

# Package ‘bmass’

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**Title** Bayesian Multivariate Analysis of Summary Statistics

**Description** Multivariate tool for analyzing genome-wide association study results in the form of univariate summary statistics. The goal of 'bmass' is to comprehensively test all possible multivariate models given the phenotypes and datasets provided. Multivariate models are determined by assigning each phenotype to being either Unassociated (U), Directly associated (D) or Indirectly associated (I) with the genetic variant of interest. Test results for each model are presented in the form of Bayes factors, thereby allowing direct comparisons between models. The underlying framework implemented here is based on the modeling developed in "A Unified Framework for Association Analysis with Multiple Related Phenotypes", M. Stephens (2013) <[doi:10.1371/journal.pone.0065245](https://doi.org/10.1371/journal.pone.0065245)>.

**License** GPL (>= 3)

**URL** <https://github.com/mturchin20/bmass>

**BugReports** <https://github.com/mturchin20/bmass/issues>

**Depends** R (>= 3.3.0)

**Imports** utils, stats

**Suggests** testthat, knitr, rmarkdown

**LazyData** true

**NeedsCompilation** no

**RoxygenNote** 6.1.1

**VignetteBuilder** knitr

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## R topics documented:

bmass	2
bmass_SimulatedData1	4
bmass_SimulatedData2	4
bmass_SimulatedSigSNPs	5
bmass_TestData1	5
bmass_TestData2	6
bmass_TestSigSNPs	6
GetMarginalPosteriors	7
GetModelPriorMatrix	8
GetTopModelsPerSNPViaPosteriors	9
GlobalLipids2013.GWASsnps	10
<b>Index</b>	<b>11</b>

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bmass	<i>Bayesian multivariate analysis of summary statistics (bmass)</i>
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## Description

Run `bmass` on a set of phenotypes that each have univariate GWAS statistics on the same set of SNPs

## Usage

```
bmass(DataSources, GWASsnps = NULL,
      SNPMarginalUnivariateThreshold = 1e-06,
      SNPMarginalMultivariateThreshold = 1e-06, GWASThreshFlag = TRUE,
      GWASThreshValue = 5e-08, NminThreshold = 0,
      PrintMergedData = FALSE, PrintProgress = FALSE, ...)
```

## Arguments

<code>DataSources</code>	A string indicating the variable names of the input datafiles and phenotypes. No default value.
<code>GWASsnps</code>	A <code>data.table</code> containing rows of SNPs that were univariate genome-wide significant in the phenotypes being used for analysis; <code>GWASsnps</code> input file should have two columns, one for chromosome and another for basepair position (with column headers of <code>Chr</code> and <code>BP</code> ). No default value.
<code>SNPMarginalUnivariateThreshold</code>	A numerical value indicating the univariate p-value threshold to use when collecting marginally significant SNPs for final <code>bmass</code> analysis. Default is $1e-6$ .

SNPMarginalMultivariateThreshold	A numerical value indicating the basic multivariate p-value threshold to use when collecting marginally significant SNPs for final bmass analysis. Default is 1e-6.
GWASThreshFlag	A logical TRUE/FALSE flag that indicates whether to threshold input GWASsnps list by a univariate GWAS p-value or not (eg the input GWASsnps list contains variants that are significant from discovery + replication data, but the input summary statistics are just from the discovery cohort). Default is TRUE.
GWASThreshValue	A numerical value indicating the univariate p-value threshold to use in conjunction with the GWASThreshFlag. Default is 5e-8.
NminThreshold	A numerical value that indicates a sample size threshold to use where SNPs below which are removed. Default is 0.
PrintMergedData	A logical TRUE/FALSE flag that indicates whether the intermediary 'merged datafile' should be included in the final bmass output; this file combines all the phenotypes for every SNP provided just prior to thresholding for marginally significant SNPs. Default is FALSE.
PrintProgress	A logical TRUE/FALSE flag that indicates whether progress statements should be printed to stderr during the course of running bmass or not. Default is FALSE.
...	Additional optional arguments.

### Value

A list containing model, SNP, and posterior information for both the previously significant univariate SNPs (PreviousSNPs) and the newly significant multivariate SNPs (NewSNPs). For a full breakdown of the bmass output list structure, please see the associated vignettes.

### Other Examples

```
bmass(c("HDL", "LDL", "TG", "TC"), GWASsnps, NminThreshold = 50000) bmass(c("HDL", "LDL", "TG", "TC"),
GWASsnps, GWASThreshValue = 1e-8, NminThreshold = 50000, PrintProgress = TRUE) bmass(c("HDL",
"LDL", "TG", "TC"), GWASsnps, GWASThreshFlag = FALSE, SNPMarginalUnivariateThreshold
= 1e-4, SNPMarginalMultivariateThreshold = 1e-4, PrintMergedData = TRUE) bmassOutput
<- bmass(c("HDL", "LDL", "TG", "TC"), GWASsnps, NminThreshold = 50000)
```

### Examples

```
Phenotypes <- c("bmass_SimulatedData1", "bmass_SimulatedData2")
bmassOutput <- bmass(Phenotypes, bmass_SimulatedSigSNPs)
summary(bmassOutput)
bmassOutput$NewSNPs$SNPs
```

---

bmass\_SimulatedData1 *bmass Simulated Dataset 1*

---

**Description**

A manually created sample dataset for use in Roxygen2 documents and vignettes.

**Format**

A data frame with 11 rows and 9 variables:

**Chr** chromosome

**BP** basepair position

**Marker** rsID# or other identifier

**MAF** Minor Allele Frequency

**A1** reference allele

**A2** alternative allele

**Direction** direction of association effect size, + or -

**pValue** p-Value of GWAS association

**N** sample size

**Source**

Manually created

---

bmass\_SimulatedData2 *bmass Simulated Dataset 2*

---

**Description**

A manually created sample dataset for use in Roxygen2 documents and vignettes.

**Format**

A data frame with 11 rows and 9 variables:

**Chr** chromosome

**BP** basepair position

**Marker** rsID# or other identifier

**MAF** Minor Allele Frequency

**A1** reference allele

**A2** alternative allele

**Direction** direction of association effect size, + or -

**pValue** p-Value of GWAS association

**N** sample size

**Source**

Manually created

---

bmass\_SimulatedSigSNPs

*bmass Simulated GWAS SNPs*

---

**Description**

A manually created list of GWAS significant SNPs to be used in conjunction with 'bmass\_SimulatedData1' and 'bmass\_SimulatedData2'.

**Format**

A data frame with 2 rows and 2 variables:

**Chr** chromosome

**BP** basepair position

**Source**

Manually created

---

bmass\_TestData1

*bmass Test Dataset 1*

---

**Description**

A manually created sample dataset for use in unit tests.

**Format**

A data frame with 11 rows and 9 variables:

**Chr** chromosome

**BP** basepair position

**Marker** rsID# or other identifier

**MAF** Minor Allele Frequency

**A1** reference allele

**A2** alternative allele

**Direction** direction of association effect size, + or -

**pValue** p-Value of GWAS association

**N** sample size

**Source**

Manually created

---

bmass\_TestData2      *bmass Test Dataset 2*

---

**Description**

A manually created sample dataset for use in unit tests.

**Format**

A data frame with 11 rows and 9 variables:

**Chr** chromosome

**BP** basepair position

**Marker** rsID# or other identifier

**MAF** Minor Allele Frequency

**A1** reference allele

**A2** alternative allele

**Direction** direction of association effect size, + or -

**pValue** p-Value of GWAS association

**N** sample size

**Source**

Manually created

---

bmass\_TestSigSNPs      *bmass Test GWAS SNPs*

---

**Description**

A manually created list of GWAS significant SNPs to be used in conjunction with 'bmass\_TestData1' and 'bmass\_TestData2'.

**Format**

A data frame with 2 rows and 2 variables:

**Chr** chromosome

**BP** basepair position

**Source**

Manually created

---

GetMarginalPosteriors *Get Marginal {U,D,I} Posteriors*

---

## Description

Get marginal posteriors for how much every individual phenotype belongs to categories {U,D,I} across each SNP

## Usage

```
GetMarginalPosteriors(DataSources, ListSNPs, Models, LogFile)
```

## Arguments

DataSources	A string indicating the variable names of the input datafiles and phenotypes.
ListSNPs	A list produced from running <a href="#">bmass</a> containing the SNPs of interest to get marginal posteriors for.
Models	A matrix describing the models being explored (default output from running <a href="#">bmass</a> ).
LogFile	A matrix of string outputs for function logging purposes (default output from running <a href="#">bmass</a> ).

## Value

A list containing three matrices of SNPs x Phenotypes marginal posteriors for each category {U,D,I}; this list is appended to the input ListSNPs as a new object, Marginals (the full returned object is a list containing the input ListSNPs and the input LogFile).

## Examples

```
Phenotypes <- c("bmass_SimulatedData1", "bmass_SimulatedData2")
bmassOutput <- bmass(Phenotypes, bmass_SimulatedSigSNPs)
bmassOutput[c("PreviousSNPs", "LogFile")] <-
  GetMarginalPosteriors(Phenotypes, bmassOutput$PreviousSNPs,
    bmassOutput$Models, bmassOutput$LogFile)
bmassOutput$PreviousSNPs$Marginals
```

---

GetModelPriorMatrix     *Get Model Prior Matrix*

---

## Description

Creates a matrix containing the model descriptions and their associated priors.

## Usage

```
GetModelPriorMatrix(DataSources, Models, ModelPriors, LogFile,
  SigmaAlphas = c(0.005, 0.0075, 0.01, 0.015, 0.02, 0.03, 0.04, 0.05,
  0.06, 0.07, 0.08, 0.09, 0.1, 0.15))
```

## Arguments

DataSources	A string indicating the variable names of the input datafiles and phenotypes.
Models	A matrix describing the models being explored (default output from running <a href="#">bmass</a> ).
ModelPriors	A vector containing the priors on each model across each tranche of sigma alpha (default output from running <a href="#">bmass</a> ; length is number of models times number of sigma alphas).
LogFile	A matrix of string outputs for function logging purposes (default output from running <a href="#">bmass</a> ).
SigmaAlphas	A vector containing the different values traversed for this 'effect size controlling' hyperparameter (see "Prior on Sigma_Alpha" in Stephens 2013 PLoS ONE, <a href="https://doi.org/10.1371/journal.pone.0065245">https://doi.org/10.1371/journal.pone.0065245</a> ).

## Value

A matrix containing the original description of each model sort by prior, each model's trained prior, the cumulative prior distribution, and the model's original order position.

## Examples

```
Phenotypes <- c("bmass_SimulatedData1", "bmass_SimulatedData2")
bmassOutput <- bmass(Phenotypes,bmass_SimulatedSigSNPs)
bmassOutput[c("ModelPriorMatrix", "LogFile")] <-
  GetModelPriorMatrix(Phenotypes, bmassOutput$Models,
  bmassOutput$ModelPriors, bmassOutput$LogFile)
head(bmassOutput$ModelPriorMatrix)
```



---

GetTopModelsPerSNPViaPosteriors  
*Get Top Multivariate Models*

---

## Description

Get a summary of the top models per SNP across all multivariate {**U,D,I**} combinations based on posterior probabilities.

## Usage

```
GetTopModelsPerSNPViaPosteriors(DataSources, ListSNPs, ModelPriorMatrix,  
  LogFile)
```

## Arguments

DataSources	A string indicating the variable names of the input datafiles and phenotypes.
ListSNPs	A list produced from running <code>bmass</code> containing the SNPs of interest to get marginal posteriors for.
ModelPriorMatrix	A matrix detailing the models being explored and their associated priors (obtained by running <code>GetModelPriorMatrix</code> )
LogFile	A matrix of string outputs for function logging purposes (default output from running <code>bmass</code> ).

## Value

A matrix containing each model that was a SNP's top model at least once, along with related information; this matrix is appended to the input ListSNPs as a new object, TopModels (the full returned object is a list containing the input ListSNPs and the input LogFile).

## Examples

```
Phenotypes <- c("bmass_SimulatedData1", "bmass_SimulatedData2")  
bmassOutput <- bmass(Phenotypes, bmass_SimulatedSigSNPs)  
bmassOutput[c("ModelPriorMatrix", "LogFile")] <-  
  GetModelPriorMatrix(Phenotypes, bmassOutput$Models,  
    bmassOutput$ModelPriors, bmassOutput$LogFile)  
bmassOutput[c("PreviousSNPs", "LogFile")] <-  
  GetTopModelsPerSNPViaPosteriors(Phenotypes,  
    bmassOutput$PreviousSNPs, bmassOutput$ModelPriorMatrix, bmassOutput$LogFile)  
head(bmassOutput$PreviousSNPs$TopModels)
```

---

GlobalLipids2013.GWASsnps

*GlobalLipids2013 GWAS SNPs*

---

**Description**

A list of the univariate GWAS significant SNPs from the GlobalLipids2013 dataset to be used in the second introductory bmass vignette.

**Format**

A data frame with 157 rows and 2 variables:

**Chr** chromosome

**BP** basepair position

**Source**

Supplementary Tables 2 and 3 from <https://doi.org/10.1038/ng.2797>.

# Index

## \* data

- bmass\_SimulatedData1, [4](#)
- bmass\_SimulatedData2, [4](#)
- bmass\_SimulatedSigSNPs, [5](#)
- bmass\_TestData1, [5](#)
- bmass\_TestData2, [6](#)
- bmass\_TestSigSNPs, [6](#)
- GlobalLipids2013.GWASsnps, [10](#)

bmass, [2](#), [7-9](#)

- bmass\_SimulatedData1, [4](#)
- bmass\_SimulatedData2, [4](#)
- bmass\_SimulatedSigSNPs, [5](#)
- bmass\_TestData1, [5](#)
- bmass\_TestData2, [6](#)
- bmass\_TestSigSNPs, [6](#)

- GetMarginalPosteriors, [7](#)
- GetModelPriorMatrix, [8](#), [9](#)
- GetTopModelsPerSNPViaPosteriors, [9](#)
- GlobalLipids2013.GWASsnps, [10](#)