV = TRUE
)
```

Arguments

maf	MAF object
clinicalFeature	columns names from 'clinical.data' slot of MAF to be analysed for.
annotationDat	If MAF file was read without clinical data, provide a custom data.frame or a tsv file with a column containing Tumor_Sample_Barcodes along with clinical features. Default NULL.
minMut	Consider only genes with minimum this number of samples mutated. Default 5.
useCNV	whether to include copy number events. Only applicable when MAF is read along with copy number data. Default TRUE if available.

Value

result list containing p-values

See Also

[plotEnrichmentResults](#)

Examples

```
## Not run:
laml.maf = system.file('extdata', 'tcga_laml.maf.gz', package = 'maftools')
laml.clin = system.file('extdata', 'tcga_laml_annot.tsv', package = 'maftools')
laml = read.maf(maf = laml.maf, clinicalData = laml.clin)
clinicalEnrichment(laml, 'FAB_classification')

## End(Not run)
```

compareSignatures	<i>Compares identified denovo mutational signatures to known COSMIC signatures</i>
-------------------	--

Description

Takes results from [extractSignatures](#) and compares them known COSMIC signatures. Two COSMIC databases are used for comparisons - "legacy" which includes 30 signaures, and "SBS" - which includes updated/refined 65 signatures

Usage

```
compareSignatures(nmfRes, sig_db = "legacy", verbose = TRUE)
```

Arguments

nmfRes	results from extractSignatures
sig_db	can be legacy or SBS. Default legacy
verbose	Default TRUE

Details

SBS signature database was obtained from <https://www.synapse.org/#!/Synapse:syn11738319.7>

Value

list containing cosine similarities, aetiologies if available, and best match.

See Also

[trinucleotideMatrix](#) [extractSignatures](#) [plotSignatures](#)

coOncoplot

Draw two oncoplots side by side for cohort comparison.

Description

Draw two oncoplots side by side for cohort comparison.

Usage

```
coOncoplot(  
  m1,  
  m2,  
  genes = NULL,  
  m1Name = NULL,  
  m2Name = NULL,  
  clinicalFeatures1 = NULL,  
  clinicalFeatures2 = NULL,  
  annotationColor1 = NULL,  
  annotationColor2 = NULL,  
  annotationFontSize = 1.2,  
  sortByAnnotation1 = FALSE,  
  sortByAnnotation2 = FALSE,  
  additionalFeature1 = NULL,  
  additionalFeaturePch1 = 20,  
  additionalFeatureCol1 = "white",  
  additionalFeatureCex1 = 0.9,  
  additionalFeature2 = NULL,  
  additionalFeaturePch2 = 20,  
  additionalFeatureCol2 = "white",  
  additionalFeatureCex2 = 0.9,  
  sepwd_genes1 = 0.5,  
  sepwd_samples1 = 0.5,  
  sepwd_genes2 = 0.5,  
  sepwd_samples2 = 0.5,  
  colors = NULL,  
  removeNonMutated = TRUE,  
  geneNamefont = 0.8,  
  showSampleNames = FALSE,  
  SampleNamefont = 1,  
  legendFontSize = 1.2,  
)
```

```

    titleFontSize = 1.5,
    keepGeneOrder = FALSE,
    bgCol = "#CCCCCC",
    borderCol = "white"
)

```

Arguments

m1	first MAF object
m2	second MAF object
genes	draw these genes. Default plots top 5 mutated genes from two cohorts.
m1Name	optional name for first cohort
m2Name	optional name for second cohort
clinicalFeatures1	columns names from 'clinical.data' slot of m1 MAF to be drawn in the plot. Dafault NULL.
clinicalFeatures2	columns names from 'clinical.data' slot of m2 MAF to be drawn in the plot. Dafault NULL.
annotationColor1	list of colors to use for 'clinicalFeatures1' Default NULL.
annotationColor2	list of colors to use for 'clinicalFeatures2' Default NULL.
annotationFontSize	font size for annotations Default 1.2
sortByAnnotation1	logical sort oncomatrix (samples) by provided 'clinicalFeatures1'. Sorts based on first 'clinicalFeatures1'. Defaults to FALSE. column-sort
sortByAnnotation2	same as above but for m2
additionalFeature1	a vector of length two indicating column name in the MAF and the factor level to be highlighted.
additionalFeaturePch1	Default 20
additionalFeatureCol1	Default "white"
additionalFeatureCex1	Default 0.9
additionalFeature2	a vector of length two indicating column name in the MAF and the factor level to be highlighted.
additionalFeaturePch2	Default 20
additionalFeatureCol2	Default "white"
additionalFeatureCex2	Default 0.9
sepwd_genes1	Default 0.5
sepwd_samples1	Default 0.5

sepwd_genes2 Default 0.5
 sepwd_samples2 Default 0.5
 colors named vector of colors for each Variant_Classification.
 removeNonMutated
 Logical. If TRUE removes samples with no mutations in the oncoplot for better
 visualization. Default TRUE.
 geneNamefont font size for gene names. Default 1
 showSampleNames
 whether to show sample names. Default FALSE.
 SampleNamefont font size for sample names. Default 1
 legendFontSize font size for legend. Default 1.2
 titleFontSize font size for title. Default 1.5
 keepGeneOrder force the resulting plot to use the order of the genes as specified. Default FALSE
 bgCol Background grid color for wild-type (not-mutated) samples. Default gray -
 "#CCCCCC"
 borderCol border grid color for wild-type (not-mutated) samples. Default 'white'

Details

Draws two oncoplots side by side to display difference between two cohorts.

Value

Returns nothing. Just draws plot.

Examples

```

#' ##Primary and Relapse APL
primary.apl <- system.file("extdata", "APL_primary.maf.gz", package = "maftools")
relapse.apl <- system.file("extdata", "APL_relapse.maf.gz", package = "maftools")
##Read mafs
primary.apl <- read.maf(maf = primary.apl)
relapse.apl <- read.maf(maf = relapse.apl)
##Plot
coOncoplot(m1 = primary.apl, m2 = relapse.apl, m1Name = 'Primary APL', m2Name = 'Relapse APL')
dev.off()

```

drugInteractions

Drug-Gene Interactions

Description

Checks for drug-gene interactions and druggable categories

Usage

```
drugInteractions(  
  maf,  
  top = 20,  
  genes = NULL,  
  plotType = "bar",  
  drugs = FALSE,  
  fontSize = 0.8  
)
```

Arguments

maf	an MAF object generated by read.maf
top	Top number genes to check for. Default 20
genes	Manually specify gene list
plotType	Can be bar, pie, or wordCloud. Default bar plot.
drugs	Check for known/reported drugs. Default FALSE
fontSize	Default 0.8

Details

This function takes a list of genes and checks for known/reported drug-gene interactions or Drug-gable categories. All gene-drug interactions and drug claims are compiled from Drug Gene Interaction Database. See reference for details and cite it if you use this function.

References

Griffith, M., Griffith, O. L., Coffman, A. C., Weible, J. V., McMichael, J. F., Spies, N. C., et. al., 2013. DGIdb - Mining the druggable genome. Nature Methods.

Examples

```
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "maftools")  
lam1 <- read.maf(maf = lam1.maf)  
drugInteractions(maf = lam1)
```

estimateSignatures	<i>Estimate number of signatures based on cophenetic correlation metric</i>
--------------------	---

Description

Estimate number of signatures based on cophenetic correlation metric

Usage

```
estimateSignatures(
  mat,
  nMin = 2,
  nTry = 6,
  nrun = 10,
  parallel = 4,
  pConstant = NULL,
  verbose = TRUE,
  plotBestFitRes = FALSE
)
```

Arguments

mat	Input matrix of dimension nx96 generated by trinucleotideMatrix
nMin	Minimum number of signatures to try. Default 2.
nTry	Maximum number of signatures to try. Default 6.
nrun	numeric giving the number of run to perform for each value in range. Default 5
parallel	Default 4. Number of cores to use.
pConstant	A small positive value to add to the matrix. Use it ONLY if the functions throws a non-conformable arrays error
verbose	Default TRUE
plotBestFitRes	plots consensus heatmap for range of values tried. Default FALSE

Details

This function decomposes a non-negative matrix into n signatures. Extracted signatures are compared against 30 experimentally validated signatures by calculating cosine similarity. See <http://cancer.sanger.ac.uk/cosm> for details.

Value

a list with NMF.rank object and summary stats.

See Also

[plotCophenetic](#) [extractSignatures](#) [trinucleotideMatrix](#)

Examples

```
## Not run:
laml.tnm <- trinucleotideMatrix(maf = laml, ref_genome = 'hg19.fa', prefix = 'chr',
add = TRUE, useSyn = TRUE)
laml.sign <- estimateSignatures(mat = laml.tnm, plotBestFitRes = FALSE)

## End(Not run)
```

extractSignatures *Extract mutational signatures from trinucleotide context.*

Description

Decompose a matrix of 96 substitution classes into n signatures.

Usage

```
extractSignatures(  
  mat,  
  n = NULL,  
  plotBestFitRes = FALSE,  
  parallel = 4,  
  pConstant = NULL  
)
```

Arguments

mat	Input matrix of dimension nx96 generated by trinucleotideMatrix
n	decompose matrix into n signatures. Default NULL. Tries to predict best value for n by running NMF on a range of values and chooses based on cophenetic correlation coefficient.
plotBestFitRes	plots consensus heatmap for range of values tried. Default FALSE
parallel	Default 4. Number of cores to use.
pConstant	A small positive value to add to the matrix. Use it ONLY if the functions throws a non-conformable arrays error

Details

This function decomposes a non-negative matrix into n signatures.

Value

a list with decomposed scaled signatures, signature contributions in each sample and NMF object.

See Also

[trinucleotideMatrix](#) [plotSignatures](#) [compareSignatures](#)

Examples

```
## Not run:  
laml.tnm <- trinucleotideMatrix(maf = laml, ref_genome = 'hg19.fa', prefix = 'chr',  
  add = TRUE, useSyn = TRUE)  
laml.sign <- extractSignatures(mat = laml.tnm, plotBestFitRes = FALSE)  
  
## End(Not run)
```

`forestPlot`*Draw forest plot for differences between cohorts.*

Description

Draw forest plot for differences between cohorts.

Usage

```
forestPlot(  
  mafCompareRes,  
  pVal = 0.05,  
  fdr = NULL,  
  color = NULL,  
  geneFontSize = 1.2,  
  titleSize = 1.2,  
  lineWidth = 2.2  
)
```

Arguments

<code>mafCompareRes</code>	results from mafCompare
<code>pVal</code>	p-value threshold. Default 0.05.
<code>fdr</code>	fdr threshold. Default NULL. If provided uses adjusted pvalues (fdr).
<code>color</code>	vector of colors for cohorts. Default NULL.
<code>geneFontSize</code>	Font size for gene symbols. Default 1.2
<code>titleSize</code>	font size for titles. Default 1.2
<code>lineWidth</code>	line width for CI bars. Default 2.2

Details

Plots results from `link{mafCompare}` as a forest plot with x-axis as log10 converted odds ratio and differentially mutated genes on y-axis.

Value

Nothing

See Also

[mafCompare](#)

Examples

```
##Primary and Relapse APL  
primary.apl <- system.file("extdata", "APL_primary.maf.gz", package = "maftools")  
relapse.apl <- system.file("extdata", "APL_relapse.maf.gz", package = "maftools")  
##Read mafs  
primary.apl <- read.maf(maf = primary.apl)  
relapse.apl <- read.maf(maf = relapse.apl)
```

```
##Perform analysis and draw forest plot.
pt.vs.rt <- mafCompare(m1 = primary.ap1, m2 = relapse.ap1, m1Name = 'Primary',
m2Name = 'Relapse', minMut = 5)
forestPlot(mafCompareRes = pt.vs.rt)
```

geneCloud

Plots wordcloud.

Description

Plots word cloud of mutated genes or altered cytobands with size proportional to the event frequency.

Usage

```
geneCloud(input, minMut = 3, col = NULL, top = NULL, genesToIgnore = NULL, ...)
```

Arguments

input	an MAF or GISTIC object generated by read.maf or readGistic
minMut	Minimum number of samples in which a gene is required to be mutated.
col	vector of colors to choose from.
top	Just plot these top n number of mutated genes.
genesToIgnore	Ignore these genes.
...	Other options passed to wordcloud

Value

nothing.

Examples

```
lam1.input <- system.file("extdata", "tcga_lam1.maf.gz", package = "maftools")
lam1 <- read.maf(maf = lam1.input, useAll = FALSE)
geneCloud(input = lam1, minMut = 5)
```

genesToBarcodes

Extracts Tumor Sample Barcodes where the given genes are mutated.

Description

Extracts Tumor Sample Barcodes where the given genes are mutated.

Usage

```
genesToBarcodes(maf, genes = NULL, justNames = FALSE)
```

Arguments

maf	an MAF object generated by read.maf
genes	Hugo_Symbol for which sample names to be extracted.
justNames	if TRUE, just returns samples names instead of summarized tables.

Value

list of data . tables with samples in which given genes are mutated.

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml <- read.maf(maf = laml.maf)
genesToBarcodes(maf = laml, genes = 'DNMT3A')
```

genotypeMatrix	<i>Creates a Genotype Matrix for every variant</i>
----------------	--

Description

Creates a Genotype matrix using allele frequencies or by mutation status.

Usage

```
genotypeMatrix(
  maf,
  genes = NULL,
  tsb = NULL,
  includeSyn = FALSE,
  vafCol = NULL,
  vafCutoff = c(0.1, 0.75)
)
```

Arguments

maf	an MAF object generated by read.maf
genes	create matrix for only these genes. Define NULL
tsb	create matrix for only these tumor sample barcodes/samples. Define NULL
includeSyn	whether to include silent mutations. Default FALSE
vafCol	specify column name for vaf's. Default NULL. If not provided simply assumes all mutations are heterozygous.
vafCutoff	specify minimum and maximum vaf to define mutations as heterozygous. Default range 0.1 to 0.75. Mutations above maximum vafs are defined as homozygous.

Value

matrix

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml <- read.maf(maf = laml.maf)
genotypeMatrix(maf = laml, genes = "RUNX1")
```

getClinicalData *extract annotations from MAF object*

Description

extract annotations from MAF object

Usage

```
getClinicalData(x)

## S4 method for signature 'MAF'
getClinicalData(x)
```

Arguments

x An object of class MAF

Value

annotations associated with samples in MAF

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml <- read.maf(maf = laml.maf)
getClinicalData(x = laml)
```

getCytobandSummary *extract cytoband summary from GISTIC object*

Description

extract cytoband summary from GISTIC object

Usage

```
getCytobandSummary(x)

## S4 method for signature 'GISTIC'
getCytobandSummary(x)
```

Arguments

x An object of class GISTIC

Value

summarized gistic results by altered cytobands.

Examples

```
all.lesions <- system.file("extdata", "all_lesions.conf_99.txt", package = "maftools")
amp.genes <- system.file("extdata", "amp_genes.conf_99.txt", package = "maftools")
del.genes <- system.file("extdata", "del_genes.conf_99.txt", package = "maftools")
scores.gistic <- system.file("extdata", "scores.gistic", package = "maftools")
laml.gistic = readGistic(gisticAllLesionsFile = all.lesions, gisticAmpGenesFile = amp.genes, gisticDelGenesFi
getCytobandSummary(laml.gistic)
```

getFields

extract available fields from MAF object

Description

extract available fields from MAF object

Usage

```
getFields(x)

## S4 method for signature 'MAF'
getFields(x)
```

Arguments

x An object of class MAF

Value

Field names in MAF file

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml <- read.maf(maf = laml.maf)
getFields(x = laml)
```

getGeneSummary *extract gene summary from MAF or GISTIC object*

Description

extract gene summary from MAF or GISTIC object

Usage

```
getGeneSummary(x)

## S4 method for signature 'MAF'
getGeneSummary(x)

## S4 method for signature 'GISTIC'
getGeneSummary(x)
```

Arguments

x An object of class MAF or GISTIC

Value

gene summary table

Examples

```
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "maftools")
lam1 <- read.maf(maf = lam1.maf)
getGeneSummary(lam1)
```

getSampleSummary *extract sample summary from MAF or GISTIC object*

Description

extract sample summary from MAF or GISTIC object

Usage

```
getSampleSummary(x)

## S4 method for signature 'MAF'
getSampleSummary(x)

## S4 method for signature 'GISTIC'
getSampleSummary(x)
```

Arguments

x An object of class MAF or GISTIC

Value

sample summary table

Examples

```
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "maftools")
lam1 <- read.maf(maf = lam1.maf)
getSampleSummary(x = lam1)
```

GISTIC-class	<i>Class GISTIC</i>
--------------	---------------------

Description

S4 class for storing summarized MAF.

Slots

`data` data.table of summarized GISTIC file.
`cnv.summary` table containing alterations per sample
`cytoband.summary` table containing alterations per cytoband
`gene.summary` table containing alterations per gene
`cnMatrix` character matrix of dimension n*m where n is number of genes and m is number of samples
`numericMatrix` numeric matrix of dimension n*m where n is number of genes and m is number of samples
`gis.scores` gistic.scores
`summary` table with basic GISTIC summary stats
`classCode` mapping between numeric values in numericMatrix and copy number events.

See Also

[getGeneSummary](#) [getSampleSummary](#) [getCytobandSummary](#)

<code>gisticBubblePlot</code>	<i>Plot gistic results as a bubble plot</i>
-------------------------------	---

Description

Plots significantly altered cytobands as a function of number samples in which it is altered and number genes it contains. Size of each bubble is according to $-\log_{10}$ transformed q values.

Usage

```
gisticBubblePlot(
  gistic = NULL,
  color = NULL,
  markBands = NULL,
  fdrCutoff = 0.1,
  log_y = TRUE,
  txtSize = 3
)
```

Arguments

gistic	an object of class GISTIC generated by readGistic
color	colors for Amp and Del events.
markBands	any cytobands to label. Default top 5 lowest q values.
fdrCutoff	fdr cutoff to use. Default 0.1
log_y	log10 scale y-axis (# genes affected). Default TRUE
txtSize	label size for bubbles.

Value

Nothing

Examples

```
all.lesions <- system.file("extdata", "all_lesions.conf_99.txt", package = "maftools")
amp.genes <- system.file("extdata", "amp_genes.conf_99.txt", package = "maftools")
del.genes <- system.file("extdata", "del_genes.conf_99.txt", package = "maftools")
scores.gistic <- system.file("extdata", "scores.gistic", package = "maftools")
laml.gistic = readGistic(gisticAllLesionsFile = all.lesions, gisticAmpGenesFile = amp.genes, gisticDelGenesFile = del.genes, gisticScoresFile = scores.gistic)
gisticBubblePlot(gistic = laml.gistic, markBands = "")
```

gisticChromPlot

Plot gistic results along linearized chromosome

Description

A genomic plot with segments highlighting significant Amplifications and Deletion regions.

Usage

```
gisticChromPlot(
  gistic = NULL,
  fdrCutoff = 0.1,
  markBands = NULL,
  color = NULL,
  ref.build = "hg19",
  cytobandOffset = 0.01,
  txtSize = 0.8,
  cytobandTxtSize = 0.6
)
```

Arguments

<code>gistic</code>	an object of class GISTIC generated by <code>readGistic</code>
<code>fdrCutOff</code>	fdr cutoff to use. Default 0.1
<code>markBands</code>	any cytobands to label. Default top 5 lowest q values.
<code>color</code>	colors for Amp and Del events.
<code>ref.build</code>	reference build. Could be hg18, hg19 or hg38.
<code>cytobandOffset</code>	if <code>scores.gistic</code> file is given use this to adjust cytoband size.
<code>txtSize</code>	label size for labels
<code>cytobandTxtSize</code>	label size for cytoband

Value

nothing

Examples

```
all.lesions <- system.file("extdata", "all_lesions.conf_99.txt", package = "maftools")
amp.genes <- system.file("extdata", "amp_genes.conf_99.txt", package = "maftools")
del.genes <- system.file("extdata", "del_genes.conf_99.txt", package = "maftools")
scores.gistic <- system.file("extdata", "scores.gistic", package = "maftools")
laml.gistic = readGistic(gisticAllLesionsFile = all.lesions, gisticAmpGenesFile = amp.genes, gisticDelGenesFile = del.genes, gisticScoresFile = scores.gistic)
gisticChromPlot(laml.gistic)
```

<code>gisticOncoPlot</code>	<i>Plot gistic results.</i>
-----------------------------	-----------------------------

Description

takes output generated by `readGistic` and draws a plot similar to `oncoplot`.

Usage

```
gisticOncoPlot(
  gistic = NULL,
  top = NULL,
  showTumorSampleBarcodes = FALSE,
  gene_mar = 5,
  barcode_mar = 6,
  sepwd_genes = 0.5,
  sepwd_samples = 0.25,
  clinicalData = NULL,
  clinicalFeatures = NULL,
  sortByAnnotation = FALSE,
  sampleOrder = NULL,
  annotationColor = NULL,
  bandsToIgnore = NULL,
  removeNonAltered = TRUE,
  colors = NULL,
```

```

    SampleNamefontSize = 0.6,
    fontSize = 0.8,
    legendFontSize = 1.2,
    annotationFontSize = 1.2
)

```

Arguments

<code>gistic</code>	an GISTIC object generated by readGistic
<code>top</code>	how many top cytobands to be drawn. defaults to all.
<code>showTumorSampleBarcodes</code>	logical to include sample names.
<code>gene_mar</code>	Default 5
<code>barcode_mar</code>	Default 6
<code>sepwd_genes</code>	Default 0.5
<code>sepwd_samples</code>	Default 0.25
<code>clinicalData</code>	data.frame with columns containing Tumor_Sample_Barcodes and rest of columns with annotations.
<code>clinicalFeatures</code>	columns names from 'clinicalData' to be drawn in the plot. Default NULL.
<code>sortByAnnotation</code>	logical sort oncomatrix (samples) by provided 'clinicalFeatures'. Defaults to FALSE. column-sort
<code>sampleOrder</code>	Manually specify sample names for oncolplot ordering. Default NULL.
<code>annotationColor</code>	list of colors to use for clinicalFeatures. Default NULL.
<code>bandsToIgnore</code>	do not show these bands in the plot Default NULL.
<code>removeNonAltered</code>	Logical. If TRUE removes samples with no mutations in the oncoplot for better visualization. Default FALSE.
<code>colors</code>	named vector of colors Amp and Del events.
<code>SampleNamefontSize</code>	font size for sample names. Default 0.6
<code>fontSize</code>	font size for cytoband names. Default 0.8
<code>legendFontSize</code>	font size for legend. Default 1.2
<code>annotationFontSize</code>	font size for annotations. Default 1.2

Details

Takes gistic file as input and plots it as a matrix. Any desired annotations can be added at the bottom of the oncoplot by providing annotation

Value

None.

See Also

[oncostrip](#)

Examples

```

all.lesions <- system.file("extdata", "all_lesions.conf_99.txt", package = "maftools")
amp.genes <- system.file("extdata", "amp_genes.conf_99.txt", package = "maftools")
del.genes <- system.file("extdata", "del_genes.conf_99.txt", package = "maftools")
scores.gistic <- system.file("extdata", "scores.gistic", package = "maftools")
laml.gistic = readGistic(gisticAllLesionsFile = all.lesions, gisticAmpGenesFile = amp.genes, gisticDelGenesFi
gisticOncoPlot(laml.gistic)

```

```
icgcSimpleMutationToMAF
```

Converts ICGC Simple Somatic Mutation format file to MAF

Description

Converts ICGC Simple Somatic Mutation format file to Mutation Annotation Format. Basic fields are converted as per MAF specifications, rest of the fields are retained as in the input file. Ensemble gene IDs are converted to HGNC Symbols. Note that by default Simple Somatic Mutation format contains all affected transcripts of a variant resulting in multiple entries of the same variant in same sample. It is hard to choose a single affected transcript based on annotations alone and by default this program removes repeated variants as duplicated entries. If you wish to keep all of them, set `removeDuplicatedVariants` to `FALSE`.

Usage

```

icgcSimpleMutationToMAF(
  icgc,
  basename = NA,
  MAFobj = FALSE,
  clinicalData = NULL,
  removeDuplicatedVariants = TRUE,
  addHugoSymbol = FALSE
)

```

Arguments

<code>icgc</code>	Input data in ICGC Simple Somatic Mutation format. Can be gz compressed.
<code>basename</code>	If given writes to output file with <code>basename</code> .
<code>MAFobj</code>	If <code>TRUE</code> returns results as an <code>MAF</code> object.
<code>clinicalData</code>	Clinical data associated with each sample/ <code>Tumor_Sample_Barcode</code> in MAF. Could be a text file or a <code>data.frame</code> . Default <code>NULL</code> .
<code>removeDuplicatedVariants</code>	removes repeated variants in a particular sample, mapped to multiple transcripts of same Gene. See Description. Default <code>TRUE</code> .
<code>addHugoSymbol</code>	If <code>TRUE</code> replaces ensemble gene IDs with <code>Hugo_Symbols</code> . Default <code>FALSE</code> .

Details

ICGC Simple Somatic Mutation format specification can be found here: <http://docs.icgc.org/submission/guide/icgc-simple-somatic-mutation-format/>

Value

tab delimited MAF file.

Examples

```
esca.icgc <- system.file("extdata", "simple_somatic_mutation.open.ESCA-CN.sample.tsv.gz", package = "maftool")
esca.maf <- icgcSimpleMutationToMAF(icgc = esca.icgc)
```

inferHeterogeneity *Clusters variants based on Variant Allele Frequencies (VAF).*

Description

takes output generated by read.maf and clusters variants to infer tumor heterogeneity. This function requires VAF for clustering and density estimation. VAF can be on the scale 0-1 or 0-100. Optionally if copy number information is available, it can be provided as a segmented file (e.g, from Circular Binary Segmentation). Those variants in copy number altered regions will be ignored.

Usage

```
inferHeterogeneity(
  maf,
  tsb = NULL,
  top = 5,
  vafCol = NULL,
  segFile = NULL,
  ignChr = NULL,
  minVaf = 0,
  maxVaf = 1,
  useSyn = FALSE,
  dirichlet = FALSE
)
```

Arguments

maf	an MAF object generated by read.maf
tsb	specify sample names (Tumor_Sample_Barcodes) for which clustering has to be done.
top	if tsb is NULL, uses top n number of most mutated samples. Defaults to 5.
vafCol	manually specify column name for vafs. Default looks for column 't_vaf'
segFile	path to CBS segmented copy number file. Column names should be Sample, Chromosome, Start, End, Num_Probes and Segment_Mean (log2 scale).
ignChr	ignore these chromosomes from analysis. e.g, sex chromosomes chrX, chrY. Default NULL.
minVaf	filter low frequency variants. Low vaf variants maybe due to sequencing error. Default 0. (on the scale of 0 to 1)
maxVaf	filter high frequency variants. High vaf variants maybe due to copy number alterations or impure tumor. Default 1. (on the scale of 0 to 1)
useSyn	Use synonymous variants. Default FALSE.
dirichlet	Deprecated! No longer supported. uses nonparametric dirichlet process for clustering. Default FALSE - uses finite mixture models.

Details

This function clusters variants based on VAF to estimate univariate density and cluster classification. There are two methods available for clustering. Default using parametric finite mixture models and another method using nonparametric infinite mixture models (Dirichlet process).

Value

list of clustering tables.

References

Chris Fraley and Adrian E. Raftery (2002) Model-based Clustering, Discriminant Analysis and Density Estimation *Journal of the American Statistical Association* 97:611-631

Jara A, Hanson TE, Quintana FA, Muller P, Rosner GL. DPpackage: Bayesian Semi- and Nonparametric Modeling in R. *Journal of statistical software*. 2011;40(5):1-30.

Olshen AB, Venkatraman ES, Lucito R, Wigler M. Circular binary segmentation for the analysis of array-based DNA copy number data. *Biostatistics*. 2004;5(4):557-72.

See Also

[plotClusters](#)

Examples

```
## Not run:
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "maftools")
lam1 <- read.maf(maf = lam1.maf)
TCGA.AB.2972.clust <- inferHeterogeneity(maf = lam1, tsb = 'TCGA-AB-2972', vafCol = 'i_TumorVAF_WU')

## End(Not run)
```

lollipopPlot

Draws lollipop plot of amino acid changes on to Protein structure.

Description

Draws lollipop plot of amino acid changes. Protein domains are derived from PFAM database.

Usage

```
lollipopPlot(
  maf,
  gene = NULL,
  AACol = NULL,
  labelPos = NULL,
  labPosSize = 0.9,
  showMutationRate = TRUE,
  showDomainLabel = TRUE,
  cBioPortal = FALSE,
  refSeqID = NULL,
  proteinID = NULL,
```



```

    repel = FALSE,
    collapsePosLabel = TRUE,
    legendTxtSize = 0.8,
    labPosAngle = 0,
    domainLabelSize = 0.8,
    axisTextSize = c(1, 1),
    printCount = FALSE,
    colors = NULL,
    domainColors = NULL,
    labelOnlyUniqueDoamins = TRUE,
    defaultYaxis = FALSE,
    titleSize = c(1.2, 1),
    pointSize = 1.5
)

```

Arguments

maf	an MAF object generated by read.maf
gene	HGNC symbol for which protein structure to be drawn.
AACol	manually specify column name for amino acid changes. Default looks for fields 'HGVS _{Sp} _Short', 'AAC _{Change} ' or 'Protein_Change'. Changes can be of any format i.e. can be a numeric value or HGVS _{Sp} annotations (e.g; p.P459L, p.L2195Pfs*30 or p.Leu2195ProfsTer30)
labelPos	Amino acid positions to label. If 'all', labels all variants.
labPosSize	Text size for labels. Default 0.9
showMutationRate	Whether to show the somatic mutation rate on the title. Default TRUE
showDomainLabel	Label domains within the plot. Default TRUE. If FALSE they will be annotated in legend.
cBioPortal	Adds annotations similar to cBioPortals MutationMapper and collapse Variants into Truncating and rest.
refSeqID	RefSeq transcript identifier for gene if known.
proteinID	RefSeq protein identifier for gene if known.
repel	If points are too close to each other, use this option to repel them. Default FALSE. Warning: naive method, might make plot ugly in case of too many variants!
collapsePosLabel	Collapses overlapping labels at same position. Default TRUE
legendTxtSize	Text size for legend. Default 0.8
labPosAngle	angle for labels. Defaults to horizontal 0 degree labels. Set to 90 for vertical; 45 for diagonal labels.
domainLabelSize	text size for domain labels. Default 0.8
axisTextSize	text size x and y tick labels. Default c(1,1).
printCount	If TRUE, prints number of summarized variants for the given protein.
colors	named vector of colors for each Variant_Classification. Default NULL.
domainColors	Manual colors for protein domains

labelOnlyUniqueDoamins Default TRUE only labels unique doamins.
 defaultYaxis If FALSE, just labels min and maximum y values on y axis.
 titleSize font size for title and subtitle. Default c(1.2, 1)
 pointSize size of lollipop heads. Default 1.5

Details

This function by default looks for fields 'HGVS_Short', 'AACChange' or 'Protein_Change' in maf file. One can also manually specify field name containing amino acid changes.

Value

Nothing

Examples

```
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "maftools")
lam1 <- read.maf(maf = lam1.maf)
lollipopPlot(maf = lam1, gene = 'KIT', AACol = 'Protein_Change')
```

lollipopPlot2

Compare two lollipop plots

Description

Compare two lollipop plots

Usage

```
lollipopPlot2(
  m1,
  m2,
  gene = NULL,
  AACol1 = NULL,
  AACol2 = NULL,
  m1_name = NULL,
  m2_name = NULL,
  m1_label = NULL,
  m2_label = NULL,
  refSeqID = NULL,
  proteinID = NULL,
  labPosAngle = 0,
  labPosSize = 0.9,
  colors = NULL,
  axisTextSize = c(1, 1),
  pointSize = 1.2,
  domainLabelSize = 1,
  legendTxtSize = 1
)
```

Arguments

m1	first MAF object
m2	second MAF object
gene	HGNC symbol for which protein structure to be drawn.
AACol1	manually specify column name for amino acid changes in m1. Default looks for fields 'HGVS_Short', 'AAChange' or 'Protein_Change'.
AACol2	manually specify column name for amino acid changes in m2. Default looks for fields 'HGVS_Short', 'AAChange' or 'Protein_Change'.
m1_name	name for m1 cohort. optional.
m2_name	name for m2 cohort. optional.
m1_label	Amino acid positions to label for m1 cohort. If 'all', labels all variants.
m2_label	Amino acid positions to label for m2 cohort. If 'all', labels all variants.
refSeqID	RefSeq transcript identifier for gene if known.
proteinID	RefSeq protein identifier for gene if known.
labPosAngle	angle for labels. Defaults to horizontal 0 degree labels. Set to 90 for vertical; 45 for diagonal labels.
labPosSize	Text size for labels. Default 3
colors	named vector of colors for each Variant_Classification. Default NULL.
axisTextSize	text size for axis labels. Default 1.
pointSize	size of lollipop heads. Default 1.2
domainLabelSize	text size for domain labels. Default 1.
legendTxtSize	Default 1.

Details

Draws lollipop plot for a gene from two cohorts

See Also

[lollipopPlot](#)

[mafCompare](#)

Examples

```
primary.apl <- system.file("extdata", "APL_primary.maf.gz", package = "maftools")
relapse.apl <- system.file("extdata", "APL_relapse.maf.gz", package = "maftools")
primary.apl <- read.maf(maf = primary.apl)
relapse.apl <- read.maf(maf = relapse.apl)
lollipopPlot2(m1 = primary.apl, m2 = relapse.apl, gene = "FLT3", AACol1 = "amino_acid_change", AACol2 = "amino_a
```

MAF-class	<i>Class MAF</i>
-----------	------------------

Description

S4 class for storing summarized MAF.

Slots

`data` `data.table` of MAF file containing all non-synonymous variants.
`variants.per.sample` table containing variants per sample
`variant.type.summary` table containing variant types per sample
`variant.classification.summary` table containing variant classification per sample
`gene.summary` table containing variant classification per gene
`summary` table with basic MAF summary stats
`maf.silent` subset of main MAF containing only silent variants
`clinical.data` clinical data associated with each sample/`Tumor_Sample_Barcode` in MAF.

See Also

[getGeneSummary](#) [getSampleSummary](#) [getFields](#)

mafCompare	<i>compare two cohorts (MAF).</i>
------------	-----------------------------------

Description

compare two cohorts (MAF).

Usage

```
mafCompare(m1, m2, m1Name = NULL, m2Name = NULL, minMut = 5, useCNV = TRUE)
```

Arguments

<code>m1</code>	first MAF object
<code>m2</code>	second MAF object
<code>m1Name</code>	optional name for first cohort
<code>m2Name</code>	optional name for second cohort
<code>minMut</code>	Consider only genes with minimum this number of samples mutated in at least one of the cohort for analysis. Helpful to ignore single mutated genes. Default 5.
<code>useCNV</code>	whether to include copy number events to compare MAFs. Only applicable when MAF is read along with copy number data. Default TRUE if available.

Details

Performs fisher test on 2x2 contingency table generated from two cohorts to find differentially mutated genes.

Value

result list

See Also

[forestPlot](#)

[lollipopPlot2](#)

Examples

```
primary.apl <- system.file("extdata", "APL_primary.maf.gz", package = "maftools")
relapse.apl <- system.file("extdata", "APL_relapse.maf.gz", package = "maftools")
primary.apl <- read.maf(maf = primary.apl)
relapse.apl <- read.maf(maf = relapse.apl)
pt.vs.rt <- mafCompare(m1 = primary.apl, m2 = relapse.apl, m1Name = 'Primary',
m2Name = 'Relapse', minMut = 5)
```

mafSummary

Summary statistics of MAF

Description

Summarizes genes and samples irrespective of the type of alteration. This is different from [getSampleSummary](#) and [getGeneSummary](#) which returns summaries of only non-synonymous variants.

Usage

```
mafSummary(maf)
```

Arguments

maf an MAF object generated by [read.maf](#)

Details

This function takes MAF object as input and returns summary table.

Value

Returns a list of summarized tables

See Also

[getGeneSummary](#) [getSampleSummary](#)

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml <- read.maf(maf = laml.maf)
mafSummary(maf = laml)
```

mafSurvGroup

Performs survival analysis for a geneset

Description

Similar to [mafSurvival](#) but for a geneset

Usage

```
mafSurvGroup(
  maf,
  geneSet = NULL,
  clinicalData = NULL,
  time = "Time",
  Status = "Status"
)
```

Arguments

maf	an MAF object generated by read.maf
geneSet	gene names for which survival analysis needs to be performed. Samples with mutations in ALL of the genes provided are used as genes-set mutants.
clinicalData	dataframe containing events and time to events. Default looks for clinical data in annotation slot of MAF.
time	column name containing time in clinicalData
Status	column name containing status of patients in clinicalData. must be logical or numeric. e.g, TRUE or FALSE, 1 or 0.

Value

Survival plot

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml.clin <- system.file("extdata", "tcga_laml_annot.tsv", package = "maftools")
laml <- read.maf(maf = laml.maf, clinicalData = laml.clin)
mafSurvGroup(maf = laml, geneSet = c('DNMT3A', 'FLT3'), time = 'days_to_last_followup', Status = 'Overall_Survival')
```

mafSurvival	<i>Performs survival analysis</i>
-------------	-----------------------------------

Description

Performs survival analysis by grouping samples from maf based on mutation status of given gene(s) or manual grouping of samples.

Usage

```
mafSurvival(
  maf,
  genes = NULL,
  samples = NULL,
  clinicalData = NULL,
  time = "Time",
  Status = "Status",
  groupNames = c("Mutant", "WT"),
  showConfInt = TRUE,
  addInfo = TRUE,
  col = c("maroon", "royalblue"),
  isTCGA = FALSE,
  textSize = 12,
  fn = NULL,
  width = 6,
  height = 6
)
```

Arguments

maf	an MAF object generated by read.maf
genes	gene names for which survival analysis needs to be performed. Samples with mutations in any one of the genes provided are used as mutants.
samples	samples to group by. Genes and samples are mutually exclusive.
clinicalData	dataframe containing events and time to events. Default looks for clinical data in annotation slot of MAF .
time	column name containing time in clinicalData
Status	column name containing status of patients in clinicalData. must be logical or numeric. e.g, TRUE or FALSE, 1 or 0.
groupNames	names for groups. Should be of length two. Default c("Mutant", "WT")
showConfInt	TRUE. Whether to show confidence interval in KM plot.
addInfo	TRUE. Whether to show survival info in the plot.
col	colors for plotting.
isTCGA	FALSE. Is data is from TCGA.
textSize	Text size for surv table. Default 7.
fn	NULL. If provided saves pdf plot with basename fn.
width	width of plot to be saved. Default 6
height	height of plot to be saved. Default 6

Details

This function takes MAF file and groups them based on mutation status associated with given gene(s) and performs survival analysis. Requires dataframe containing survival status and time to event. Make sure sample names match to Tumor Sample Barcodes from MAF file.

Value

Survival plot

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "mafTools")
laml.clin <- system.file("extdata", "tcga_laml_annot.tsv", package = "mafTools")
laml <- read.maf(maf = laml.maf, clinicalData = laml.clin)
mafSurvival(maf = laml, genes = 'DNMT3A', time = 'days_to_last_followup', Status = 'Overall_Survival_Status',
```

math.score

calculates MATH (Mutant-Allele Tumor Heterogeneity) score.

Description

calculates MATH scores from variant allele frequencies. Mutant-Allele Tumor Heterogeneity (MATH) score is a measure of intra-tumor genetic heterogeneity. High MATH scores are related to lower survival rates. This function requires vafs.

Usage

```
math.score(maf, vafCol = NULL, sampleName = NULL, vafCutOff = 0.075)
```

Arguments

maf	an MAF object generated by read.maf
vafCol	manually specify column name for vafs. Default looks for column 't_vaf'
sampleName	sample name for which MATH score to be calculated. If NULL, calculates for all samples.
vafCutOff	minimum vaf for a variant to be considered for score calculation. Default 0.075

Value

data.table with MATH score for every Tumor_Sample_Barcode

References

Mroz, Edmund A. et al. Intra-Tumor Genetic Heterogeneity and Mortality in Head and Neck Cancer: Analysis of Data from The Cancer Genome Atlas. Ed. Andrew H. Beck. PLoS Medicine 12.2 (2015): e1001786.

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml <- read.maf(maf = laml.maf)
laml.math <- math.score(maf = laml, vafCol = 'i_TumorVAF_WU',
  sampleName = c('TCGA-AB-3009', 'TCGA-AB-2849', 'TCGA-AB-3002', 'TCGA-AB-2972'))
```

merge_mafs	<i>Merge multiple mafs into single MAF</i>
------------	--

Description

Merges multiple maf files/objects/data.frames into a single MAF.

Usage

```
merge_mafs(mafs, verbose = TRUE, ...)
```

Arguments

mafs	a list of MAF objects or data.frames or paths to MAF files.
verbose	Default TRUE
...	additional arguments passed read.maf

Value

[MAF](#) object

mutCountMatrix	<i>Generates count matrix of mutations.</i>
----------------	---

Description

Generates a count matrix of mutations. i.e, number of mutations per gene per sample.

Usage

```
mutCountMatrix(
  maf,
  includeSyn = FALSE,
  countOnly = NULL,
  removeNonMutated = TRUE
)
```

Arguments

maf	an MAF object generated by read.maf
includeSyn	whether to include synonymous variants in output matrix. Default FALSE
countOnly	Default NULL - counts all variants. You can specify type of 'Variant_Classification' to count. For e.g, countOnly = 'Splice_Site' will generate matrix for only Splice_Site variants.
removeNonMutated	Logical Default TRUE, removes samples with no mutations from the matrix.

Value

Integer Matrix

See Also[getFields](#) [getGeneSummary](#) [getSampleSummary](#)**Examples**

```

laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml <- read.maf(maf = laml.maf)
##Generate matrix
mutCountMatrix(maf = laml)
##Generate count matrix of Splice_Site mutations
mutCountMatrix(maf = laml, countOnly = 'Splice_Site')

```

oncodrive

*Detect cancer driver genes based on positional clustering of variants.***Description**

Clusters variants based on their position to detect disease causing genes.

Usage

```

oncodrive(
  maf,
  AACol = NULL,
  minMut = 5,
  pvalMethod = "zscore",
  nBgGenes = 100,
  bgEstimate = TRUE,
  ignoreGenes = NULL
)

```

Arguments

maf	an MAF object generated by read.maf
AACol	manually specify column name for amino acid changes. Default looks for field 'AChange'
minMut	minimum number of mutations required for a gene to be included in analysis. Default 5.
pvalMethod	either zscore (default method for oncodriveCLUST), poisson or combined (uses lowest of the two pvalues).
nBgGenes	minimum number of genes required to estimate background score. Default 100. Do not change this unless its necessary.
bgEstimate	If FALSE skips background estimation from synonymous variants and uses predefined values estimated from COSMIC synonymous variants.
ignoreGenes	Ignore these genes from analysis. Default NULL. Helpful in case data contains large number of variants belonging to polymorphic genes such as mucins and TTN.

Details

This is the re-implimentation of algorithm defined in OncodriveCLUST article. Concept is based on the fact that most of the variants in cancer causing genes are enriched at few specific loci (aka hotspots). This method takes advantage of such positions to identify cancer genes. Cluster score of 1 means, a single hotspot hosts all observed variants. If you use this function, please cite OncodriveCLUST article.

Value

data table of genes ordered according to p-values.

References

Tamborero D, Gonzalez-Perez A and Lopez-Bigas N. OncodriveCLUST: exploiting the positional clustering of somatic mutations to identify cancer genes. *Bioinformatics*. 2013; doi: 10.1093/bioinformatics/btt395s

See Also

[plotOncodrive](#)

Examples

```
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "mafTools")
lam1 <- read.maf(maf = lam1.maf)
lam1.sig <- oncodrive(maf = lam1, AACol = 'Protein_Change', minMut = 5)
```

OncogenicPathways

Enrichment of known oncogenic pathways

Description

Checks for enrichment of known oncogenic pathways

Usage

```
OncogenicPathways(maf)
```

Arguments

maf an [MAF](#) object generated by [read.maf](#)

Details

Oncogenic signalling pathways are derived from TCGA cohorts. See reference for details.

Value

Prints fraction of altered pathway

References

Sanchez-Vega F, Mina M, Armenia J, Chatila WK, Luna A, La KC, Dimitriadoy S, Liu DL, Kantheti HS, Saghaflinia S et al. 2018. Oncogenic Signaling Pathways in The Cancer Genome Atlas. Cell 173: 321-337 e310

See Also

[PlotOncogenicPathways](#)

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml <- read.maf(maf = laml.maf)
OncogenicPathways(maf = laml)
```

oncoplot

draw an oncoplot

Description

takes output generated by read.maf and draws an oncoplot

Usage

```
oncoplot(
  maf,
  top = 20,
  genes = NULL,
  altered = FALSE,
  mutsig = NULL,
  mutsigQval = 0.1,
  drawRowBar = TRUE,
  drawColBar = TRUE,
  includeColBarCN = TRUE,
  draw_titv = FALSE,
  logColBar = FALSE,
  clinicalFeatures = NULL,
  exprsTbl = NULL,
  additionalFeature = NULL,
  additionalFeaturePch = 20,
  additionalFeatureCol = "white",
  additionalFeatureCex = 0.9,
  annotationDat = NULL,
  annotationColor = NULL,
  genesToIgnore = NULL,
  showTumorSampleBarcodes = FALSE,
  barcode_mar = 4,
  gene_mar = 5,
  removeNonMutated = TRUE,
  fill = TRUE,
  cohortSize = NULL,
```

```

colors = NULL,
sortByMutation = FALSE,
sortByAnnotation = FALSE,
numericAnnoCol = NULL,
groupAnnotationBySize = TRUE,
annotationOrder = NULL,
keepGeneOrder = FALSE,
GeneOrderSort = TRUE,
sampleOrder = NULL,
writeMatrix = FALSE,
sepwd_genes = 0.5,
sepwd_samples = 0.25,
fontSize = 0.8,
SampleNamefontSize = 1,
showTitle = TRUE,
titleFontSize = 1.5,
legendFontSize = 1.2,
annotationFontSize = 1.2,
bgCol = "#CCCCCC",
borderCol = "white",
colbar_pathway = FALSE
)

```

Arguments

maf	an MAF object generated by read.maf
top	how many top genes to be drawn. defaults to 20.
genes	Just draw oncoplot for these genes. Default NULL.
altered	Default FALSE. Chooses top genes based on mutation status. If TRUE chooses top genes based alterations (CNV or mutation).
mutsig	Mutsig results if available. Usually file named sig_genes.txt If provided plots significant genes and corresponding Q-values as side row-bar. Default NULL.
mutsigQval	Q-value to choose significant genes from mutsig results. Default 0.1
drawRowBar	logical plots barplot for each gene. Default TRUE.
drawColBar	logical plots barplot for each sample. Default TRUE.
includeColBarCN	Whether to include CN in column bar plot. Default TRUE
draw_titv	logical Includes TiTv plot. FALSE
logColBar	Plot top bar plot on log10 scale. Default FALSE.
clinicalFeatures	columns names from 'clinical.data' slot of MAF to be drawn in the plot. Default NULL.
exprsTbl	Expression values if available. Must be a data.frame with two columns containing to gene names and expression values.
additionalFeature	a vector of length two indicating column name in the MAF and the factor level to be highlighted.
additionalFeaturePch	Default 20

additionalFeatureCol	Default "white"
additionalFeatureCex	Default 0.9
annotationDat	If MAF file was read without clinical data, provide a custom data.frame with a column Tumor_Sample_Barcode containing sample names along with rest of columns with annotations. You can specify which columns to be drawn using 'clinicalFeatures' argument.
annotationColor	Custom colors to use for 'clinicalFeatures'. Must be a named list containing a named vector of colors. Default NULL. See example for more info.
genesToIgnore	do not show these genes in Oncoplot. Default NULL.
showTumorSampleBarcodes	logical to include sample names.
barcode_mar	Default 4
gene_mar	Default 5
removeNonMutated	Logical. If TRUE removes samples with no mutations in the oncoplot for better visualization. Default TRUE.
fill	Logical. If TRUE draws genes and samples as blank grids even when they are not altered.
cohortSize	Number of sequenced samples in the cohort. Default all samples from Cohort. You can manually specify the cohort size. Default NULL
colors	named vector of colors for each Variant_Classification.
sortByMutation	Force sort matrix according mutations. Helpful in case of MAF was read along with copy number data. Default FALSE.
sortByAnnotation	logical sort oncomatrix (samples) by provided 'clinicalFeatures'. Sorts based on first 'clinicalFeatures'. Defaults to FALSE. column-sort
numericAnnoCol	color palette used for numeric annotations. Default 'YlOrBr' from RColorBrewer
groupAnnotationBySize	Further group 'sortByAnnotation' orders by their size. Defaults to TRUE. Largest groups comes first.
annotationOrder	Manually specify order for annotations. Works only for first 'clinicalFeatures'. Default NULL.
keepGeneOrder	logical whether to keep order of given genes. Default FALSE, order according to mutation frequency
GeneOrderSort	logical this is applicable when 'keepGeneOrder' is TRUE. Default TRUE
sampleOrder	Manually specify sample names for oncolplot ordering. Default NULL.
writeMatrix	writes character coded matrix used to generate the plot to an output file.
sepwd_genes	Default 0.5
sepwd_samples	Default 0.25
fontSize	font size for gene names. Default 0.8.
SampleNamefontSize	font size for sample names. Default 1

showTitle	Default TRUE
titleFontSize	font size for title. Default 1.5
legendFontSize	font size for legend. Default 1.2
annotationFontSize	font size for annotations. Default 1.2
bgCol	Background grid color for wild-type (not-mutated) samples. Default gray - "#CCCCCC"
borderCol	border grid color (not-mutated) samples. Default 'white'.
colbar_pathway	Draw top column bar with respect to displayed pathway. Default FALSE.

Details

Takes maf file as input and plots it as a matrix. Any desired clinical features can be added at the bottom of the oncoplot by providing `clinicalFeatures`. Oncoplot can be sorted either by mutations or by `clinicalFeatures` using arguments `sortByMutation` and `sortByAnnotation` respectively.

Value

None.

See Also

[oncostrip](#)

Examples

```

laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml.clin = system.file('extdata', 'tcga_laml_annot.tsv', package = 'maftools')
laml <- read.maf(maf = laml.maf, clinicalData = laml.clin)
#Basic oncoplot
oncoplot(maf = laml, top = 3)
#Changing colors for variant classifications (You can use any colors, here in this example we will use a color palette)
col = RColorBrewer::brewer.pal(n = 8, name = 'Paired')
names(col) = c('Frame_Shift_Del', 'Missense_Mutation', 'Nonsense_Mutation', 'Multi_Hit', 'Frame_Shift_Ins',
              'In_Frame_Ins', 'Splice_Site', 'In_Frame_Del')
#Color coding for FAB classification; try getAnnotations(x = laml) to see available annotations.
fabcolors = RColorBrewer::brewer.pal(n = 8, name = 'Spectral')
names(fabcolors) = c("M0", "M1", "M2", "M3", "M4", "M5", "M6", "M7")
fabcolors = list(FAB_classification = fabcolors)
oncoplot(maf = laml, colors = col, clinicalFeatures = 'FAB_classification', sortByAnnotation = TRUE, annotation

```

oncostrip

draw an oncostrip similar to cBioportal oncoprinter output.

Description

draw an oncostrip similar to cBioportal oncoprinter output.

Usage

```
oncostrip(maf = NULL, ...)
```

Arguments

maf an [MAF](#) object generated by `read.maf`
... arguments passed [oncoplot](#)

Details

This is just a wrapper around [oncoplot](#) with `drawRowBar` and `drawColBar` set to `FALSE`

Value

None.

See Also

[oncoplot](#)

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml <- read.maf(maf = laml.maf)
dev.new()
oncostrip(maf = laml, genes = c('NPM1', 'RUNX1'))
```

pancanComparison

Perform PacCancer analysis

Description

Takes MutSig results and compares them against PanCancer results.

Usage

```
pancanComparison(  
  mutsigResults,  
  qval = 0.1,  
  cohortName = "input",  
  inputSampleSize = NULL,  
  label = 1,  
  genesToLabel = NULL,  
  pointSize = 0.1,  
  labelSize = 0.8  
)
```

Arguments

`mutsigResults` MutSig results (usually `sig_genes.txt`). Can be gz compressed.
`qval` qvalue threshold to define SMG. Default 0.1
`cohortName` Input cohort name.
`inputSampleSize` Sample size from MAF file used to generate mutSig results. Optional.

label	Default 1. Can be 1, 2 or 3.
genesToLabel	Default NULL. Exclusive with label argument.
pointSize	size for scatter plot. Default 1.
labelSize	label text size. Default 1

Details

This function takes MutSig results and compares them against panCancer cohort (~5000 tumor samples from 21 cancer types). This analysis can reveal novel genes exclusively mutated in input cohort.

Value

result table

References

Lawrence MS, Stojanov P, Mermel CH, et al. Discovery and saturation analysis of cancer genes across 21 tumor types. *Nature*. 2014;505(7484):495-501. doi:10.1038/nature12912.

Examples

```
lam1.mutsig <- system.file("extdata", "LAML_sig_genes.txt.gz", package = "maftools")
pancanComparison(mutsigResults = lam1.mutsig, qval = 0.1, cohortName = 'LAML', inputSampleSize = 200, label = 1)
```

pfamDomains	<i>pfam domain annotation and summarization.</i>
-------------	--

Description

Summarizes amino acid positions and annotates them with pfam domain information.

Usage

```
pfamDomains(  
  maf = NULL,  
  AACol = NULL,  
  summarizeBy = "AAPos",  
  top = 5,  
  domainsToLabel = NULL,  
  baseName = NULL,  
  varClass = "nonSyn",  
  width = 5,  
  height = 5,  
  labelSize = 1  
)
```

Arguments

maf	an MAF object generated by <code>read.maf</code>
AACol	manually specify column name for amino acid changes. Default looks for field 'AChange'
summarizeBy	Summarize domains by amino acid position or conversions. Can be "AAPos" or "AChange"
top	How many top mutated domains to label in the scatter plot. Defaults to 5.
domainsToLabel	Default NULL. Exclusive with top argument.
baseName	If given writes the results to output file. Default NULL.
varClass	which variants to consider for summarization. Can be nonSyn, Syn or all. Default nonSyn.
width	width of the file to be saved.
height	height of the file to be saved.
labelSize	font size for labels. Default 1.

Value

returns a list two tables summarized by amino acid positions and domains respectively. Also plots top 5 most mutated domains as scatter plot.

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml <- read.maf(maf = laml.maf)
pfamDomains(maf = laml, AACol = 'Protein_Change')
```

plotApobecDiff	<i>Plot differences between APOBEC enriched and non-APOBEC enriched samples.</i>
----------------	--

Description

Plots differences between APOBEC enriched and non-APOBEC enriched samples

Usage

```
plotApobecDiff(
  tnm,
  maf,
  pVal = 0.05,
  title_size = 1,
  axis_lwd = 1,
  font_size = 1.2
)
```

Arguments

tnm	output generated by trinucleotideMatrix
maf	an MAF object used to generate the matrix
pVal	p-value threshold for fisher's test. Default 0.05.
title_size	size of title. Default 1.3
axis_lwd	axis width. Default 1
font_size	font size. Default 1.2

Details

Plots differences between APOBEC enriched and non-APOBEC enriched samples (TCW). Plot includes differences in mutations load, tCw motif distribution and top genes altered.

Value

list of table containing differentially altered genes. This can be passed to [forestPlot](#) to plot results.

See Also

[trinucleotideMatrix](#) [plotSignatures](#)

Examples

```
## Not run:
lam1.tnm <- trinucleotideMatrix(maf = lam1, ref_genome = 'hg19.fa', prefix = 'chr',
add = TRUE, useSyn = TRUE)
plotApobecDiff(lam1.tnm)

## End(Not run)
```

plotCBSsegments	<i>Plots segmented copy number data.</i>
-----------------	--

Description

Plots segmented copy number data.

Usage

```
plotCBSsegments(
  cbsFile = NULL,
  maf = NULL,
  tsb = NULL,
  savePlot = FALSE,
  ylims = NULL,
  seg_size = 0.1,
  width = 6,
  height = 3,
  genes = NULL,
```

```

    ref.build = "hg19",
    writeTable = FALSE,
    removeXY = FALSE,
    color = NULL
  )

```

Arguments

cbsFile	CBS segmented copy number file. Column names should be Sample, Chromosome, Start, End, Num_Probes and Segment_Mean (log2 scale).
maf	optional MAF
tsb	If segmentation file contains many samples (as in gistic input), specify sample name here. Default plots all samples. If you are mapping maf, make sure sample names in Sample column of segmentation file matches to those Tumor_Sample_Barcodes in MAF.
savePlot	If true plot is saved as pdf.
ylims	Default NULL
seg_size	Default 0.1
width	width of plot
height	height of plot
genes	If given and maf object is specified, maps all mutations from maf onto segments. Default NULL
ref.build	Reference build for chromosome sizes. Can be hg18, hg19 or hg38. Default hg19.
writeTable	If true and if maf object is specified, writes plot data with each variant and its corresponding copynumber to an output file.
removeXY	don not plot sex chromosomes.
color	Manually specify color scheme for chromosomes. Default NULL. i.e, alternating Gray70 and midnightblue

Details

this function takes segmented copy number data and plots it. If MAF object is specified, all mutations are highlighted on the plot.

Value

Draws plot

Examples

```

tcga.ab.009.seg <- system.file("extdata", "TCGA.AB.3009.hg19.seg.txt", package = "maftools")
plotCBSsegments(cbsFile = tcga.ab.009.seg)

```

plotClusters	<i>Plot density plots from clustering results.</i>
--------------	--

Description

Plots results from `inferHeterogeneity`.

Usage

```
plotClusters(  
  clusters,  
  tsb = NULL,  
  genes = NULL,  
  showCNvars = FALSE,  
  colors = NULL  
)
```

Arguments

<code>clusters</code>	clustering results from inferHeterogeneity
<code>tsb</code>	sample to plot from clustering results. Default plots all samples from results.
<code>genes</code>	genes to highlight on the plot. Can be a vector of gene names, <code>CN_altered</code> to label copy number altered variants. or <code>all</code> to label all genes. Default <code>NULL</code> .
<code>showCNvars</code>	show copy numbered altered variants on the plot. Default <code>FALSE</code> .
<code>colors</code>	manual colors for clusters. Default <code>NULL</code> .

Value

returns nothing.

See Also

[inferHeterogeneity](#)

Examples

```
## Not run:  
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "maftools")  
lam1 <- read.maf(maf = lam1.maf)  
seg = system.file('extdata', 'TCGA.AB.3009.hg19.seg.txt', package = 'maftools')  
TCGA.AB.3009.clust <- inferHeterogeneity(maf = lam1, tsb = 'TCGA-AB-3009',  
  segFile = seg, vafCol = 'i_TumorVAF_WU')  
plotClusters(TCGA.AB.3009.clust, genes = c('NF1', 'SUZ12'), showCNvars = TRUE)  
  
## End(Not run)
```

plotCophenetic	<i>Draw an elbow plot of cophenetic correlation metric.</i>
----------------	---

Description

Draw an elbow plot of cophenetic correlation metric.

Usage

```
plotCophenetic(res = NULL, bestFit = NULL)
```

Arguments

res	output from estimateSignatures
bestFit	rank to highlight. Default NULL

Details

This function draws an elbow plot of cophenetic correlation metric.

See Also

[estimateSignatures](#) [plotCophenetic](#)

plotEnrichmentResults	<i>Plots results from clinicalEnrichment analysis</i>
-----------------------	---

Description

Plots results from clinicalEnrichment analysis

Usage

```
plotEnrichmentResults(  
  enrich_res,  
  pVal = 0.05,  
  cols = NULL,  
  annoFontSize = 0.8,  
  geneFontSize = 0.8,  
  legendFontSize = 0.8,  
  showTitle = TRUE  
)
```

Arguments

enrich_res	results from clinicalEnrichment or signatureEnrichment
pVal	Default 0.05
cols	named vector of colors for factor in a clinical feature. Default NULL
annoFontSize	cex for annotation font size. Default 0.8
geneFontSize	cex for gene font size. Default 0.8
legendFontSize	cex for legend font size. Default 0.8
showTitle	Default TRUE

Value

returns nothing.

See Also

[clinicalEnrichment](#) [signatureEnrichment](#)

plotmafSummary	<i>Plots maf summary.</i>
----------------	---------------------------

Description

Plots maf summary.

Usage

```
plotmafSummary(
  maf,
  rmOutlier = TRUE,
  dashboard = TRUE,
  titvRaw = TRUE,
  log_scale = FALSE,
  addStat = NULL,
  showBarcodes = FALSE,
  fs = 1,
  textSize = 0.8,
  color = NULL,
  titleSize = c(1, 0.8),
  titvColor = NULL,
  top = 10
)
```

Arguments

maf	an MAF object generated by read.maf
rmOutlier	If TRUE removes outlier from boxplot.
dashboard	If FALSE plots simple summary instead of dashboard style.
titvRaw	TRUE. If false instead of raw counts, plots fraction.

log_scale	FALSE. If TRUE log10 transforms Variant Classification, Variant Type and Variants per sample sub-plots.
addStat	Can be either mean or median. Default NULL.
showBarcodes	include sample names in the top bar plot.
fs	base size for text. Default 1
textSize	font size if showBarcodes is TRUE. Default 0.8
color	named vector of colors for each Variant_Classification.
titleSize	font size for title and subtitle. Default c(10, 8)
titvColor	colors for SNV classifications.
top	include top n genes dashboard plot. Default 10.

Value

Prints plot.

See Also

[read.maf](#) [MAF](#)

Examples

```
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "maftools")
lam1 <- read.maf(maf = lam1.maf, useAll = FALSE)
plotmafSummary(maf = lam1, addStat = 'median')
```

plotOncodrive

Plots results from oncodrive

Description

Takes results from oncodrive and plots them as a scatter plot. Size of the gene shows number of clusters (hotspots), x-axis can either be an absolute number of variants accumulated in these clusters or a fraction of total variants found in these clusters. y-axis is fdr values transformed into $-\log_{10}$ for better representation. Labels indicate Gene name with number clusters observed.

Usage

```
plotOncodrive(
  res = NULL,
  fdrCutOff = 0.05,
  useFraction = FALSE,
  colCode = NULL,
  bubbleSize = 1,
  labelSize = 1
)
```


Arguments

res	results from oncodrive
fdrCutOff	fdr cutoff to call a gene as a driver.
useFraction	if TRUE uses a fraction of total variants as X-axis scale instead of absolute counts.
colCode	Colors to use for indicating significant and non-significant genes. Default NULL
bubbleSize	Size for bubbles. Default 2.
labelSize	font size for labelling genes. Default 1.

Value

Nothing

See Also

[oncodrive](#)

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "mafTools")
laml <- read.maf(maf = laml.maf)
laml.sig <- oncodrive(maf = laml, AACol = 'Protein_Change', minMut = 5)
plotOncodrive(res = laml.sig, fdrCutOff = 0.1)
```

PlotOncogenicPathways *Plot oncogenic pathways*

Description

Plot oncogenic pathways

Usage

```
PlotOncogenicPathways(  
  maf,  
  pathways = NULL,  
  fullPathway = FALSE,  
  removeNonMutated = TRUE,  
  tsgCol = "red",  
  ogCol = "royalblue",  
  fontSize = 0.6,  
  showTumorSampleBarcodes = FALSE,  
  sampleOrder = NULL,  
  SampleNameFontSize = 0.6  
)
```

Arguments

maf	an MAF object generated by read.maf
pathways	Name of pathways to be drawn
fullPathway	Include all genes from the pathway. Default FALSE only plots mutated genes
removeNonMutated	Default TRUE
tsgCol	Color for tumor suppressor genes. Default red
ogCol	Color for onco genes. Default royalblue
fontSize	Default 0.6
showTumorSampleBarcodes	logical to include sample names.
sampleOrder	Manually specify sample names for oncolplot ordering. Default NULL.
SampleNamefontSize	font size for sample names. Default 10

Details

Draws oncolplot of oncogenic pathway.

References

Sanchez-Vega F, Mina M, Armenia J, Chatila WK, Luna A, La KC, Dimitriadoy S, Liu DL, Kantheti HS, Saghafeina S et al. 2018. Oncogenic Signaling Pathways in The Cancer Genome Atlas. Cell 173: 321-337 e310

See Also

[OncogenicPathways](#)

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml <- read.maf(maf = laml.maf)
PlotOncogenicPathways(maf = laml, pathways = "RTK-RAS")
```

plotSignatures

Plots decomposed mutational signatures

Description

Takes results from [extractSignatures](#) and plots decomposed mutational signatures as a barplot.

Usage

```
plotSignatures(
  nmfRes = NULL,
  contributions = FALSE,
  color = NULL,
  patient_order = NULL,
  font_size = 1.2,
  show_title = TRUE,
  sig_db = "legacy",
  axis_lwd = 2,
  title_size = 0.9,
  show_barcodes = FALSE,
  yaxisLim = 0.3,
  ...
)
```

Arguments

nmfRes	results from extractSignatures
contributions	If TRUE plots contribution of signatures in each sample.
color	colors for each Ti/Tv conversion class. Default NULL
patient_order	User defined ordering of samples. Default NULL.
font_size	font size. Default 1.2
show_title	If TRUE compares signatures to COSMIC signatures and prints them as title
sig_db	Only applicable if show_title is TRUE. Can be legacy or SBS. Default legacy
axis_lwd	axis width. Default 2.
title_size	size of title. Default 1.3
show_barcodes	Default FALSE
yaxisLim	Default 0.3. If NA autoscales.
...	further plot options passed to barplot

Value

Nothing

See Also

[trinucleotideMatrix](#) [plotSignatures](#)

plotTiTv

Plot Transition and Trasnversion ratios.

Description

Takes results generated from `titv` and plots the Ti/Tv ratios and contributions of 6 mutational conversion classes in each sample.

Usage

```
plotTiTv(
  res = NULL,
  plotType = "both",
  sampleOrder = NULL,
  color = NULL,
  showBarcodes = FALSE,
  textSize = 0.8,
  baseFontSize = 1,
  axisTextSize = c(1, 1),
  plotNotch = FALSE
)
```

Arguments

res	results generated by titv
plotType	Can be 'bar', 'box' or 'both'. Defaults to 'both'
sampleOrder	Sample names in which the barplot should be ordered. Default NULL
color	named vector of colors for each coversion class.
showBarcodes	Whether to include sample names for barplot
textSize	fontsize if showBarcodes is TRUE. Deafult 2.
baseFontSize	font size. Deafult 1.
axisTextSize	text size x and y tick labels. Default c(1,1).
plotNotch	logical. Include notch in boxplot.

Value

None.

See Also

[titv](#)

Examples

```
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "maftools")
lam1 <- read.maf(maf = lam1.maf)
lam1.titv = titv(maf = lam1, useSyn = TRUE)
plotTiTv(lam1.titv)
```

plotVaf

Plots vaf distribution of genes

Description

Plots vaf distribution of genes as a boxplot. Each dot in the jitter is a variant.

Usage

```
plotVaf(  
  maf,  
  vafCol = NULL,  
  genes = NULL,  
  top = 10,  
  orderByMedian = TRUE,  
  keepGeneOrder = FALSE,  
  flip = FALSE,  
  fn = NULL,  
  gene_fs = 0.8,  
  axis_fs = 0.8,  
  height = 5,  
  width = 5,  
  showN = TRUE  
)
```

Arguments

maf	an MAF object generated by read.maf
vafCol	manually specify column name for vafs. Default looks for column 't_vaf'
genes	specify genes for which plots has to be generated
top	if genes is NULL plots top n number of genes. Defaults to 5.
orderByMedian	Orders genes by decreasing median VAF. Default TRUE
keepGeneOrder	keep gene order. Default FALSE
flip	if TRUE, flips axes. Default FALSE
fn	Filename. If given saves plot as a output pdf. Default NULL.
gene_fs	font size for gene names. Default 0.8
axis_fs	font size for axis. Default 0.8
height	Height of plot to be saved. Default 5
width	Width of plot to be saved. Default 4
showN	if TRUE, includes number of observations

Value

Nothing.

Examples

```
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "maftools")  
lam1 <- read.maf(maf = lam1.maf)  
plotVaf(maf = lam1, vafCol = 'i_TumorVAF_WU')
```

prepareMutSig	<i>Prepares MAF file for MutSig analysis.</i>
---------------	---

Description

Corrects gene names for MutSig compatibility.

Usage

```
prepareMutSig(maf, fn = NULL)
```

Arguments

maf	an MAF object generated by read.maf
fn	basename for output file. If provided writes MAF to an output file with the given basename.

Details

MutSig/MutSigCV is most widely used program for detecting driver genes. However, we have observed that covariates files (gene.covariates.txt and exome_full192.coverage.txt) which are bundled with MutSig have non-standard gene names (non Hugo_Symbols). This discrepancy between Hugo_Symbols in MAF and non-Hugo_symbols in covariates file causes MutSig program to ignore such genes. For example, KMT2D - a well known driver gene in Esophageal Carcinoma is represented as MLL2 in MutSig covariates. This causes KMT2D to be ignored from analysis and is represented as an insignificant gene in MutSig results. This function attempts to correct such gene symbols with a manually curated list of gene names compatible with MutSig covariates list.

Value

returns a MAF with gene symbols corrected.

Examples

```
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "mafTools")
lam1 <- read.maf(maf = lam1.maf)
prepareMutSig(maf = lam1)
```

rainfallPlot	<i>Rainfall plot to display hyper mutated genomic regions.</i>
--------------	--

Description

Plots inter variant distance as a function of genomic locus.

Usage

```
rainfallPlot(  
  maf,  
  tsb = NULL,  
  detectChangePoints = FALSE,  
  ref.build = "hg19",  
  color = NULL,  
  savePlot = FALSE,  
  width = 6,  
  height = 3,  
  fontSize = 1.2,  
  pointSize = 0.4  
)
```

Arguments

maf	an MAF object generated by <code>read.maf</code> . Required.
tsb	specify sample names (Tumor_Sample_Barcodes) for which plotting has to be done. If NULL, draws plot for most mutated sample.
detectChangePoints	If TRUE, detects genomic change points where potential kataegis are formed. Results are written to an output tab delimited file.
ref.build	Reference build for chromosome sizes. Can be hg18, hg19 or hg38. Default hg19.
color	named vector of colors for each conversion class.
savePlot	If TRUE plot is saved to output pdf. Default FALSE.
width	width of plot to be saved.
height	height of plot to be saved.
fontSize	Default 12.
pointSize	Default 0.8.

Details

If ‘detectChangePoints’ is set to TRUE, this function will identify Kataegis loci. Kataegis detection algorithm by Moritz Goretzky at WWU Munster, which exploits the definition of Kataegis (six consecutive mutations with an avg. distance of 1000bp) to identify hyper mutated genomic loci. Algorithm starts with a double-ended queue to which six consecutive mutations are added and their average intermutation distance is calculated. If the average intermutation distance is larger than 1000, one element is added at the back of the queue and one is removed from the front. If the average intermutation distance is less or equal to 1000, further mutations are added until the average intermutation distance is larger than 1000. After that all mutations in the double-ended queue are written into output as one kataegis and the double-ended queue is reinitialized with six mutations.

Value

Results are written to an output file with suffix changePoints.tsv

read.maf	<i>Read MAF files.</i>
----------	------------------------

Description

Takes tab delimited MAF (can be plain text or gz compressed) file as an input and summarizes it in various ways. Also creates oncomatrix - helpful for visualization.

Usage

```
read.maf(
  maf,
  clinicalData = NULL,
  removeDuplicatedVariants = TRUE,
  useAll = TRUE,
  gisticAllLesionsFile = NULL,
  gisticAmpGenesFile = NULL,
  gisticDelGenesFile = NULL,
  gisticScoresFile = NULL,
  cnLevel = "all",
  cnTable = NULL,
  isTCGA = FALSE,
  vc_nonSyn = NULL,
  verbose = TRUE
)
```

Arguments

maf	tab delimited MAF file. File can also be gz compressed. Required. Alternatively, you can also provide already read MAF file as a dataframe.
clinicalData	Clinical data associated with each sample/Tumor_Sample_Barcode in MAF. Could be a text file or a data.frame. Default NULL.
removeDuplicatedVariants	removes repeated variants in a particular sample, mapped to multiple transcripts of same Gene. See Description. Default TRUE.
useAll	logical. Whether to use all variants irrespective of values in Mutation_Status. Defaults to TRUE. If FALSE, only uses with values Somatic.
gisticAllLesionsFile	All Lesions file generated by gistic. e.g; all_lesions.conf_XX.txt, where XX is the confidence level. Default NULL.
gisticAmpGenesFile	Amplification Genes file generated by gistic. e.g; amp_genes.conf_XX.txt, where XX is the confidence level. Default NULL.
gisticDelGenesFile	Deletion Genes file generated by gistic. e.g; del_genes.conf_XX.txt, where XX is the confidence level. Default NULL.
gisticScoresFile	scores.gistic file generated by gistic. Default NULL
cnLevel	level of CN changes to use. Can be 'all', 'deep' or 'shallow'. Default uses all i.e, genes with both 'shallow' or 'deep' CN changes

cnTable	Custom copynumber data if gistic results are not available. Input file or a data.frame should contain three columns in aforementioned order with gene name, Sample name and copy number status (either 'Amp' or 'Del'). Default NULL.
isTCGA	Is input MAF file from TCGA source. If TRUE uses only first 12 characters from Tumor_Sample_Barcode.
vc_nonSyn	NULL. Provide manual list of variant classifications to be considered as non-synonymous. Rest will be considered as silent variants. Default uses Variant Classifications with High/Moderate variant consequences. http://asia.ensembl.org/Help/Glossary?id= "Frame_Shift_Del", "Frame_Shift_Ins", "Splice_Site", "Translation_Start_Site", "Nonsense_Mutation", "Nonstop_Mutation", "In_Frame_Del", "In_Frame_Ins", "Missense_Mutation"
verbose	TRUE logical. Default to be talkative and prints summary.

Details

This function takes MAF file as input and summarizes them. If copy number data is available, e.g from GISTIC, it can be provided too via arguments `gisticAllLesionsFile`, `gisticAmpGenesFile`, and `gisticDelGenesFile`. Copy number data can also be provided as a custom table containing Gene name, Sample name and Copy Number status.

Note that if input MAF file contains multiple affected transcripts of a variant, this function by default removes them as duplicates, while keeping single unique entry per variant per sample. If you wish to keep all of them, set `removeDuplicatedVariants` to FALSE.

FLAGS - If you get a note on possible FLAGS while reading MAF, it means some of the top mutated genes are fishy. These genes are often non-pathogenic and passengers, but are frequently mutated in most of the public exome studies. Examples of such genes include TTN, MUC16, etc. This note can be ignored without any harm, it's only generated as to make user aware of such genes. See references for details on FLAGS.

Value

An object of class MAF.

References

Shyr C, Tarailo-Graovac M, Gottlieb M, Lee JJ, van Karnebeek C, Wasserman WW. FLAGS, frequently mutated genes in public exomes. *BMC Med Genomics* 2014; 7: 64.

See Also

[plotmafSummary](#) [write.mafSummary](#)

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "mafTools")
laml <- read.maf(maf = laml.maf)
```

readGistic	<i>Read and summarize gistic output.</i>
------------	--

Description

A little function to summarize gistic output files. Summarized output is returned as a list of tables.

Usage

```
readGistic(
  gisticAllLesionsFile = NULL,
  gisticAmpGenesFile = NULL,
  gisticDelGenesFile = NULL,
  gisticScoresFile = NULL,
  cnLevel = "all",
  isTCGA = FALSE,
  verbose = TRUE
)
```

Arguments

gisticAllLesionsFile	All Lesions file generated by gistic. e.g; all_lesions.conf_XX.txt, where XX is the confidence level. Required. Default NULL.
gisticAmpGenesFile	Amplification Genes file generated by gistic. e.g; amp_genes.conf_XX.txt, where XX is the confidence level. Default NULL.
gisticDelGenesFile	Deletion Genes file generated by gistic. e.g; del_genes.conf_XX.txt, where XX is the confidence level. Default NULL.
gisticScoresFile	scores.gistic file generated by gistic.
cnLevel	level of CN changes to use. Can be 'all', 'deep' or 'shallow'. Default uses all i.e, genes with both 'shallow' or 'deep' CN changes
isTCGA	Is the data from TCGA. Default FALSE.
verbose	Default TRUE

Details

Requires output files generated from GISTIC. Gistic documentation can be found here <ftp://ftp.broadinstitute.org/pub/GIS>

Value

A list of summarized data.

Examples

```
all.lesions <- system.file("extdata", "all_lesions.conf_99.txt", package = "maftools")
amp.genes <- system.file("extdata", "amp_genes.conf_99.txt", package = "maftools")
del.genes <- system.file("extdata", "del_genes.conf_99.txt", package = "maftools")
scores.gistic <- system.file("extdata", "scores.gistic", package = "maftools")
laml.gistic = readGistic(gisticAllLesionsFile = all.lesions, gisticAmpGenesFile = amp.genes, gisticDelGenesFi
```

signatureEnrichment	<i>Performs sample stratification based on signature contribution and enrichment analysis.</i>
---------------------	--

Description

Performs k-means clustering to assign signature to samples and performs enrichment analysis.

Usage

```
signatureEnrichment(maf, sig_res, minMut = 5, useCNV = FALSE, fn = NULL)
```

Arguments

maf	an MAF object used for signature analysis.
sig_res	Signature results from extractSignatures
minMut	Consider only genes with minimum this number of samples mutated. Default 5.
useCNV	whether to include copy number events. Only applicable when MAF is read along with copy number data. Default TRUE if available.
fn	basename for output file. Default NULL.

Value

result list containing p-values

See Also

[plotEnrichmentResults](#)

somaticInteractions	<i>Exact tests to detect mutually exclusive, co-occurring and altered genes.</i>
---------------------	--

Description

Performs Pair-wise Fisher's Exact test to detect mutually exclusive or co-occurring events.

Usage

```
somaticInteractions(  
  maf,  
  top = 25,  
  genes = NULL,  
  pvalue = c(0.05, 0.01),  
  returnAll = TRUE,  
  geneOrder = NULL,  
  fontSize = 0.8,  
  showSigSymbols = TRUE,  
  showCounts = FALSE,
```

```

countStats = "sig",
countType = "all",
countsFontSize = 0.8,
countsFontColor = "black",
colPal = "BrBG"
)

```

Arguments

maf	an MAF object generated by <code>read.maf</code>
top	check for interactions among top 'n' number of genes. Defaults to top 25. genes
genes	List of genes among which interactions should be tested. If not provided, test will be performed between top 25 genes.
pvalue	Default c(0.05, 0.01) p-value threshold. You can provide two values for upper and lower threshold.
returnAll	If TRUE returns test statistics for all pair of tested genes. Default FALSE, returns for only genes below pvalue threshold.
geneOrder	Plot the results in given order. Default NULL.
fontSize	cex for gene names. Default 0.8
showSigSymbols	Default TRUE. Highlight significant pairs
showCounts	Default TRUE. Include number of events in the plot
countStats	Default 'all'. Can be 'all' or 'sig'
countType	Default 'cooccur'. Can be 'all', 'cooccur', 'mutexcl'
countsFontSize	Default 0.8
countsFontColor	Default 'black'
colPal	colPalBrewer palettes. See <code>RColorBrewer::display.brewer.all()</code> for details

Details

This function and plotting is inspired from genetic interaction analysis performed in the published study combining gene expression and mutation data in MDS. See reference for details.

Value

list of data.tables

References

Gerstung M, Pellagatti A, Malcovati L, et al. Combining gene mutation with gene expression data improves outcome prediction in myelodysplastic syndromes. *Nature Communications*. 2015;6:5901. doi:10.1038/ncomms6901.

Examples

```

lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "maftools")
lam1 <- read.maf(maf = lam1.maf)
somaticInteractions(maf = lam1, top = 5)

```

subsetMaf	<i>Subset MAF objects</i>
-----------	---------------------------

Description

Subsets MAF based on given conditions.

Usage

```
subsetMaf(  
  maf,  
  tsb = NULL,  
  genes = NULL,  
  query = NULL,  
  clinQuery = NULL,  
  fields = NULL,  
  mafObj = TRUE,  
  includeSyn = TRUE,  
  isTCGA = FALSE,  
  dropLevels = TRUE,  
  restrictTo = "all"  
)
```

Arguments

maf	an MAF object generated by read.maf
tsb	subset by these samples (Tumor Sample Barcodes)
genes	subset by these genes
query	query string. e.g. "Variant_Classification == 'Missense_Mutation'" returns only Missense variants.
clinQuery	query by clinical variable.
fields	include only these fields along with necessary fields in the output
mafObj	returns output as MAF class MAF-class . Default TRUE
includeSyn	Default TRUE, only applicable when mafObj = FALSE. If mafObj = TRUE, synonymous variants will be stored in a separate slot of MAF object.
isTCGA	Is input MAF file from TCGA source.
dropLevels	Default TRUE.
restrictTo	restrict subset operations to these. Can be 'all', 'cnv', or 'mutations'. Default 'all'. If 'cnv' or 'mutations', subset operations will only be applied on copy-number or mutation data respectively, while retaining other parts as is.

Value

subset table or an object of class [MAF-class](#)

See Also

[getFields](#)

Examples

```

laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml <- read.maf(maf = laml.maf)
##Select all Splice_Site mutations from DNMT3A and NPM1
subsetMaf(maf = laml, genes = c('DNMT3A', 'NPM1'),
query = "Variant_Classification == 'Splice_Site'")
##Select all variants with VAF above 30%
subsetMaf(maf = laml, query = "i_TumorVAF_WU > 30")
##Extract data for samples 'TCGA.AB.3009' and 'TCGA.AB.2933' but only include vaf filed.
subsetMaf(maf = laml, tsb = c('TCGA-AB-3009', 'TCGA-AB-2933'), fields = 'i_TumorVAF_WU')

```

survGroup

*Predict genesets associated with survival***Description**

Predict genesets associated with survival

Usage

```

survGroup(
  maf,
  top = 20,
  genes = NULL,
  geneSetSize = 2,
  minSamples = 5,
  clinicalData = NULL,
  time = "Time",
  Status = "Status",
  verbose = TRUE
)

```

Arguments

maf	an MAF object generated by read.maf
top	If genes is NULL by default used top 20 genes
genes	Manual set of genes
geneSetSize	Default 2
minSamples	minimum number of samples to be mutated to be considered for analysis. Default 5
clinicalData	dataframe containing events and time to events. Default looks for clinical data in annotation slot of MAF .
time	column name containing time in clinicalData
Status	column name containing status of patients in clinicalData. must be logical or numeric. e.g, TRUE or FALSE, 1 or 0.
verbose	Default TRUE

Examples

```
laml.maf <- system.file("extdata", "tcga_laml.maf.gz", package = "maftools")
laml.clin <- system.file("extdata", "tcga_laml_annot.tsv", package = "maftools")
laml <- read.maf(maf = laml.maf, clinicalData = laml.clin)
survGroup(maf = laml, top = 20, geneSetSize = 1, time = "days_to_last_followup", Status = "Overall_Survival_Sta
```

tcgaCompare

*Compare mutation load against TCGA cohorts***Description**

Compares mutation load in input MAF against all of 33 TCGA cohorts derived from MC3 project.

Usage

```
tcgaCompare(
  maf,
  capture_size = NULL,
  tcga_capture_size = 50,
  cohortName = NULL,
  tcga_cohorts = NULL,
  primarySite = FALSE,
  col = c("gray70", "black"),
  bg_col = c("#EDF8B1", "#2C7FB8"),
  medianCol = "red",
  logscale = TRUE,
  rm_hyper = FALSE
)
```

Arguments

maf	MAF object(s) generated by read.maf
capture_size	capture size for input MAF in MBs. Default NULL. If provided plot will be scaled to mutations per mb. TCGA capture size is assumed to be 50mb.
tcga_capture_size	capture size for TCGA cohort in MB. Default 50
cohortName	name for the input MAF cohort. Default "Input"
tcga_cohorts	restrict tcga data to these cohorts.
primarySite	If TRUE uses primary site of cancer as labels instead of TCGA project IDs. Default FALSE.
col	color vector for length 2 TCGA cohorts and input MAF cohort. Default gray70 and black.
bg_col	background color. Default '#EDF8B1', '#2C7FB8'
medianCol	color for median line. Default red.
logscale	Default TRUE
rm_hyper	Remove hyper mutated samples (outliers)? Default FALSE

Value

data.table with median mutations per cohort

Examples

```
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "mafTools")
lam1 <- read.maf(maf = lam1.maf)
tcgaCompare(maf = lam1, cohortName = "AML")
```

titv

Classifies SNPs into transitions and transversions

Description

takes output generated by read.maf and classifies Single Nucleotide Variants into Transitions and Transversions.

Usage

```
titv(maf, useSyn = FALSE, plot = TRUE, file = NULL)
```

Arguments

maf	an MAF object generated by read.maf
useSyn	Logical. Whether to include synonymous variants in analysis. Defaults to FALSE.
plot	plots a titv fractions. default TRUE.
file	basename for output file name. If given writes summaries to output file. Default NULL.

Value

list of data.frames with Transitions and Transversions summary.

See Also

[plotTiTv](#)

Examples

```
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "mafTools")
lam1 <- read.maf(maf = lam1.maf)
lam1.titv = titv(maf = lam1, useSyn = TRUE)
```

trinucleotideMatrix *Extract single 5' and 3' bases flanking the mutated site for de-novo signature analysis. Also estimates APOBEC enrichment scores.*

Description

Extract single 5' and 3' bases flanking the mutated site for de-novo signature analysis. Also estimates APOBEC enrichment scores.

Usage

```
trinucleotideMatrix(
  maf,
  ref_genome = NULL,
  prefix = NULL,
  add = TRUE,
  ignoreChr = NULL,
  useSyn = TRUE,
  fn = NULL
)
```

Arguments

maf	an MAF object generated by read.maf
ref_genome	BSgenome object or name of the installed BSgenome package. Example: BSgenome.Hsapiens.UCSC. Default NULL, tries to auto-detect from installed genomes.
prefix	Prefix to add or remove from contig names in MAF file.
add	If prefix is used, default is to add prefix to contig names in MAF file. If false prefix will be removed from contig names.
ignoreChr	Chromosomes to ignore from analysis. e.g. chrM
useSyn	Logical. Whether to include synonymous variants in analysis. Defaults to TRUE
fn	If given writes APOBEC results to an output file with basename fn. Default NULL.

Details

Extracts immediate 5' and 3' bases flanking the mutated site and classifies them into 96 substitution classes. Requires BSgenome data packages for sequence extraction.

APOBEC Enrichment: Enrichment score is calculated using the same method described by Roberts et al.

$$E = (n_tcw * background_c) / (n_C * background_tcw)$$

where, n_tcw = number of mutations within T[C>T]W and T[C>G]W context. (W -> A or T)

n_C = number of mutated C and G

background_C and background_tcw motifs are number of C and TCW motifs occurring around +/- 20bp of each mutation.

One-sided Fisher's Exact test is performed to determine the enrichment of APOBEC tcw mutations over background.

Value

list of 2. A matrix of dimension $n \times 96$, where n is the number of samples in the MAF and a table describing APOBEC enrichment per sample.

References

Roberts SA, Lawrence MS, Klimczak LJ, et al. An APOBEC Cytidine Deaminase Mutagenesis Pattern is Widespread in Human Cancers. *Nature genetics*. 2013;45(9):970-976. doi:10.1038/ng.2702.

See Also

[extractSignatures](#) [plotApobecDiff](#)

Examples

```
## Not run:
laml.tnm <- trinucleotideMatrix(maf = laml, ref_genome = 'BSgenome.Hsapiens.UCSC.hg19',
prefix = 'chr', add = TRUE, useSyn = TRUE)

## End(Not run)
```

write.GisticSummary *Writes GISTIC summaries to output tab-delimited text files.*

Description

Writes GISTIC summaries to output tab-delimited text files.

Usage

```
write.GisticSummary(gistic, basename = NULL)
```

Arguments

gistic	an object of class GISTIC generated by readGistic
basename	basename for output file to be written.

Value

None. Writes output as tab delimited text files.

See Also

[readGistic](#)

Examples

```
all.lesions <- system.file("extdata", "all_lesions.conf_99.txt", package = "maftools")
amp.genes <- system.file("extdata", "amp_genes.conf_99.txt", package = "maftools")
del.genes <- system.file("extdata", "del_genes.conf_99.txt", package = "maftools")
scores.gistic <- system.file("extdata", "scores.gistic", package = "maftools")
laml.gistic = readGistic(gisticAllLesionsFile = all.lesions, gisticAmpGenesFile = amp.genes, gisticDelGenesFi
write.GisticSummary(gistic = laml.gistic, basename = 'laml')
```

write.mafSummary	<i>Writes maf summaries to output tab-delimited text files.</i>
------------------	---

Description

Writes maf summaries to output tab-delimited text files.

Usage

```
write.mafSummary(maf, basename = NULL)
```

Arguments

maf	an MAF object generated by read.maf
basename	basename for output file to be written.

Details

Writes MAF and related summaries to output files.

Value

None. Writes output as text files.

See Also

[read.maf](#)

Examples

```
lam1.maf <- system.file("extdata", "tcga_lam1.maf.gz", package = "maftools")
lam1 <- read.maf(maf = lam1.maf)
write.mafSummary(maf = lam1, basename = 'lam1')
```

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