Package 'destiny'

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Type Package

Title Creates diffusion maps

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Description Create and plot diffusion maps.

License GPL

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BugReports https://github.com/theislab/destiny/issues

Encoding UTF-8

Depends R (>= 3.4.0)

Imports methods, graphics, grDevices, utils, stats, Matrix, Rcpp (>= 0.10.3), RcppEigen, RSpectra (>= 0.14-0), irlba, pcaMethods, Biobase, BiocGenerics, SummarizedExperiment, SingleCellExperiment, ggplot2, ggplot.multistats, tidyr, tidyselect, ggthemes, VIM, knn.covertree, proxy, RcppHNSW, smoother, scales, scatterplot3d

LinkingTo Rcpp, RcppEigen, grDevices

SystemRequirements C++11, jupyter nbconvert (see nbconvertR's INSTALL file)

NeedsCompilation yes

Enhances rgl, SingleCellExperiment

Suggests nbconvertR (>= 1.3.2), igraph, testthat, FNN, tidyr

VignetteBuilder nbconvertR

biocViews CellBiology, CellBasedAssays, Clustering, Software, Visualization

Collate 'RcppExports.R' 'aaa.r' 'accessor-generics.r' 'censoring.r' 'colorlegend.r' 'compat.r' 'cube_helix.r' 'dataset-helpers.r' 'destiny-package.r' 's4-unions.r' 'dist-matrix-coerce.r'

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'sigmas.r' 'diffusionmap.r' 'diffusionmap-methods-accession.r'
'diffusionmap-methods.r' 'plothelpers.r'
'diffusionmap-plotting.r' 'dpt-branching.r' 'dpt-helpers.r'
'dpt.r' 'dpt-methods-matrix.r' 'dpt-methods.r' 'utils.r'
'dpt-plotting.r' 'eig_decomp.r' 'expressionset-helpers.r'
'find_dm_k.r' 'gene-relevance.r' 'gene-relevance-methods.r'
'gene-relevance-plotting-differential-map.r'
'gene-relevance-plotting-gr-map.r'
'gene-relevance-plotting-rank.r' 'gene-relevance-plotting.r'
'guo-data.r' 'knn.r' 'l_which.r' 'methods-coercion.r'
'methods-extraction.r' 'methods-update.r' 'predict.r'
'projection-dist.r' 'rankcor.r' 'sigmas-plotting.r'

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R **topics documented:**

Coercion methods   *Coercion methods*

Description

Convert a [DiffusionMap](#) or [DPT](#) object to other classes

Usage

```
## S4 method for signature 'DiffusionMap'
as.data.frame(x, row.names = NULL,
  optional = FALSE, ...)

fortify.DiffusionMap(model, data, ...)

## S4 method for signature 'DPT'
as.data.frame(x, row.names = NULL, optional = FALSE,
  ...)

fortify.DPT(model, data, ...)

## S4 method for signature 'DPT'
as.matrix(x, ...)
```

Arguments

x, model	A [DiffusionMap](#) or [DPT](#) object
row.names	NULL or a character vector giving the row names for the data frame. Missing values are not allowed.
optional	logical. If TRUE, setting row names and converting column names (to syntactic names: see make.names) is optional.
...	Passed to [as.data.frame](#)
data	ignored

Details

[fortify](#) is a ggplot2 generic allowing a diffusion map to be used as data parameter in [ggplot](#) or [qplot](#).

Value

An object of the desired class

See Also

DiffusionMap accession methods, Extraction methods, DiffusionMap methods for more

Examples

```
library(Biobase)
data(guo)
dm <- DiffusionMap(guo)
classes <- vapply(as.data.frame(dm), class, character(1L))
stopifnot(all(classes[paste0('DC', 1:20)] == 'numeric'))
stopifnot(all(classes[featureNames(guo) ] == 'numeric'))
stopifnot(all(classes[   varLabels(guo) ] == c('factor', 'integer')))
```

---

colorlegend   *Color legend*

Description

Creates a color legend for a vector used to color a plot. It will use the current palette() or the specified pal as reference.

Usage

```
colorlegend(col, pal = palette(), log = FALSE, posx = c(0.9, 0.93),
  posy = c(0.05, 0.9), main = NULL, cex_main = par("cex.sub"),
  cex_axis = par("cex.axis"), col_main = par("col.sub"),
  col_lab = par("col.lab"), steps = 5, steps_color = 100,
  digit = 2, left = FALSE, ..., cex.main = NULL, cex.axis = NULL,
  col.main = NULL, col.lab = NULL)
```

Arguments

col	Vector of factor, integer, or double used to determine the ticks.
pal	If col is double, pal is used as a continuous palette, else as categorical one
log	Use logarithmic scale?
posx	Left and right borders of the color bar relative to plot area (Vector of length 2; 0-1)
posy	Bottom and top borders of color bar relative to plot area (Vector of length 2; 0-1)
main	Legend title
cex_main	Size of legend title font (default: subtitle font size par('cex.sub'))
cex_axis	Size of ticks/category labels (default: axis font size par('cex.axis'))
col_main	Color of legend title (default: subtitle color par('col.sub'))
col_lab	Color of tick or category labels (default: axis color par('col.lab'))
steps	Number of labels in case of a continuous axis. If 0 or FALSE, draw no ticks

steps_color	Number of gradient samples in case of continuous axis
digit	Number of digits for continuous axis labels
left	logical. If TRUE, invert posx
...	Additional parameters for the text call used for labels
cex.main, cex.axis, col.main, col.lab	
	For compatibility with par

Details

When passed a factor or integer vector, it will create a discrete legend, whereas a double vector will result in a continuous bar.

Value

This function is called for the side effect of adding a colorbar to a plot and returns nothing/NULL.

Examples

```
color_data <- 1:6
par(mar = par('mar') + c(0, 0, 0, 3))
plot(sample(6), col = color_data)
colorlegend(color_data)
```

---

cube_helix   *Sequential color palette using the cube helix system*

Description

Creates a perceptually monotonously decreasing (or increasing) lightness color palette with different tones. This was necessary in pre-viridis times, by now you can probably just use hcl.colors

Usage

```
cube_helix(n = 6, start = 0, r = 0.4, hue = 0.8, gamma = 1,
  light = 0.85, dark = 0.15, reverse = FALSE)

scale_colour_cube_helix(..., start = 0, r = 0.4, hue = 0.8,
  gamma = 1, light = 0.85, dark = 0.15, reverse = FALSE,
  discrete = TRUE, guide = if (discrete) "legend" else "colourbar")

scale_color_cube_helix(..., start = 0, r = 0.4, hue = 0.8,
  gamma = 1, light = 0.85, dark = 0.15, reverse = FALSE,
  discrete = TRUE, guide = if (discrete) "legend" else "colourbar")

scale_fill_cube_helix(..., start = 0, r = 0.4, hue = 0.8,
  gamma = 1, light = 0.85, dark = 0.15, reverse = FALSE,
  discrete = TRUE, guide = if (discrete) "legend" else "colourbar")
```

Arguments

n	Number of colors to return (default: 6)
start	Hue to start helix at (start $\in [0, 3]$, default: 0)
r	Number of rotations of the helix. Can be negative. (default: 0.4)
hue	Saturation. 0 means greyscale, 1 fully saturated colors (default: 0.8)
gamma	Emphasize darker (gamma < 1) or lighter (gamma > 1) colors (default: 1)
light	Lightest lightness (default: 0.85)
dark	Darkest lightness (default: 0.15)
reverse	logical. If TRUE, reverse lightness (default: FALSE)
...	parameters passed to `discrete_scale` or `continuous_scale`
discrete	If TRUE, return a discrete scale, if FALSE a continuous one (default: TRUE)
guide	Type of scale guide to use. See `guides`

Value

A `character` vector of hex colors with length n

Examples

```
palette(cube_helix())
image(matrix(1:6), col = 1:6, pch = 19, axes = FALSE)

cr <- scales::colour_ramp(cube_helix(12, r = 3))
r <- runif(100)
plot(1:100, r, col = cr(r), type = 'b', pch = 20)
```

---

destiny   *Create and plot diffusion maps*

Description

The main function is `DiffusionMap`, which returns an object you can `plot` (`plot.DiffusionMap` is then called).

Examples

```
demo(destiny, ask = FALSE)
```

destiny generics	*destiny generics*

Description

destiny provides several generic methods and implements them for the [DiffusionMap](#) and [Sigmas](#) classes.

Usage

```
eigenvalues(object)

eigenvalues(object) <- value

eigenvectors(object)

eigenvectors(object) <- value

sigmas(object)

sigmas(object) <- value

dataset(object)

dataset(object) <- value

distance(object)

distance(object) <- value

optimal_sigma(object)
```

Arguments

object	Object from which to extract or to which to assign a value
value	Value to assign within an object

Value

eigenvalues retrieves the numeric eigenvalues

eigenvectors retrieves the eigenvectors matrix

sigmas retrieves the [Sigmas](#) from an object utilizing it as kernel width

dataset retrieves the data the object was created from

distance retrieves the distance metric used to create the object, e.g. euclidean

optimal_sigma retrieves the numeric value of the optimal sigma or local sigmas

See Also

[DiffusionMap methods](#) and [Sigmas](#) class for implementations

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
eigenvalues(dm)
eigenvectors(dm)
sigmas(dm)
optimal_sigma(dm)
dataset(dm)
distance(dm)
```

```
DiffusionMap accession methods
```
DiffusionMap accession methods

Description

Get and set eigenvalues, eigenvectors, and sigma(s) of a DiffusionMap object.

Usage

```
## S4 method for signature 'DiffusionMap'
eigenvalues(object)

## S4 replacement method for signature 'DiffusionMap'
eigenvalues(object) <- value

## S4 method for signature 'DiffusionMap'
eigenvectors(object)

## S4 replacement method for signature 'DiffusionMap'
eigenvectors(object) <- value

## S4 method for signature 'DiffusionMap'
sigmas(object)

## S4 replacement method for signature 'DiffusionMap'
sigmas(object) <- value

## S4 method for signature 'DiffusionMap'
dataset(object)

## S4 replacement method for signature 'DiffusionMap'
dataset(object) <- value

## S4 method for signature 'DiffusionMap'
distance(object)

## S4 replacement method for signature 'DiffusionMap'
distance(object) <- value
```

```
## S4 method for signature 'DiffusionMap'
optimal_sigma(object)
```

Arguments

object   A DiffusionMap

value   Vector of eigenvalues or matrix of eigenvectors to get/set

Value

The assigned or retrieved value

See Also

Extraction methods, DiffusionMap methods, Coercion methods for more

Examples

```
data(guo)
dm <- DiffusionMap(guo)
eigenvalues(dm)
eigenvectors(dm)
sigmas(dm)
dataset(dm)
optimal_sigma(dm)
```

DiffusionMap methods   *DiffusionMap methods*

Description

Methods for external operations on diffusion maps

Usage

```
## S4 method for signature 'DiffusionMap'
print(x)

## S4 method for signature 'DiffusionMap'
show(object)
```

Arguments

x, object   A DiffusionMap

Value

The DiffusionMap object (print), or NULL (show), invisibly

See Also

DiffusionMap accession methods, Extraction methods, Coercion methods for more

Examples

```
data(guo)
dm <- DiffusionMap(guo)
print(dm)
show(dm)
```

DiffusionMap-class   *Create a diffusion map of cells*

Description

The provided data can be a double [matrix](#) of expression data or a [data.frame](#) with all non-integer (double) columns being treated as expression data features (and the others ignored), an [Expression-Set](#), or a [SingleCellExperiment](#).

Usage

```
DiffusionMap(data = stopifnot_distmatrix(distance), sigma = "local",
  k = find_dm_k(dataset_n_observations(data, distance) - 1L),
  n_eigs = min(20L, dataset_n_observations(data, distance) - 2L),
  density_norm = TRUE, ..., distance = c("euclidean", "cosine",
  "rankcor", "l2"), n_pcs = NULL, n_local = seq(to = min(k, 7L),
  length.out = min(k, 3L)), rotate = FALSE, censor_val = NULL,
  censor_range = NULL, missing_range = NULL, vars = NULL,
  knn_params = list(), verbose = !is.null(censor_range),
  suppress_dpt = FALSE)
```

Arguments

data	Expression data to be analyzed and covariates. Provide vars to select specific columns other than the default: all double value columns. If distance is a distance matrix, data has to be a [data.frame](#) with covariates only.
sigma	Diffusion scale parameter of the Gaussian kernel. One of 'local', 'global', a ([numeric](#)) global sigma or a [Sigmas](#) object. When choosing 'global', a global sigma will be calculated using [find_sigmas](#). (Optional. default: 'local') A larger sigma might be necessary if the eigenvalues can not be found because of a singularity in the matrix
k	Number of nearest neighbors to consider (default: a guess betweeen 100 and $n - 1$. See [find_dm_k](#)).
n_eigs	Number of eigenvectors/values to return (default: 20)
density_norm	logical. If TRUE, use density normalisation
...	Unused. All parameters to the right of the ... have to be specified by name (e.g. DiffusionMap(data, distance = 'cosine'))
distance	Distance measurement method applied to data or a distance matrix/[dist](#). For the allowed values, see [find_knn](#). If this is a [sparseMatrix](#), zeros are interpreted as "not a close neighbors", which allows the use of kNN-sparsified matrices (see the return value of [find_knn](#).

n_pcs	Number of principal components to compute to base calculations on. Using e.g. 50 DCs results in more regular looking diffusion maps. The default NULL will not compute principal components, but use reducedDims(data,'pca') if present. Set to NA to suppress using PCs.
n_local	If sigma == 'local', the n_localth nearest neighbor(s) determine(s) the local sigma
rotate	logical. If TRUE, rotate the eigenvalues to get a slimmer diffusion map
censor_val	Value regarded as uncertain. Either a single value or one for every dimension (Optional, default: censor_val)
censor_range	Uncertainity range for censoring (Optional, default: none). A length-2-vector of certainty range start and end. TODO: also allow $2 \times G$ matrix
missing_range	Whole data range for missing value model. Has to be specified if NAs are in the data
vars	Variables (columns) of the data to use. Specifying NULL will select all columns (default: All floating point value columns)
knn_params	Parameters passed to `find_knn`
verbose	Show a progressbar and other progress information (default: do it if censoring is enabled)
suppress_dpt	Specify TRUE to skip calculation of necessary (but spacious) information for `DPT` in the returned object (default: FALSE)

Value

A DiffusionMap object:

Slots

eigenvalues Eigenvalues ranking the eigenvectors

eigenvectors Eigenvectors mapping the datapoints to n_eigs dimensions

sigmas Sigmas object with either information about the find_sigmas heuristic run or just local or optimal_sigma.

data_env Environment referencing the data used to create the diffusion map

eigenvec0 First (constant) eigenvector not included as diffusion component.

transitions Transition probabilities. Can be NULL

d Density vector of transition probability matrix

d_norm Density vector of normalized transition probability matrix

k The k parameter for kNN

n_pcs Number of principal components used in kNN computation (NA if raw data was used)

n_local The n_localth nearest neighbor(s) is/are used to determine local kernel density

density_norm Was density normalization used?

rotate Were the eigenvectors rotated?

distance Distance measurement method used

censor_val Censoring value

censor_range Censoring range

missing_range Whole data range for missing value model

vars Vars parameter used to extract the part of the data used for diffusion map creation

knn_params Parameters passed to `find_knn`

See Also

DiffusionMap methods to get and set the slots. find_sigmas to pre-calculate a fitting global sigma parameter

Examples

```
data(guo)
DiffusionMap(guo)
DiffusionMap(guo, 13, censor_val = 15, censor_range = c(15, 40), verbose = TRUE)

covars <- data.frame(covar1 = letters[1:100])
dists <- dist(matrix(rnorm(100*10), 100))
DiffusionMap(covars, distance = dists)
```

---

dm_predict   *Predict new data points using an existing DiffusionMap. The resulting*
                              matrix can be used in the plot method for the DiffusionMap

---

Description

Predict new data points using an existing DiffusionMap. The resulting matrix can be used in the plot method for the DiffusionMap

Usage

```
dm_predict(dm, new_data, ..., verbose = FALSE)
```

Arguments

dm	A DiffusionMap object.
new_data	New data points to project into the diffusion map. Can be a matrix, data.frame, ExpressionSet, or SingleCellExperiment.
...	Passed to proxy::dist(new_data, data, dm@distance, ...).
verbose	Show progress messages?

Value

A $nrow(new\_data) \times ncol(eigenvectors(dif))$ matrix of projected diffusion components for the new data.

Examples

```
data(guo)
g1 <- guo[, guo$num_cells != 32L]
g2 <- guo[, guo$num_cells == 32L]
dm <- DiffusionMap(g1)
dc2 <- dm_predict(dm, g2)
plot(dm, new_dcs = dc2)
```

DPT matrix methods   *DPT Matrix methods*

Description

Treat DPT object as a matrix of cell-by-cell DPT distances.

Usage

```
## S4 method for signature 'DPT,index,index,logicalOrMissing'
x[i, j, ..., drop = TRUE]

## S4 method for signature 'DPT,index,missing,logicalOrMissing'
x[i, j, ...,
  drop = TRUE]

## S4 method for signature 'DPT,missing,index,logicalOrMissing'
x[i, j, ...,
  drop = TRUE]

## S4 method for signature 'DPT,missing,missing,logicalOrMissing'
x[i, j, ...,
  drop = TRUE]

## S4 method for signature 'DPT,index,index'
x[[i, j, ...]]

## S4 method for signature 'DPT'
nrow(x)

## S4 method for signature 'DPT'
ncol(x)

## S4 method for signature 'DPT'
dim(x)
```

Arguments

x	DPT object.
i, j	Numeric or logical index.
...	ignored
drop	If TRUE, coerce result to a vector if it would otherwise have 1 %in% dim(result).

See Also

as.matrix.DPT

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
dpt <- DPT(dm)
set.seed(1)
plot(dpt[random_root(dpt), ], Biobase::exprs(guo_norm)['DppaI', ])
```

DPT methods   *DPT methods*

Description

Methods for the DPT class. `branch_divide` subdivides branches for plotting (see the examples).

Usage

```
branch_divide(dpt, divide = integer(0L))

tips(dpt)

## S4 method for signature 'DPT'
dataset(object)

## S4 replacement method for signature 'DPT'
dataset(object) <- value
```

Arguments

`dpt, object`	DPT object
`divide`	Vector of branch numbers to use for division
`value`	Value of slot to set

Value

`branch_divide` and `dataset<-` return the changed object, `dataset` the extracted data, and `tips` the tip indices.

See Also

plot.DPT uses `branch_divide` for its `divide` argument.

Examples

```
data(guo_norm)
dpt <- DPT(DiffusionMap(guo_norm))
dpt_9_branches <- branch_divide(dpt, 1:3)
plot(dpt_9_branches, col_by = 'branch')
```

DPT-class   *Diffusion Pseudo Time*

Description

Create pseudotime ordering and assigns cell to one of three branches

Usage

```
DPT(dm, tips = random_root(dm), ..., w_width = 0.1)
```

Arguments

dm	A [DiffusionMap](#) object. Its transition probabilities will be used to calculate the DPT
tips	The cell index/indices from which to calculate the DPT(s) (integer of length 1-3)
...	Unused. All parameters to the right of the ... have to be specified by name (e.g. DPT(dm,w_width = 0.2))
w_width	Window width to use for deciding the branch cutoff

Details

Treat it as a matrix of pseudotime by subsetting ([dim nrow ncol as.matrix), and as a list of pseudodime, and expression vectors ($ [[names as.data.frame).

Value

A DPT object:

Slots

branch [matrix](#) (of [integer](#)) recursive branch labels for each cell (row); NA for undeceided. Use [branch_divide](#) to modify this.

tips [matrix](#) (of [logical](#)) indicating if a cell (row) is a tip of the corresponding banch level (col)

dm [DiffusionMap](#) used to create this DPT object

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
dpt <- DPT(dm)
str(dpt)
```

---

eig_decomp   *Fast eigen decomposition using* eigs

Description

By default uses a random initialization vector that you can make deterministic using set.seed or override by specifying opts = list(initvec = ...).

Usage

```
eig_decomp(M, n_eigs, sym, ..., opts = list())
```

Arguments

M	A matrix (e.g. from the Matrix package) or a function (see eigs).
n_eigs	Number of eigenvectors to return.
sym	defunct and ignored.
...	Passed to eigs.
opts	Passed to eigs.

Value

see eigs.

Examples

```
eig_decomp(cbind(c(1,0,-1), c(0,1,0), c(-1,0,1)), 2)
```

ExpressionSet helper methods
                          Convert object to [ExpressionSet](#) or read it from a file

---

Description

These functions present quick way to create [ExpressionSet](#) objects.

Usage

```
as.ExpressionSet(x, ...)

## S4 method for signature 'data.frame'
as.ExpressionSet(x, annotation_cols = !sapply(x,
  is.double))

read.ExpressionSet(file, header = TRUE, ...)
```

Arguments

`x`	data.frame to convert to an ExpressionSet.
`...`	Additional parameters to read.table
`annotation_cols`	
	The data.frame columns used as annotations. All others are used as expressions. (Logical, character or numerical index array)
`file`	File path to read ASCII data from
`header`	Specifies if the file has a header row.

Details

They work by using all continuous (double) columns as expression data, and all others as observation annotations.

Value

an ExpressionSet object

See Also

read.table on which `read.ExpressionSet` is based, and ExpressionSet.

Examples

```
library(Biobase)
df <- data.frame(Time  = seq_len(3), #integer column
                 Actb  = c(0.05, 0.3, 0.8),
                 Gapdh = c(0.2, 0.03, 0.1))
set <- as.ExpressionSet(df)
rownames(exprs(set)) == c('Actb', 'Gapdh')
phenoData(set)$Time == 1:3
```

Extraction methods *Extraction methods*

Description

Extraction methods

Usage

```
## S4 method for signature 'DiffusionMap'
names(x)

## S4 method for signature 'DPT'
names(x)

## S4 method for signature 'DiffusionMap,character,missing'
x[[i, j, ...]]
```

```
## S4 method for signature 'DPT,character,missing'
x[[i, j, ...]]

## S4 method for signature 'DiffusionMap'
x$name

## S4 method for signature 'DPT'
x$name
```

Arguments

x	[DiffusionMap](#) or [DPT](#) object
i, name	Name of a diffusion component 'DCx', 'DPTx', 'Branch' or column from the data
j	N/A
...	ignored

Value

The names or data row, see respective generics.

See Also

[Extract](#), [names](#) for the generics. [DiffusionMap accession methods](#), [DiffusionMap methods](#), [Coercion methods](#) for more

Examples

```
data(guo)
dm <- DiffusionMap(guo)
dm$DC1        # A diffusion component
dm$Actb       # A gene expression vector
dm$num_cells  # Phenotype metadata

dpt <- DPT(dm)
dm$Branch
dm$DPT1
```

---

find_dm_k   *Find a suitable k*

Description

The k parameter for the k nearest neighbors used in [DiffusionMap](#) should be as big as possible while still being computationally feasible. This function approximates it depending on the size of the dataset n.

Usage

```
find_dm_k(n, min_k = 100L, small = 1000L, big = 10000L)
```

Arguments

n	Number of possible neighbors (nrow(dataset) - 1)
min_k	Minimum number of neighbors. Will be chosen for $n \geq big$
small	Number of neighbors considered small. If/where $n \leq small$, n itself will be returned.
big	Number of neighbors considered big. If/where $n \geq big$, min_k will be returned.

Value

A vector of the same length as n that contains suitable k values for the respective n

Examples

```
curve(find_dm_k(n),     0, 13000, xname = 'n')
curve(find_dm_k(n) / n, 0, 13000, xname = 'n')
```

---

find_knn   *kNN search*

Description

Approximate k nearest neighbor search with flexible distance function.

Usage

```
find_knn(data, k, ..., query = NULL, distance = c("euclidean",
  "cosine", "rankcor", "l2"), method = c("covertree", "hnsw"),
  sym = TRUE, verbose = FALSE)
```

Arguments

data	Data matrix
k	Number of nearest neighbors
...	Parameters passed to hnsw_knn
query	Query matrix. Leave it out to use data as query
distance	Distance metric to use. Allowed measures: Euclidean distance (default), cosine distance ($1 - corr(c_1, c_2)$) or rank correlation distance ($1 - corr(rank(c_1), rank(c_2))$)
method	Method to use. 'hnsw' is tunable with ... but generally less exact than 'covertree' (default: 'covertree')
sym	Return a symmetric matrix (as long as query is NULL)?
verbose	Show a progressbar? (default: FALSE)

Value

A [list](#) with the entries:

index A $nrow(data) \times k$ [integer matrix](#) containing the indices of the k nearest neighbors for each cell.

dist A $nrow(data) \times k$ [double matrix](#) containing the distances to the k nearest neighbors for each cell.

dist_mat A [dgCMatrix](#) if sym == TRUE, else a [dsCMatrix](#) ($nrow(query) \times nrow(data)$). Any zero in the matrix (except for the diagonal) indicates that the cells in the corresponding pair are close neighbors.

---

find_sigmas	*Calculate the average dimensionality for m different gaussian kernel widths (σ).*

Description

The sigma with the maximum value in average dimensionality is close to the ideal one. Increasing step number gets this nearer to the ideal one.

Usage

```
find_sigmas(data, step_size = 0.1, steps = 10L, start = NULL,
  sample_rows = 500L, early_exit = FALSE, ..., censor_val = NULL,
  censor_range = NULL, missing_range = NULL, vars = NULL,
  verbose = TRUE)
```

Arguments

data	Data set with n observations. Can be a [data.frame](#), [matrix](#), [ExpressionSet](#) or [SingleCellExperiment](#).
step_size	Size of log-sigma steps
steps	Number of steps/calculations
start	Initial value to search from. (Optional. default: $\log_{10}(min(dist(data)))$)
sample_rows	Number of random rows to use for sigma estimation or vector of row indices/names to use. In the first case, only used if actually smaller than the number of available rows (Optional. default: 500)
early_exit	logical. If TRUE, return if the first local maximum is found, else keep running
...	Unused. All parameters to the right of the ... have to be specified by name (e.g. find_sigmas(data, verbose = FALSE))
censor_val	Value regarded as uncertain. Either a single value or one for every dimension
censor_range	Uncertainty range for censoring. A length-2-vector of certainty range start and end. TODO: also allow $2 \times G$ matrix
missing_range	Whole data range for missing value model. Has to be specified if NAs are in the data
vars	Variables (columns) of the data to use. Specifying TRUE will select all columns (default: All floating point value columns)
verbose	logical. If TRUE, show a progress bar and plot the output

Value

Object of class Sigmas

See Also

Sigmas, the class returned by this; DiffusionMap, the class this is used for

Examples

```
data(guo)
sigs <- find_sigmas(guo, verbose = TRUE)
DiffusionMap(guo, sigs)
```

---

find_tips   *Find tips in a DiffusionMap object*

Description

Find tips in a DiffusionMap object

Usage

```
find_tips(dm_or_dpt, root = random_root(dm_or_dpt))
```

Arguments

dm_or_dpt	A DiffusionMap or [DPT](DPT) object
root	Root cell index from which to find tips. (default: random)

Value

An integer vector of length 3

Examples

```
data(guo)
dm <- DiffusionMap(guo)
is_tip <- l_which(find_tips(dm), len = ncol(guo))
plot(dm, col = factor(is_tip))
```

Gene Relevance methods

Gene Relevance methods

Description

featureNames <-... can be used to set the gene names used for plotting (e.g. if the data contains hardly readably gene or transcript IDs). dataset gets the expressions used for the gene relevance calculations, and distance the distance measure.

Usage

```
## S4 method for signature 'GeneRelevance'
print(x)

## S4 method for signature 'GeneRelevance'
show(object)

## S4 method for signature 'GeneRelevance'
featureNames(object)


  ## S4 replacement method for signature 'GeneRelevance,characterOrFactor'
featureNames(object) <- value

## S4 method for signature 'GeneRelevance'
dataset(object)

## S4 replacement method for signature 'GeneRelevance'
dataset(object) <- value

## S4 method for signature 'GeneRelevance'
distance(object)

## S4 replacement method for signature 'GeneRelevance'
distance(object) <- value
```

Arguments

x, object	GeneRelevance object
value	A text vector ([character](#) or [factor](#))

Value

dataset, distance, and featureNames return the stored properties. The other methods return a GeneRelevance object (print, ... <-...), or NULL (show), invisibly

See Also

[gene_relevance](#), [Gene Relevance plotting](#)

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
gr <- gene_relevance(dm)
stopifnot(distance(gr) == distance(dm))
featureNames(gr)[[37]] <- 'Id2 (suppresses differentiation)'
# now plot it with the changed gene name(s)
```

GeneRelevance-class   *Gene relevances for entire data set*

Description

The relevance map is cached insided of the `DiffusionMap`.

Usage

```
gene_relevance(coords, exprs, ..., k = 20L, dims = 1:2,
  distance = NULL, smooth = TRUE, remove_outliers = FALSE,
  verbose = FALSE)

## S4 method for signature 'DiffusionMap,missing'
gene_relevance(coords, exprs, ...,
  k = 20L, dims = 1:2, distance = NULL, smooth = TRUE,
  remove_outliers = FALSE, verbose = FALSE)

## S4 method for signature 'matrix,dMatrixOrMatrix'
gene_relevance(coords, exprs, ...,
  pcs = NULL, knn_params = list(), weights = 1, k, dims, distance,
  smooth, remove_outliers, verbose)
```

Arguments

coords	A `DiffusionMap` object or a cells × dims `matrix`.
exprs	An cells × genes `matrix`. Only provide if coords is no `DiffusionMap`.
...	Unused. All parameters to the right of the ... have to be specified by name.
k	Number of nearest neighbors to use
dims	Index into columns of `coord`
distance	Distance measure to use for the nearest neighbor search.
smooth	Smoothing parameters `c(window,alpha)` (see `smth.gaussian`). Alternatively `TRUE` to use the smoother defaults or `FALSE` to skip smoothing,
remove_outliers	
	Remove cells that are only within one other cell's nearest neighbor, as they tend to get large norms.
verbose	If TRUE, log additional info to the console
pcs	A cell × `n_pcs` matrix of principal components to use for the distances.
knn_params	A `list` of parameters for `find_knn`.
weights	Weights for the partial derivatives. A vector of the same length as `dims`.

Value

A GeneRelevance object:

Slots

coords A cells × dims [matrix](#) or [sparseMatrix](#) of coordinates (e.g. diffusion components), reduced to the dimensions passed as dims

exprs A cells × genes matrix of expressions

partials Array of partial derivatives wrt to considered dimensions in reduced space (genes × cells × dimensions)

partials_norm Matrix with norm of aforementioned derivatives. (n_genes × cells)

nn_index Matrix of k nearest neighbor indices. (cells × k)

dims Column index for plotted dimensions. Can [character](#), [numeric](#) or [logical](#)

distance Distance measure used in the nearest neighbor search. See [find_knn](#)

smooth_window Smoothing window used (see [smth.gaussian](#))

smooth_alpha Smoothing kernel width used (see [smth.gaussian](#))

See Also

[Gene Relevance methods](#), [Gene Relevance plotting](#): plot_differential_map/plot_gene_relevance

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
gr <- gene_relevance(dm)

m <- t(Biobase::exprs(guo_norm))
gr_pca <- gene_relevance(prcomp(m)$x, m)
# now plot them!
```

---

guo   *Guo at al. mouse embryonic stem cell qPCR data*

Description

Gene expression data of 48 genes and an annotation column $num_cells containing the cell stage at which the embryos were harvested.

Usage

```
data(guo)
data(guo_norm)
```

Format

An [ExpressionSet](#) with 48 features, 428 observations and 2 [phenoData](#) annotations.

Details

The data is normalized using the mean of two housekeeping genes. The difference between guo and guo_norm is the LoD being set to 10 in the former, making it usable with the `censor_val` parameter of DiffusionMap.

Value

an ExpressionSet with 48 features and 428 observations containing qPCR Ct values and a "num.cells" observation annotation.

Author(s)

Guoji Guo, Mikael Huss, Guo Qing Tong, Chaoyang Wang, Li Li Sun, Neil D. Clarke, Paul Robson
<robsonp@gis.a-star.edu.sg>

References

http://www.sciencedirect.com/science/article/pii/S1534580710001103

---

l_which   *Logical which*

Description

Inverse of which. Converts an array of numeric or character indices to a logical index array. This function is useful if you need to perform logical operation on an index array but are only given numeric indices.

Usage

```
l_which(idx, nms = seq_len(len), len = length(nms), useNames = TRUE)
```

Arguments

idx	Numeric or character indices.
nms	Array of names or a sequence. Required if idx is a character array
len	Length of output array. Alternative to nms if idx is numeric
useNames	Use the names of nms or idx

Details

Either `nms` or `len` has to be specified.

Value

Logical vector of length `len` or the same length as `nms`

Examples

```
all(l_which(2, len = 3L) == c(FALSE, TRUE, FALSE))
all(l_which(c('a', 'c'), letters[1:3]) == c(TRUE, FALSE, TRUE))
```

plot.DiffusionMap   *3D or 2D plot of diffusion map*

Description

If you want to plot the eigenvalues, simply plot(eigenvalues(dm)[start:end],...)

Usage

```
plot.DiffusionMap(x, dims = 1:3, new_dcs = if (!is.null(new_data))
  dm_predict(x, new_data), new_data = NULL, col = NULL,
  col_by = NULL, col_limits = NULL, col_new = "red", pal = NULL,
  pal_new = NULL, ..., ticks = FALSE, axes = TRUE, box = FALSE,
  legend_main = col_by, legend_opts = list(), interactive = FALSE,
  draw_legend = !is.null(col_by) || (length(col) > 1 &&
  !is.character(col)), consec_col = TRUE, col_na = "grey",
  plot_more = function(p, ..., rescale = NULL) p)

## S4 method for signature 'DiffusionMap,numeric'
plot(x, y, ...)

## S4 method for signature 'DiffusionMap,missing'
plot(x, y, ...)
```

Arguments

x	A DiffusionMap
dims, y	Diffusion components (eigenvectors) to plot (default: first three components; 1:3)
new_dcs	An optional matrix also containing the rows specified with y and plotted. (default: no more points)
new_data	A data set in the same format as x that is used to create new_dcs <-dm_predict(dif,new_data)
col	Single color string or vector of discrete or categoric values to be mapped to colors. E.g. a column of the data matrix used for creation of the diffusion map. (default: cluster_louvain if igraph is installed)
col_by	Specify a dataset(x) or phenoData(dataset(x)) column to use as color
col_limits	If col is a continuous (=double) vector, this can be overridden to map the color range differently than from min to max (e.g. specify c(0,1))
col_new	If new_dcs is given, it will take on this color. A vector is also possible. (default: red)
pal	Palette used to map the col vector to colors. (default: use hcl.colors for continuous and palette() for discrete data)
pal_new	Palette used to map the col_new vector to colors. (default: see pal argument)
...	Parameters passed to plot, scatterplot3d, or plot3d (if interactive == TRUE)
ticks	logical. If TRUE, show axis ticks (default: FALSE)
axes	logical. If TRUE, draw plot axes (default: Only if ticks is TRUE)

box	logical. If TRUE, draw plot frame (default: TRUE or the same as axes if specified)
legend_main	Title of legend. (default: nothing unless col_by is given)
legend_opts	Other colorlegend options (default: empty list)
interactive	Use plot3d to plot instead of scatterplot3d?
draw_legend	logical. If TRUE, draw color legend (default: TRUE if col_by is given or col is given and a vector to be mapped)
consec_col	If col or col_by refers to an integer column, with gaps (e.g. c(5,0,0,3)) use the palette color consecutively (e.g. c(3,1,1,2))
col_na	Color for NA in the data. specify NA to hide.
plot_more	Function that will be called while the plot margins are temporarily changed (its p argument is the rgl or scatterplot3d instance or NULL, its rescale argument is NULL, a list(from = c(a,b),to = c(c,d))), or an array of shape $from|to \times dims \times min|max$, i.e. $2 \times length(dims) \times 2$. In case of 2d plotting, it should take and return a ggplot2 object.

Details

If you specify negative numbers as diffusion components (e.g. plot(dm,c(-1,2))), then the corresponding components will be flipped.

Value

The return value of the underlying call is returned, i.e. a scatterplot3d or rgl object.

Examples

```
data(guo)
plot(DiffusionMap(guo))
```

---

plot.DPT	*Plot DPT*

Description

Plots diffusion components from a Diffusion Map and the accompanying Diffusion Pseudo Time (DPT)

Usage

```
plot.DPT(x, root = NULL, paths_to = integer(0L), dcs = 1:2,
  divide = integer(0L), w_width = 0.1, col_by = "dpt",
  col_path = rev(palette()), col_tip = "red", ..., col = NULL,
  legend_main = col_by)

## S4 method for signature 'DPT,numeric'
plot(x, y, ...)

## S4 method for signature 'DPT,missing'
plot(x, y, ...)
```

Arguments

x	A [DPT](#) object.
paths_to	Numeric Branch IDs. Are used as target(s) for the path(s) to draw.
dcs	The dimensions to use from the DiffusionMap
divide	If `col_by = 'branch'`, this specifies which branches to divide. (see [branch_divide](#))
w_width	Window width for smoothing the path (see [smth.gaussian](#))
col_by	Color by 'dpt' (DPT starting at `branches[[1]]`), 'branch', or a veriable of the data.
col_path	Colors for the path or a function creating n colors
col_tip	Color for branch tips
...	Graphical parameters supplied to [plot.DiffusionMap](#)
col	See [plot.DiffusionMap](#). This overrides col_by
legend_main	See [plot.DiffusionMap](#).
y, root	Root branch ID. Will be used as the start of the DPT. (default: lowest branch ID) (If longer than size 1, will be interpreted as c(root,branches))

Value

The return value of the underlying call is returned, i.e. a scatterplot3d or rgl object for 3D plots.

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
dpt <- DPT(dm)
plot(dpt)
plot(dpt, 2L,      col_by = 'branch')
plot(dpt, 1L, 2:3, col_by = 'num_cells')
plot(dpt, col_by = 'DPT3')
```

---

plot.Sigmas   *Plot [Sigmas](#) object*

Description

Plot [Sigmas](#) object

Usage

```
## S4 method for signature 'Sigmas,missing'
plot(x, col = par("fg"),
  col_highlight = "#E41A1C", col_line = "#999999", type = c("b",
  "b"), pch = c(par("pch"), 4L), only_dim = FALSE, ..., xlab = NULL,
  ylab = NULL, main = "")
```

Arguments

x	Sigmas object to plot
col	Vector of bar colors or single color for all bars
col_highlight	Color for highest bar. Overrides col
col_line	Color for the line and its axis
type	Plot type of both lines. Can be a vector of length 2 to specify both separately (default: 'b' aka "both lines and points")
pch	Point identifier for both lines. Can be a vector of length 2 to specify both separately (default: par(pch) and 4 (a '×'))
only_dim	logical. If TRUE, only plot the derivative line
...	Options passed to the call to plot
xlab	X label. NULL to use default
ylab	Either one y label or y labels for both plots. NULL to use both defauts, a NULL in a list of length 2 to use one default.
main	Title of the plot

Value

This method plots a Sigma object to the current device and returns nothing/NULL

Examples

```
data(guo)
sigs <- find_sigmas(guo)
plot(sigs)
```

---

plot_differential_map   *Plot gene relevance or differential map*

Description

plot(gene_relevance, 'Gene') plots the differential map of this/these gene(s), plot(gene_relevance) a relevance map of a selection of genes. Alternatively, you can use plot_differential_map or plot_gene_relevance on a GeneRelevance or DiffusionMap object, or with two matrices.

Usage

```
plot_differential_map(coords, exprs, ..., genes, dims = 1:2,
  pal = hcl.colors, faceter = facet_wrap(~Gene))

## S4 method for signature 'matrix,matrix'
plot_differential_map(coords, exprs, ..., genes,
  dims = 1:2, pal = hcl.colors, faceter = facet_wrap(~Gene))

## S4 method for signature 'DiffusionMap,missing'
plot_differential_map(coords, exprs, ...,
  genes, dims = 1:2, pal = hcl.colors, faceter = facet_wrap(~Gene))
```

```
## S4 method for signature 'GeneRelevance,missing'
plot_differential_map(coords, exprs, ...,
  genes, dims = 1:2, pal = hcl.colors, faceter = facet_wrap(~Gene))

plot_gene_relevance(coords, exprs, ..., iter_smooth = 2L, n_top = 10L,
  genes = NULL, dims = 1:2, pal = palette(), col_na = "grey",
  limit = TRUE)

## S4 method for signature 'matrix,matrix'
plot_gene_relevance(coords, exprs, ...,
  iter_smooth = 2L, n_top = 10L, genes = NULL, dims = 1:2,
  pal = palette(), col_na = "grey", limit = TRUE)

## S4 method for signature 'DiffusionMap,missing'
plot_gene_relevance(coords, exprs, ...,
  iter_smooth = 2L, n_top = 10L, genes = NULL, dims = 1:2,
  pal = palette(), col_na = "grey", limit = TRUE)

## S4 method for signature 'GeneRelevance,missing'
plot_gene_relevance(coords, exprs, ...,
  iter_smooth = 2L, n_top = 10L, genes = NULL, dims = 1:2,
  pal = palette(), col_na = "grey", limit = TRUE)

plot_gene_relevance_rank(coords, exprs, ..., genes, dims = 1:2,
  n_top = 10L, pal = c("#3B99B1", "#F5191C"), bins = 10L,
  faceter = facet_wrap(~Gene))

## S4 method for signature 'matrix,matrix'
plot_gene_relevance_rank(coords, exprs, ...,
  genes, dims = 1:2, n_top = 10L, pal = c("#3B99B1", "#F5191C"),
  bins = 10L, faceter = facet_wrap(~Gene))

## S4 method for signature 'DiffusionMap,missing'
plot_gene_relevance_rank(coords, exprs,
  ..., genes, dims = 1:2, n_top = 10L, pal = c("#3B99B1", "#F5191C"),
  bins = 10L, faceter = facet_wrap(~Gene))

## S4 method for signature 'GeneRelevance,missing'
plot_gene_relevance_rank(coords, exprs,
  ..., genes, dims = 1:2, n_top = 10L, pal = c("#3B99B1", "#F5191C"),
  bins = 10L, faceter = facet_wrap(~Gene))

## S4 method for signature 'GeneRelevance,character'
plot(x, y, ...)

## S4 method for signature 'GeneRelevance,numeric'
plot(x, y, ...)

## S4 method for signature 'GeneRelevance,missing'
plot(x, y, ...)
```

Arguments

coords	A [DiffusionMap](GeneRelevance)/GeneRelevance object or a cells × dims matrix.
exprs	An cells × genes matrix. Only provide if coords is a matrix.
...	Passed to plot_differential_map/plot_gene_relevance.
genes	Genes to base relevance map on (vector of strings). You can also pass an index into the gene names (vector of numbers or logicals with length > 1). The default NULL means all genes.
dims	Names or indices of dimensions to plot. When not plotting a GeneRelevance object, the relevance for the dimensions 1:max(dims) will be calculated.
pal	Palette. Either A colormap function or a list of colors.
faceter	A ggplot faceter like facet_wrap(~ Gene).
iter_smooth	Number of label smoothing iterations to perform on relevance map. The higher the more homogenous and the less local structure.
n_top	Number the top n genes per cell count towards the score defining which genes to return and plot in the relevance map.
col_na	Color for cells that end up with no most relevant gene.
limit	Limit the amount of displayed gene labels to the amount of available colors in pal?
bins	Number of hexagonal bins for plot_gene_relevance_rank.
x	GeneRelevance object.
y	Gene name(s) or index/indices to create differential map for. (integer or character)

Value

ggplot2 plot, when plotting a relevance map with a list member $ids containing the gene IDs used.

See Also

gene_relevance, Gene Relevance methods

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
gr <- gene_relevance(dm)
plot(gr)          # or plot_gene_relevance(dm)
plot(gr, 'Fgf4')  # or plot_differential_map(dm, 'Fgf4')

guo_norm_mat <- t(Biobase::exprs(guo_norm))
pca <- prcomp(guo_norm_mat)$x
plot_gene_relevance(pca, guo_norm_mat, dims = 2:3)
plot_differential_map(pca, guo_norm_mat, genes = c('Fgf4', 'Nanog'))
```

---

projection_dist   *Projection distance*

Description

Projection distance

Usage

```
projection_dist(dm, new_dcs = NULL, ..., new_data, verbose = FALSE)
```

Arguments

dm	A [DiffusionMap](#) object.
new_dcs	Diffusion component matrix of which to calculate the distance to the data.
...	Passed to [proxy::dist](#) if new_data was passed.
new_data	New data points to project into the diffusion map. Can be a [matrix](#), [data.frame](#), [ExpressionSet](#), or [SingleCellExperiment](#).
verbose	If [TRUE](#), log additional info to the console.

Examples

```
data(guo_norm)
g2_32 <- guo_norm[, guo_norm$num_cells < 64]
g64  <- guo_norm[, guo_norm$num_cells == 64]
dm <- DiffusionMap(g2_32)
d <- projection_dist(dm, new_data = g64)
```

---

random_root   *Find a random root cell index*

Description

Finds a cell that has the maximum DPT distance from a randomly selected one.

Usage

```
random_root(dm_or_dpt)
```

Arguments

dm_or_dpt	A [DiffusionMap](#) or [DPT](#) object

Value

A cell index

Examples

```
data(guo)
dm <- DiffusionMap(guo)
random_root(dm)
```

Sigmas-class   *Sigmas Object*

Description

Holds the information about how the `sigma` parameter for a [DiffusionMap](#) was obtained, and in this way provides a plotting function for the [find_sigmas](#) heuristic. You should not need to create a Sigmas object yourself. Provide `sigma` to [DiffusionMap](#) instead or use [find_sigmas](#).

Usage

```
Sigmas(...)

## S4 method for signature 'Sigmas'
optimal_sigma(object)

## S4 method for signature 'Sigmas'
print(x)

## S4 method for signature 'Sigmas'
show(object)
```

Arguments

object, x	[Sigmas](#) object
...	See "**Slots**" below

Details

A Sigmas object is either created by [find_sigmas](#) or by specifying the `sigma` parameter to [DiffusionMap](#).

In the second case, if the `sigma` parameter is just a number, the resulting Sigmas object has all slots except of `optimal_sigma` set to NULL.

Value

Sigmas creates an object of the same class

optimal_sigma retrieves the numeric value of the optimal sigma or local sigmas

Slots

log_sigmas Vector of length m containing the $\log_{10}$ of the σs

dim_norms Vector of length $m - 1$ containing the average dimensionality $\langle p \rangle$ for the respective kernel widths

optimal_sigma Multiple local sigmas or the mean of the two global σs around the highest $\langle p \rangle$ (c(optimal_idx,optimal_idx+1L))

optimal_idx The index of the highest $\langle p \rangle$.

avrd_norms Vector of length m containing the average dimensionality for the corresponding sigma.

See Also

[find_sigmas](), the function to determine a locally optimal sigma and returning this class

Examples

```
data(guo)
sigs <- find_sigmas(guo, verbose = FALSE)
optimal_sigma(sigs)
print(sigs)
```

---

updateObject methods   *Update old destiny objects to a newer version.*

Description

Handles [DiffusionMap](), [Sigmas](), and [GeneRelevance]().

Usage

```
## S4 method for signature 'DiffusionMap'
updateObject(object, ..., verbose = FALSE)

## S4 method for signature 'Sigmas'
updateObject(object, ..., verbose = FALSE)

## S4 method for signature 'GeneRelevance'
updateObject(object, ..., verbose = FALSE)
```

Arguments

object	An object created with an older destiny release
...	ignored
verbose	tells what is being updated

Value

A [DiffusionMap]() or [Sigmas]() object that is valid when used with the current destiny release

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